

## TOLERANCE VALUES AND EFFECTS OF SELECTED ENVIRONMENTAL DETERMINANTS ON DIATOM DISTRIBUTION IN NORTHWEST AND NORTH CENTRAL WASHINGTON, USA

Dean W. Blinn<sup>1</sup>

**ABSTRACT.**—Diatoms were collected at 165 stream, river, wall seep, and pond/lake sites in 9 counties in northwest and north central Washington, including the Olympic Peninsula, over a 5-year period. Four hundred and fifteen species in 77 genera were identified. Species assemblages were compared to altitude, physicochemical factors, aquatic habitats, and land use on west and east sides of the North Cascade Range. Species richness averaged 30.5 per site on the west side and 43 on the east side. Diatom species richness showed significant positive correlations to total nitrogen, total phosphorus, pH, specific conductance, channel embeddedness, and water temperature, and a significant negative correlation to percent canopy cover. A multilevel hierarchical clustering model separated streams, rivers, wall seeps, and ponds/lakes into geographic and land-use regions based on diatom assemblages. A multimetric index (diatom tolerance index [DTI]) was developed to determine environmental tolerance levels for diatom species over a large number and variety of sites. The index performed well in distinguishing between the role of embeddedness, total nitrogen, total phosphorus, and specific conductance on the distribution of diatom species and showed close agreement between land use and diatom distribution. These DTI values provide baseline information for monitoring changes in ecosystem health in drainages throughout Washington landscapes. *Didymosphenia geminata* occurred in 46% of the streams and rivers sampled and ranged from <1% to 27% in relative abundance, but it was not collected in ponds/lakes or wall seeps. *Didymosphenia geminata* density showed a significant positive correlation to suspended sediment and significant negative correlations to altitude and total nitrogen. This invasive alga attained a higher average relative abundance in rivers compared to streams and occurred in 28% of the sites examined west of the Cascade Range compared to 63% of the sites east of the Cascades.

**RESUMEN.**—Se colectaron diatomeas en 165 arroyos, ríos, paredes filtradas y estanques/lagos de 9 condados del noroeste y centro-norte de Washington, incluyendo la Península Olímpica, durante un período de 5 años. Se identificaron cuatrocientas quince especies de 77 géneros. Se compararon los grupos de especies con respecto a la altitud, los factores fisicoquímicos, los hábitats acuáticos y el uso de suelo en los este y oeste de la Cordillera de las Cascadas del Norte (North Cascade Range). La riqueza de especies tuvo un promedio de 30.5 en cada ubicación de la zona oeste y 43 en el lado este. Se obtuvieron correlaciones positivas significativas entre la riqueza de especies y el nitrógeno total, fósforo total, el pH, la conductividad específica, la incrustación en la acequia y la temperatura del agua, y una correlación negativa significativa con el % de cobertura vegetal. Un modelo de agrupación jerárquica multinivel separó los arroyos, los ríos, las paredes filtradas y los estanque/lagos en regiones geográficas y el uso de suelo, basándose en las asociaciones de diatomeas. Se desarrolló un índice multimétrico [Tolerancia Diatomea Index (DTI, por sus siglas en inglés)] para determinar los niveles de tolerancia ambiental en especies de diatomeas a lo largo de un gran número y variedad de ubicaciones. El índice funcionó bien a la hora de distinguir entre el papel jugado por la incrustación, el nitrógeno total, el fósforo total y la conducción específica sobre la distribución de especies de diatomeas, y mostró una estrecha concordancia entre el uso de suelo. Estos valores DTI proporcionan información de referencia para el seguimiento de los cambios en la salud de los ecosistemas en pozos a lo largo de Washington. Se encontró *Didymosphenia geminata* en el 46% de los arroyos y ríos medidos, la abundancia relativa osciló entre <1% al 27%, pero no se colectó en estanques/lagos o paredes filtradas. Se encontró una correlación positiva significativa entre la densidad de *D. geminata* con los sedimentos en suspensión y con la altitud y el nitrógeno total se encontraron correlaciones negativas significativas. Esta alga invasora alcanza una mayor abundancia relativa promedio en ríos en comparación con arroyos, y se encontró en el 28% de las zonas inspeccionadas al oeste de la Cordillera de las Cascadas, en comparación con el 63% de las zonas al este de las Cascadas.

Few studies have been published on the distribution and ecology of diatoms in the state of Washington, even though the state contains habitats ranging from low agricultural lands to shrubsteppe ecosystems, rainforests on the Olympic Peninsula, and true alpine

habitats. The high diversity of ecoregions in Washington should yield equally high diversity in diatom assemblages.

One of the earliest publications on diatoms in the state was by Sovereign (1963) who described 27 species of diatoms from springs

<sup>1</sup>Department of Biological Sciences, Northern Arizona University, Flagstaff, AZ 86011. E-mail: deandiacad@gmail.com

and lakes throughout the state. Blinn (1993) described the role of salinity on diatom communities in 3 salt lakes in the interior of Washington. Also, Bahls (2004–2006) provided a photographic catalogue with optimal physicochemical values, as well as a checklist from northwestern USA (Bahls 2009) of 118 genera and 380 diatom species for the state of Washington.

Other Washington-based research has examined diatoms in sediment cores to depict environmental and climatic changes over time (Bennett 1962, Stockner and Benson 1967, Tynni 1986, Wallace et al. 2006). Also, Hollingshead (2012) examined climate and hydrologic changes in Castor Lake, and Enache et al. (2013) reported 3 new *Psammothidium* species in sediment cores from oligotrophic lakes in Washington Cascade Mountain Lakes. Recently, Sheibley et al. (2014) noted an increase in *Asterionella formosa* and *Fragilaria tenera* in sediment cores from Hoh Lake in Olympic National Park beginning in 1969–1975 which suggested nitrogen enrichment during this period.

Hawkins and Norris (2000) suggested that indices based on the most robust and finest taxonomic resolution are preferable over family or generic resolution for the bioassessment of habitats and landscapes. In addition, Barbour et al. (1995) recommended a multimetric approach for measuring biological conditions. Stevenson et al. (2008) hypothesized that indicators based on species traits determined in streams with similar natural landscape features would be most effective in developing tolerance indices. More recently, Stevenson (2014) further suggested that future research on the regional-scale resilience of algal assemblages will improve water quality management. Kociolek (2006) discussed the limited checklists of diatom taxa for various regions in the USA and the need to develop species lists for various regions, especially regions such as the state of Washington with a wide variety of geomorphic conditions, land use, and aquatic habitats.

The distribution and ecology of diatoms were examined in the state of Washington, with tolerance values for common diatom species based on channel embeddedness, nutrients (total nitrogen [TN], total phosphorus [TP]), and specific conductance. These tolerance values provide a functional baseline for

monitoring changes in ecosystem health for aquatic ecosystems in the varied landscapes of the state (Blinn and Ruitter 2013).

Northwestern Washington is experiencing a rapid expansion in agricultural activities and urban centers (Washington State Agriculture: 2025; [www.agr.wa.gov](http://www.agr.wa.gov)). This study provides physicochemical and biotic baselines to monitor future changes in aquatic ecosystems in northwestern Washington. The following objectives were addressed in this paper: (1) determine the diatom flora in the northwest and central portions of the state of Washington, including the Olympic Peninsula; (2) determine the role land use plays in determining the diatom flora on the west and east sides of the Cascade Range; (3) develop a multimetric index to determine environmental tolerance levels to embeddedness, TN, TP, and specific conductance for dominant diatom species in the region; (4) determine the distribution and relative abundance of the invasive *Didymosphenia geminata* on the west and east sides of the Cascade Range.

#### STUDY AREA

The study area encompasses about 50,000 km<sup>2</sup> in 9 counties in northwest and central Washington, including Clallam and Jefferson counties on the Olympic Peninsula. The region includes the North Pacific Coastal and Columbia Glaciated freshwater ecoregions with a variety of temperate coastal streams, rivers, wall seeps, and ponds/lakes (Abell et al. 2000). The North Pacific Coastal region includes mesic landscapes on the west side of the North Cascade Range and a drier Columbia Glaciated region east of the Cascade Range. The study area also encompassed 5 physiographic regions including the Olympic Mountains, Puget Lowland, Northern Cascades, Okanogan West, and Columbia Basin (Rosenfeld 1985).

These landscapes contain a number of urban communities with a combined total of over 300,000 residents (United States Census Bureau 2013). Bellingham is the largest urban center with over 80,000 residents; most other urban centers have <10,000 residents (Blinn and Ruitter 2013). Dairy, corn, and berry farming make up a large portion of the agriculture in the lowland landscapes on the west side of the Cascade Range, while dryland crops and rangelands are common on the east side of

the Cascade Range (personal observation). The urban and agricultural landscapes west of the Cascades are typically <300 m in elevation, and most forest landscapes range from 300 to 2000 m. In contrast, agricultural landscapes east of the Cascade range are typically >300 m. Average elevation for the study area on the western side of the Cascade Range was 226 m (SE 27), while study areas east of the range averaged 638 m (SE 30).

Riparian vegetation in urban landscapes includes *Acer macrophyllum*, *Alnus rubra*, and *Pseudotsuga menziesii*, while agricultural landscapes include grasses, sedges, and *A. rubra*. Riparian vegetation along lowland forest sites contain *A. macrophyllum*, *A. rubra*, *Salix scouleriana*, *Salix sitchensis*, *P. menziesii*, and *Populus balsamifera* ssp. *trichocarpa*, which is largely replaced by *Tsuga heterophylla* and *Thuja plicata* in lowland riparian systems (Blinn and Ruiter 2013). *Picea sitchensis* is also present along the streams in the lowlands on the Olympic Peninsula. Riparian vegetation in drier eastern Washington includes *P. menziesii*, *Pinus ponderosa*, and dry grassland vegetation (personal observation).

#### METHODS

Diatoms were collected from 99 streams, 41 rivers, 9 wall seeps (water running down rock faces), and 16 ponds/lakes based on access during mid-June through September from 2008 through 2012 in northwest and north central Washington, including the Olympic Peninsula (Appendix 1). Diatoms were collected by scraping algal material from an approximate 3 × 3-cm area from rock and sediment substrates and by collecting submerged leaves and moss at all sites. The diatom floras by Krammer and Lange-Bertalot (1986, 1988, 1991a, 1991b, 2000) were routinely used for the identification of species.

Land use (urban, agriculture, forest) was noted and physicochemical information was collected at each site. Agricultural regions included cornfields and livestock pastures with cattle and horses; urban areas had ≥1000 residents; and forested land contained >50% tree cover. Maximum water temperatures were measured with a handheld thermometer on-site. Specific conductance (at 25 °C) of water was determined with a conductance meter (Radiometer, Copenhagen, Denmark;

Model CDM2e) in the laboratory, and pH was measured with an Oakton pH 6 meter on-site. Channel embeddedness was determined to the nearest 5% interval at each site following Platts et al. (1983). Suspended sediments were determined according to MacDonald et al. (1991), and canopy cover was determined with a spherical densiometer (Forest Densiometer Model A) to the nearest 5% interval.

Water collections for TN and TP were immediately placed on dry ice and analyzed within 14 days by the persulfate digestion and flow injection method on an FS3000 Flow Injection Analyzer made by OI Corporation (APHA 2005). The digestion method was an alkaline oxidation (APHA 4500-PJ) using sodium hydroxide and potassium persulfate. Following the digestion, TP was measured using the ascorbic acid method (APHA 4500-PH) and TN was measured using the cadmium reduction method on the FS3000 (APHA 4500 NO3-I).

Kendall's tau ( $\tau$ ) rank correlation coefficients were calculated between species richness and TN, TP, specific conductance, altitude, maximum water temperature, pH, and percent canopy. Principal components analysis was used based on singular value decomposition of the centered, scaled data matrix to extract the important relationships among water quality and diatom variables (Jongman et al. 1995, Ben-Hur and Guyon 2003, R Development Core Team 2011). Principal components were used as new variables to identify stable clusters using hierarchical clustering with Euclidean distance and Ward's minimum variance cluster method, following the methods outlined by Ben-Hur and Guyon (2003). Rare species were included in the analyses. The PCA variables were centered on the mean and scaled to unit variance. Shannon's diversity index was also calculated for diatom assemblages at each site.

A tolerance index was used for channel embeddedness, TN, TP, and specific conductance to compare the distribution of diatom species along each of these environmental gradients. The following equation was developed by Blinn (1993) for specific conductance (SCI) to examine the distribution of diatoms in salt lakes in western USA and expanded by Blinn and Ruiter (2006, 2013) for caddisflies:

$$SCI_x = \frac{\sum_{i=1}^{N_x} [\log_{10} (RA_i \cdot 100)] (\text{Specific Conductance, } \mu\text{S/cm})}{N_x}$$

where  $RA$  = relative abundance (percent) of species  $x$  at a given stream site, and  $N$  = number of streams in which species  $x$  occurred. The diatom tolerance index (DTI) is a summation of the adjusted combined numeric values of total nitrogen index (TNI), total phosphorus index (TPI), specific conductance index (SCI), and embedded index (EMBI). Elevated nutrients, salinization of water, and siltation of cobbles are 3 of the most influential stressors for stream biota (Laws 2000, Enger and Smith 2009). We adjusted the DTI numeric scores to a 0–10 scale for comparison.

## RESULTS

### Diatom Distribution and Physicochemical Features

Four hundred and fifteen diatom species were identified from streams, rivers, wall seeps and pond/lake sites in northwest and north central Washington (Appendix 2). Three hundred and fifty-two species were collected from streams/rivers, 87 species from wall seeps, and 213 species from ponds/lakes.

Aquatic systems sampled west of the Cascade Range averaged 278 m (SE 30.5) in altitude, while those east of the Cascade Range averaged 855 m (SE 68.3).  $H'$  ranged from 1.57 to 5.06 west of the Cascades, and from 1.85 to 4.98 east of the Cascade Range. Nearly 77% of the taxa were either mono- or biraphid diatoms. Species richness averaged 30.5 (SE 1.3) per site on the west side of the Cascades and 43 (SE 2.8) on the east side. *Achnantheidium minutissimum*, *Cocconeis placentula* f. *lineata*, *Diatoma mesodon*, *Encyonema silesiacum*, *Eunotia bilunaris*, *Fragilaria capucina*, *Fragilaria vaucheria*, *Frustulia vulgaris*, *Gomphonema parvulum*, *Meridion circulare*, *Nitzschia dissipata*, *Planothidium lanceolatum*, and *Ulnaria ulna* occurred in over 50% of the sites examined throughout the study area.

Ponds and lakes had the highest average number of species at 42.2 (SE 3.2) per site (Fig. 1).  $H'$  diversity for these habitats at lower elevations (<700 m) ranged from 2.83 to 4.77 with an average of 34 (SE 2) species, while those at higher elevations (>1150 m) ranged from 3.53 to 5.19 with an average of 56 (SE 7)

species. *Achnantheidium minutissimum*, *C. placentula* v. *lineata*, *E. bilunaris*, *F. capucina*, *F. vaucheria*, *U. ulna*, *Gomphonema truncatum*, and *Staurosira construens* f. *venter* were common in low-elevation ponds and lakes, while *A. minutissimum*, *Aulacoseira alpigena*, *D. mesodon*, *Eunotia* spp., *Frustulia* spp., *Pinnularia* spp., and *Tabellaria flocculosa* were common taxa in high-elevation lakes.

Urban streams and rivers averaged 38 (SE 2.6) species compared to 33 (SE 2.1) species in agricultural landscapes, and 26 species (SE 1.9) in forest landscapes (Fig. 1). *Cocconeis placentula* v. *lineata*, *F. capucina*, *P. lanceolatum*, *U. ulna*, and *Rhoicosphenia abbreviatum* were common species in urban streams, while *Melosira varians*, *N. dissipata*, *Nitzschia linearis*, *Nitzschia palea*, and *U. ulna* were common taxa in lowland agricultural streams and rivers.  $H'$  diversity ranged from 2.98 to 4.92 for urban streams and from 3.33 to 5.04 for lowland agricultural streams. *Aulacoseira alpigena*, *Diatoma hyemalis*, *D. mesodon*, *Eunotia* spp., *Hannaea arcus*, *M. circulare*, *Pinnularia divergens*, and *T. flocculosa* were common in streams and rivers at higher elevations (>700 m) on both sides of the Cascades.  $H'$  diversity in high-elevation streams ranged from 2.45 to 4.27 on the west side of the Cascades and from 2.90 to 4.98 on the east side. Wall seeps had the lowest average number of species (18, SE 2.1) with  $H'$  ranging from 1.57 to 2.99. *Diatoma mesodon*, *D. hyemalis*, *Diatomella balfourinia*, *F. capucina*, *F. vaucheria*, *Eunotia* spp., *T. flocculosa*, *Tetracyclus glans*, and *Tetracyclus rupestris* were common assemblages in wall seeps.

pH ranged from 6.6 to 8.3, with a mode of 7.1 throughout the study area. Average TN was nearly 6-fold higher on the west side of the Cascades compared to the east side (Table 1). TN and TP values for streams west of the Cascades were 725  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 122) and 18.5  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 3.8) compared to 88.7  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 21.7) and 21.7  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 5.3) east of the Cascades. Lowland sites in the agricultural region west of the Cascades had the highest average TN (1673  $\mu\text{g} \cdot \text{L}^{-1}$ , SE 349) and TP levels (63.5  $\mu\text{g} \cdot \text{L}^{-1}$ , SE 21) and the highest average specific conductance (2.75  $\text{mS} \cdot \text{cm}^{-1}$ , SE 2.3). Channel embeddedness averaged 85% (SE 7.1). Average TN and TP values for rivers west of the Cascades were 101.5  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 12.6) and 18.9  $\mu\text{g} \cdot \text{L}^{-1}$  (SE 3.5), respectively, compared to 115.7  $\mu\text{g} \cdot \text{L}^{-1}$

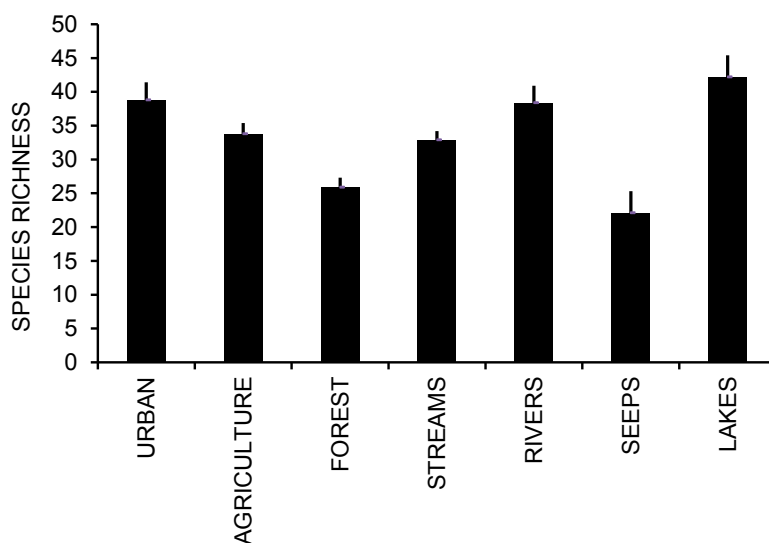


Fig. 1. Mean diatom species richness for land use and habitats in northwest and north central Washington.

TABLE 1. Average values (standard errors in parentheses) for physicochemical features of aquatic systems on the west and east sides of the North Cascade Mountain Range.

Physicochemical feature	West	East
Altitude (m)	278 (30.5)	855 (68.3)
Total phosphorus ( $\mu\text{g} \cdot \text{L}^{-1}$ )	20.5 (3)	16.2 (2.5)
Total nitrogen ( $\mu\text{g} \cdot \text{L}^{-1}$ )	452 (79)	80.2 (11.2)
Specific conductance ( $\text{mS} \cdot \text{cm}^{-1}$ )	0.379 (0.254)	0.122 (0.02)
Canopy cover (%)	47 (3.6)	38 (6)
Embeddedness (%)	35 (3.1)	16.8 (3.5)

TABLE 2. Kendall's  $\tau$  rank-based correlations between diatom species richness and environmental determinants.

	Kendall's $\tau$
Altitude	-0.050
Specific conductance**	0.146
Maximum water temperature**	0.171
Total phosphorus**	0.210
Total nitrogen*	0.106
pH**	0.156
% Canopy cover*	-0.136
Channel embeddedness**	0.213

\* $P \leq 0.05$

\*\* $P \leq 0.01$

(SE 21.7) and  $18.3 \mu\text{g} \cdot \text{L}^{-1}$  (SE 3) east of the Cascades.

Diatom species richness showed significant positive correlations to channel embeddedness, TN, TP, specific conductivity, maximum water temperature, and pH, and a significant negative correlation to percent canopy cover (Table 2). More detailed information on physicochemi-

cal features between landscapes (urban, agriculture, forest) and habitats (stream, river, seep, pond/lake) in the study area can be found in Blinn and Ruiter (2013).

Hierarchical clustering produced 6 stable cluster groups using the first 3 principal components as clustering variables (Fig 2, Table 3). These cluster groups appeared to match natural groups formed by geographic location and land-use type. A simple chi-square contingency table showed a significant association between the 6 "blind" hierarchical cluster groups and the following location/land-use groups (Crawley 2007).

Cluster A included 7 stream, 10 river, and 11 pond/lake sites primarily west of the Cascade Range at an average altitude of 88 m (SE 19), a canopy cover of 8.3% (SE 2), and a channel embeddedness of 92% (SE 1.0) (Table 3). These sites were primarily located in the lowland agricultural region (berry, dairy, and corn) west of the Cascades in Whatcom County. The only sites east of the Cascade Range included

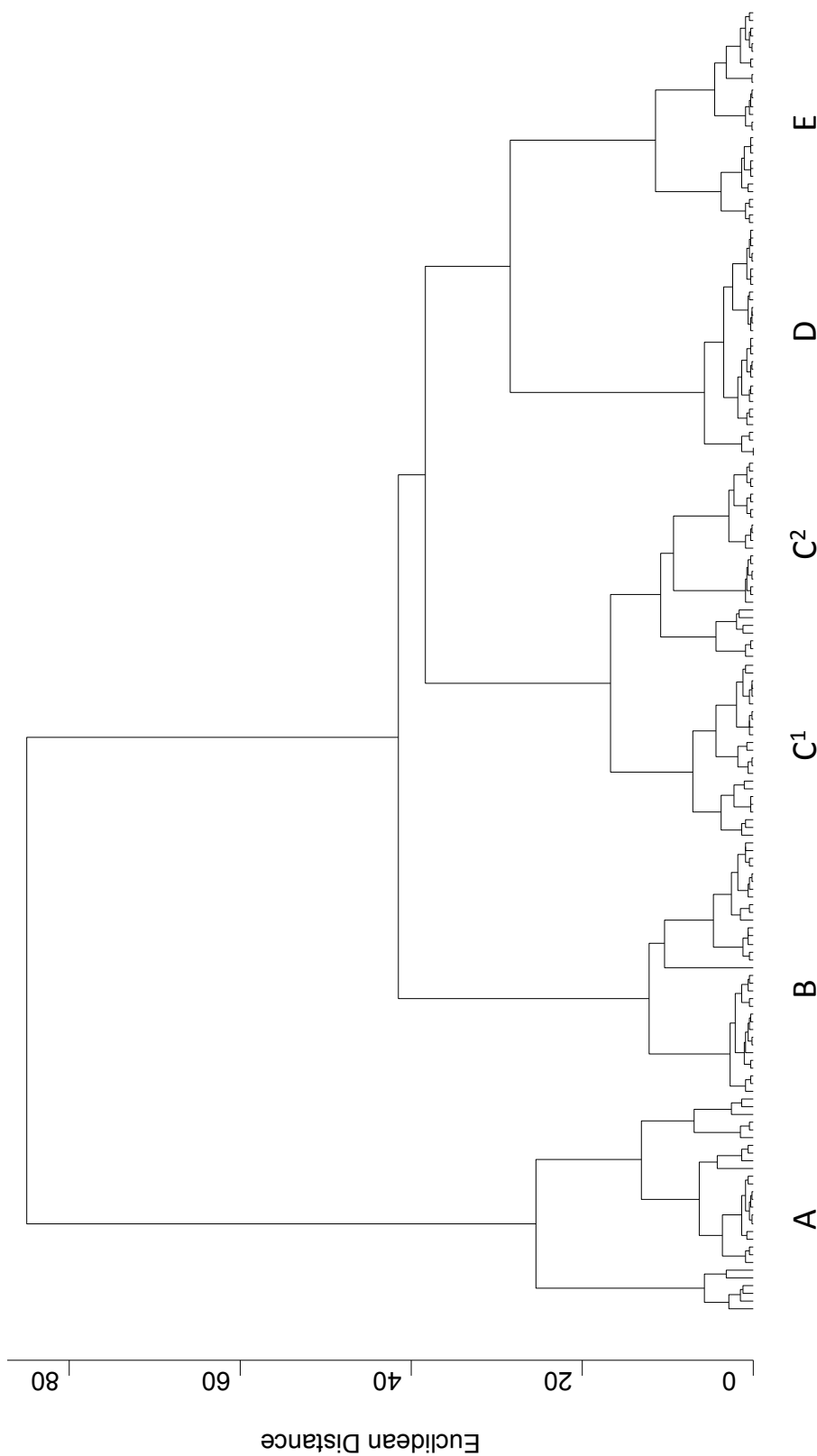


Fig. 2. Multilevel hierarchical clustering using 3 environmental factors for seeps, streams, and rivers in northwest and north central Washington. Cluster letters are provided in Appendix 1. A = sites at low altitudes in a dairy and berry agricultural region primarily on the west side of the Cascades (28 sites); B = sites located in municipal areas and regions of moderate agriculture primarily west of the Cascades; 6 of these sites are located on the Olympic Peninsula (33 sites); C1 = mainly stream sites primarily west of the Cascades at moderate altitudes, many of which are located in municipal areas and regions of moderate agriculture (21 sites); C2 = sites mainly on the east side of the Cascades with moderate agriculture (25 sites); D = stream and river sites all west of the Cascades with moderate agriculture (30 sites); E = high-altitude sites, including streams, rivers, and wall seeps on both sides of the Cascades (28 sites).

TABLE 3. Average values (standard errors in parentheses) for physicochemical features of the 6 hierarchical cluster groups for aquatic habitats in western Washington.

Physicochemical feature	Cluster group					
	A	B	C1	C2	D	E
Altitude (m)	88 (19)	337 (69)	169 (51)	487 (42)	208 (27)	850 (64)
Total phosphorus ( $\mu\text{g} \cdot \text{L}^{-1}$ )	48 (10)	129 (25)	20 (3.6)	8.8 (1.2)	13.9 (2.3)	7.1 (1.2)
Total nitrogen ( $\mu\text{g} \cdot \text{L}^{-1}$ )	898 (196)	138 (21)	526 (69)	90 (16)	285 (51)	60 (8.2)
Specific conductance ( $\text{mS} \cdot \text{cm}^{-1}$ )	1.17 (0.9)	0.08 (0.01)	0.174 (0.01)	0.09 (0.01)	0.109 (0.03)	0.07 (0.01)
% Canopy cover	8.3 (2)	18.6 (3.1)	76 (6)	54 (7)	85 (2.9)	39 (7.2)
% Embeddedness	92 (1)	55 (6.2)	18.5 (2.6)	12.2 (1.7)	12.3 (1.5)	7.3 (0.8)
Temperature ( $^{\circ}\text{C}$ )	21 (0.6)	16.3 (0.3)	18.2 (0.5)	16.6 (0.4)	16 (0.2)	14.1 (0.2)

the Columbia and Okanogan rivers. TN and TP averaged  $898 \mu\text{g} \cdot \text{L}^{-1}$  (SE 196) and  $48 \mu\text{g} \cdot \text{L}^{-1}$  (SE 10), respectively, for sites in this cluster. These sites had the lowest average altitude and the highest average channel embeddedness, TN, specific conductance, and water temperature of all sites examined.

Cluster B included 7 stream, 20 river, and 6 pond/lake sites at an average altitude of 337 m (SE 69) and a canopy cover of 18.6% (SE 3.1). This cluster included 30 sites west and 3 sites east of the Cascades. Six river and stream sites were located on the Olympic Peninsula. TN and TP averaged  $138 \mu\text{g} \cdot \text{L}^{-1}$  (SE 21) and  $19.8 \mu\text{g} \cdot \text{L}^{-1}$  (SE 3), respectively. Average water temperature and specific conductance for sites in this cluster were  $16.3^{\circ}\text{C}$  (SE 0.3) and  $0.08 \text{mS} \cdot \text{cm}^{-1}$  (SE 0.01), respectively. This cluster had one of the lowest average specific conductance values.

Cluster C1 included 21 stream and 2 river sites located at an average altitude of 169 m (SE 51) and a canopy cover of 76% (SE 6). Nineteen sites were located on the east side of the Cascades. TP averaged  $526 \mu\text{g} \cdot \text{L}^{-1}$  (SE 69) for sites on the west side and  $20 \mu\text{g} \cdot \text{L}^{-1}$  (SE 3.6) east of the Cascades. Average water temperature and specific conductance were  $18.2^{\circ}\text{C}$  (SE 0.5) and  $0.174 \text{mS} \cdot \text{cm}^{-1}$  (SE 0.01). Many of these sites were located in small municipal regions or areas with moderate agriculture. Cluster C2 included 17 stream and 9 river sites at moderate altitudes (487 m, SE 42) and canopy cover (54%, SE 7). Seventeen sites were located on the east side of the Cascades.

Average water temperatures and embeddedness were similar to C1 sites west of the Cascades, while average specific conductance, TN, and TP were considerably lower (Table 3).

Cluster D included a total of 30 stream and river sites, all west of the Cascades with 3 sites on the Olympic Peninsula. The average altitude of these sites was 208 m (SE 27) with a canopy cover of 85% (SE 2.9). TN and TP averaged  $285 \mu\text{g} \cdot \text{L}^{-1}$  (SE 51) and  $13.9 \mu\text{g} \cdot \text{L}^{-1}$  (SE 2.3), and specific conductance averaged  $0.109 \text{mS} \cdot \text{cm}^{-1}$  (SE 0.03).

Cluster E included 28 high-altitude (850 m; SE 64), cool water ( $14.1^{\circ}\text{C}$ ; SE 0.2) streams, rivers, and wall seeps. This included 20 sites west and 8 sites east of the Cascade Range. TN and TP averaged  $60 \mu\text{g} \cdot \text{L}^{-1}$  (SE 1.2) and  $7.1 \mu\text{g} \cdot \text{L}^{-1}$  (SE 1.2), while specific conductance averaged  $0.07 \text{mS} \cdot \text{cm}^{-1}$  (SE 0.01). Sites in this cluster had the highest average altitude and lowest average water temperature, specific conductance, TN, TP, and channel embeddedness of any cluster (Table 3).

The invasive alga *Didymosphenia geminata* was collected in 46% of the stream and river sites sampled and ranged from <1% to 27% in relative abundance, but was not collected in ponds/lakes or wall seeps. *Didymosphenia geminata* showed significant ( $P = 0.05$ ) negative correlations to altitude ( $r = -0.138$ ) and TN ( $r = -0.127$ ) and a significant positive correlation (0.319) to suspended sediment. The average relative abundance of *D. geminata* was 2.2% (SE 0.6) in streams compared to 4.7% (SE 2.1) in rivers. *Didymosphenia geminata* was collected

TABLE 4. Average physicochemical features (standard errors in parentheses) for sites with *Didymosphenia geminata* (all sites, sites west and east of the Cascade Mountain Range, and sites on the Olympic Peninsula [Oly Pen]). ALTI = altitude, MTEMP = maximum water temperature, TP = total phosphorus, TN = total nitrogen, SPCO = specific conductance, CAN = % canopy, EMBE = % embeddedness, SUSE = suspended sediments, % relative abundance = percent relative abundance of *Didymosphenia geminata*,  $n$  = number of sites.

	ALTI (m)	MTEMP (°C)	TP ( $\mu\text{g} \cdot \text{L}^{-1}$ )	TN ( $\mu\text{g} \cdot \text{L}^{-1}$ )	SPCO ( $\text{mS} \cdot \text{cm}^{-1}$ )	CAN (%)	EMBE (%)	SUSE ( $\text{mg} \cdot \text{L}^{-1}$ )	% Relative abundance
All sites ( $n = 64$ )	300 (28.6)	15.6 (0.5)	16.1 (1.8)	177.2 (41.5)	0.558 (0.428)	34.4 (4.2)	27.8 (3.7)	16.6 (6.2)	2.8 (0.8)
West ( $n = 37$ )	159 (23)	16.0 (0.6)	16.0 (2.5)	240 (66)	0.837 (0.71)	41.6 (5.8)	33.9 (5.1)	23.4 (10.1)	3.7 (1)
East ( $n = 21$ )	519 (29)	15.1 (0.8)	16.5 (2.8)	83 (13)	0.125 (0.03)	25.8 (5.5)	18.3 (4.7)	6.1 (1.4)	2.8 (1.2)
Oly Pen ( $n = 6$ )	97 (38)	14.4 (0.7)	8 (2.4)	126 (38)	0.092 (0.01)	29.2 (9.8)	20.0 (4)	20.0 (4)	4.3 (1.2)

in 87% of the river sites examined on the mainland and all river sites on the Olympic Peninsula except the Elwa River. This invasive alga was present throughout the main channel of the Nooksack River, including the north, middle, and south forks (Appendix 1). In contrast, *D. geminata* was only collected in 33% of the stream sites examined.

Average physicochemical features of habitats throughout the study area with *D. geminata*, including the Olympic Peninsula, are presented in Table 4. The average TN was  $134 \mu\text{g} \cdot \text{L}^{-1}$  (SE 35) in streams compared to  $93 \mu\text{g} \cdot \text{L}^{-1}$  (SE 7.9) in rivers. Also, specific conductance in streams with *D. geminata* averaged  $0.08 \text{ mS} \cdot \text{cm}^{-1}$  (SE 0.02) compared to  $3 \text{ mS} \cdot \text{cm}^{-1}$  (SE 1.9) in rivers. Average water temperature for *D. geminata* on the Olympic Peninsula was  $14.4^\circ\text{C}$  (SE 0.7;  $n = 5$ ) compared to  $17^\circ\text{C}$  (SE 0.3;  $n = 59$ ) on the mainland. Average water temperature for aquatic systems with this alga throughout the study area was  $15.6^\circ\text{C}$  (SE 0.5;  $n = 64$ ).

The average TN for rivers with *D. geminata* was  $98.4 \mu\text{g} \cdot \text{L}^{-1}$  (SE 10.7) compared to  $491 \mu\text{g} \cdot \text{L}^{-1}$  (SE 131) for those without this alga. Stream sites with *D. geminata* had a TN value of  $131.4 \mu\text{g} \cdot \text{L}^{-1}$  (SE 23.6) compared to  $838.3 \mu\text{g} \cdot \text{L}^{-1}$  (SE 182) for those without *D. geminata*. All sites with this alga had an average TN value of  $127 \mu\text{g} \cdot \text{L}^{-1}$  (SE 19.4), while sites without this alga had an average TN value of  $788 \text{ mg} \cdot \text{L}^{-1}$  (SE 163). The average relative frequency of *D. geminata* was 2.2% (SE 0.6) in streams compared to 4.7% (SE 2.1) in rivers.

The average relative abundance of *D. geminata* in habitats west of the Cascades was 3.7% (SE 1;  $n = 37$ ) compared to 2.8% (SE 1.2;  $n = 22$ ) on the east side. The relative abundance of *D. geminata* on the Olympic Peninsula was 4.3% (SE 1.2). Average TP and TN concentrations for all sites with *D. geminata* throughout the study area were  $16.1 \mu\text{g} \cdot \text{L}^{-1}$  (SE 1.8) and  $177.2 \mu\text{g} \cdot \text{L}^{-1}$  (SE 41.5), respectively. Highest TN value for streams with *D. geminata* was  $408 \mu\text{g} \cdot \text{L}^{-1}$  in Thompson Creek compared to  $278 \mu\text{g} \cdot \text{L}^{-1}$  in the Calawah River for river systems. Fifty-five percent of the sites with *D. geminata* had TP values  $\leq 10 \mu\text{g} \cdot \text{L}^{-1}$ , while 53% had TN values  $\leq 100 \mu\text{g} \cdot \text{L}^{-1}$ . Mean TP and TN for streams with *D. geminata* west of the Cascades was  $10.1$  (SE 1.9) and  $154.8 \mu\text{g} \cdot \text{L}^{-1}$  (SE 21.8), respectively, compared to 9.7



(SE 2.8) and 52.4 (SE 16.4)  $\mu\text{g} \cdot \text{L}^{-1}$  east of the Cascades. *D. geminata* made up 5.4% (SE 1.2) of the diatom assemblage in streams and rivers with  $\geq 5 \text{ mg} \cdot \text{L}^{-1}$  suspended sediment, compared to 2.2% (SE 0.4) for those with  $< 5 \text{ mg} \cdot \text{L}^{-1}$  suspended sediment. Other physicochemical features for habitats with *D. geminata* are provided in Table 4.

#### Diatom Tolerance Values

A diatom tolerance index (DTI) for channel embeddedness (EMBI), nutrients (TNI, TPI), and specific conductance (SCI) is provided in Table 5 for 114 of the most common diatom taxa in the region. The percent relative frequency for each species throughout the study area is also provided. The adjusted DTI values (ADJUS) ranged from 0 to 10. Tolerance values  $\leq 4.0$  are considered more sensitive to the measured environmental parameters, while tolerance values  $\geq 6.0$  are considered more tolerant.

The majority of taxa with DTI values  $\leq 4.0$  were either araphid or monoraphid (e.g., *Aulacoserira alpigena*, *Diatomella balfourniana*, *Eumotia paludosa*, *Gomphonema affine*, *Orthoseira dendroteres*, and *Tetracyclus glans*, etc.), and occurred on hard substrata with limited motility. These taxa were common in the higher-altitude sites. In contrast, 55% of the species with DTI values  $\geq 6.0$  were keeled or canal-raped taxa (e.g., *Cymatopleura solea*, *Nitzschia amphibia*, *N. dissipata*, *N. linearis*, *N. palea*, and *Surirella ovalis*, etc.) and occurred in habitats with elevated embeddedness and nutrients in stream channels throughout the study area.

Genera, which included species with active movement (*Navicula*, *Nitzschia*, and *Surirella*), made up  $< 10\%$  of the taxa with ADJUS values  $\leq 4.0$ , while species in these genera made up over 70% of taxa with ADJUS values  $\geq 6.0$ . *Achnanthydium minutissimum*, *C. placentula* f. *lineata*, *D. mesodon*, *E. silesiacum*, *F. capucina*, *F. vaucheria*, *F. vulgaris*, *G. parvulum*, *P. lanceolatum*, *N. dissipata*, and *U. ulna* occurred in over 50% of the sites with WADJUS values ranging from 4.6 to 6.4. Araphid, monoraphid, and centric diatoms typically had lower EMBI values, which implied species associated with vegetation and hard surface substrates. Biraphid diatoms were more common along increasing EMBI values which suggested habitats with fine sediments.

#### DISCUSSION

Diatom species richness declined as stream channel conditions deteriorated throughout the Washington landscape. Cooke and Prepas (1998) reported that crop fertilizers and animal manure constituted a large portion of non-point-source pollution of phosphorus and nitrogen in streams. Also, Lee and Ziegler (2010) found that urbanization, construction activity, and impoundments influence rates of embeddedness and salinity in streams, while Kauffman and Krueger (1984) reported livestock impacts on riparian ecosystems. These conditions also influenced both composition and species richness in diatom communities throughout the Washington landscape. Fore and Grafe (2002) developed a river diatom index (RDI) for Idaho rivers and found it to be significantly correlated with human disturbance at the site (i.e., conductivity, percentage of fine sediments, and number of human activities) and at the catchment level (i.e., percentage of urbanization and agriculture in the upstream catchment).

The hierarchical clustering model indicated that diatoms were useful environmental indicators of both land use and altitudinal gradients in Washington aquatic ecosystems. The model separated diatom assemblages into distinct clusters that ranged from lowland aquatic habitats with heavy crop and dairy farming to those with low to moderate agriculture activity at moderate to high altitudes west of the Cascades. The model also separated diatom communities located west and east of the Cascades. Use of dominant diatom taxa associated with low and highly disturbed habitats provides a useful monitoring program for the region.

DTI values for embeddedness, nutrients, and salinity for each species enhanced the ability to determine which environmental parameter(s) are responsible for conditions in the stream systems in the Washington landscape. Diatom species showed distinct associations with physicochemical conditions in aquatic habitats throughout the study area. The majority of taxa with DTI values  $\leq 4.0$  were either araphid, monoraphid, or biraphid, and their presence suggested hard substrata and limited motility. These taxa were common in the higher-altitude sites.

In contrast, 55% of the species with DTI values  $\geq 6.0$  were keeled or canal-raped taxa,

TABLE 5. Diatom tolerance index (DTI) for 114 species collected in Washington. Ranking of diatom taxa is based on combined tolerance values for channel embeddedness (EMBI), total phosphorus (TPI), total nitrogen (TNI), and specific conductance (SCI). Combined tolerance values (ADJUS) were adjusted to a scale of 0 to 10. Species were arranged from most sensitive to most tolerant based on combined tolerance values. % Relative frequency = percentage of sites in which species occurred throughout the study area.

Diatom species	EMBI	TPI	TNI	SCI	DTI	ADJUS	% Relative frequency
<i>Diatomella balfouriana</i>	21	2	38	101	162	0.8	11.0
<i>Orthoseira dendroteres</i>	11	20	52	84	167	0.8	1.2
<i>Tetracyclus rupestris</i>	15	23	81	88	207	1.0	4.5
<i>Rossithidium nodosum</i>	17	23	87	139	266	1.3	4.5
<i>Orthoseira roeseana</i>	42	12	91	123	268	1.3	2.5
<i>Epithemia argus</i>	14	11	125	120	270	1.4	4.0
<i>Pinnularia borealis</i>	48	12	204	142	406	2.0	5.0
<i>Tetracyclus glans</i>	43	17	203	149	412	2.1	3.2
<i>Aulacoseira alpigena</i>	44	18	243	152	457	2.3	7.6
<i>Surirella linearis</i>	25	40	228	165	458	2.3	5.7
<i>Eumotia soleirolii</i>	54	24	272	113	463	2.3	8.9
<i>Pinnularia subcapitata</i>	59	14	317	86	476	2.4	5.4
<i>Epithemia adnata</i>	31	49	199	236	515	2.6	4.5
<i>Caloneis tenuis</i>	62	15	263	182	522	2.6	4.5
<i>Nitzschia angustata</i>	73	26	209	253	561	2.3	3.8
<i>Rossithidium petersenii</i>	35	27	349	177	588	2.9	6.4
<i>Diatoma anceps</i>	54	21	341	175	591	3.0	5.7
<i>Eumotia paludosa</i>	66	37	330	164	597	3.0	13.3
<i>Tabellaria flocculosa</i>	72	25	348	153	598	3.0	34.4
<i>Diatoma hyemalis</i>	58	29	333	184	604	3.0	17.8
<i>Fragilaria virescens</i>	77	18	431	89	615	3.1	5.1
<i>Gomphonema affine</i>	32	16	411	184	643	3.2	6.4
<i>Aulacoseira distans</i>	65	9	470	113	657	3.3	3.9
<i>Eumotia monodon</i>	62	25	450	150	687	3.4	3.2
<i>Psammothidium subatomoides</i>	45	22	475	150	692	3.5	28.6
<i>Didymosphenia geminata</i>	73	28	406	200	707	3.5	2.6
<i>Hannaea arcus</i>	63	35	404	211	713	3.6	49.2
<i>Caloneis alpestris</i>	68	22	376	258	724	3.6	5.5
<i>Rhopalodia gibba</i>	67	39	407	226	739	3.7	15.9
<i>Achnanthyidium exile</i>	67	31	464	178	740	3.7	10.8
<i>Gomphonema olivaceum</i>	51	26	486	180	743	3.7	16.5
<i>Eucocconeis laevis</i>	45	24	467	211	747	3.7	7.0
<i>Diploneis oblongella</i>	24	23	494	219	760	3.8	3.8
<i>Tabellaria fenestrata</i>	67	51	395	249	762	3.8	5.1
<i>Diatoma vulgare</i>	43	34	486	227	790	4.0	24.2
<i>Cymbella affinis</i>	56	29	501	206	792	4.0	49.7
<i>Caloneis schumanniana</i>	29	20	549	223	821	4.1	5.5
<i>Amphipleura pellucida</i>	43	37	537	237	854	4.3	7.6
<i>Navicula margalithii</i>	54	43	493	267	857	4.3	6.3
<i>Gomphonema truncatum</i>	61	36	517	248	862	4.3	19.1
<i>Caloneis silicula</i>	56	24	620	163	863	4.3	11.5
<i>Eumotia pectinalis v. undulata</i>	61	33	586	199	879	4.4	23.0
<i>Brachysira brebissonii</i>	65	33	627	165	890	4.5	5.7
<i>Navicula recens</i>	55	24	620	204	903	4.5	11.5
<i>Diatoma mesodon</i>	56	33	584	239	912	4.6	62.0
<i>Sellaphora pupula</i>	76	58	528	256	918	4.6	11.4
<i>Gomphonema acuminatum</i>	57	29	587	246	919	4.6	21.6
<i>Hantzschia amphioxys</i>	49	51	589	232	921	3.6	11.5
<i>Caloneis sublinearis</i>	61	30	693	151	935	4.7	2.5
<i>Aulacoseira italica</i>	84	48	549	261	942	4.7	9.5
<i>Encyonema minutum</i>	69	35	628	214	946	4.7	24.0
<i>Reimeria sinuata</i>	49	31	656	215	951	4.8	47.0
<i>Fragilaria vaucheriae</i>	67	35	606	243	951	4.8	65.7
<i>Encyonopsis microcephala</i>	69	35	638	212	954	4.8	26.8
<i>Navicula radiosa</i>	65	46	627	224	962	4.8	12.0
<i>Psammothidium bioretii</i>	69	28	651	226	974	4.9	9.5
<i>Achnanthyidium minutissimum</i>	65	44	599	266	974	4.9	68.0
<i>Pinnularia divergens</i>	54	40	701	181	976	4.9	33.7

TABLE 5. Continued.

Diatom species	EMBI	TPI	TNI	SCI	DTI	ADJUS	% Relative frequency
<i>Navicula recens</i>	58	25	684	212	979	4.9	11.5
<i>Encyonema silesiacum</i>	72	41	633	244	990	5.0	64.7
<i>Fragilaria tenera</i>	51	34	688	221	994	5.0	14.6
<i>Eunotia bilunaris</i>	70	28	648	250	996	5.0	38.0
<i>Amphora pediculus</i>	42	40	675	253	1010	5.1	21.0
<i>Epithemia sorex</i>	61	33	688	236	1018	5.1	9.5
<i>Diatoma tenuis</i>	63	30	693	233	1019	5.1	16.5
<i>Nitzschia sigma</i>	76	24	723	197	1020	5.1	9.6
<i>Nitzschia fonticola</i>	76	46	670	232	1024	5.1	15.9
<i>Staurosira construens</i> f. <i>center</i>	72	43	645	270	1030	5.2	13.3
<i>Fragilaria capucina</i> v. <i>vaucheria</i>	69	40	664	258	1031	5.2	63.4
<i>Frustulia vulgaris</i>	64	34	713	227	1038	5.2	54.0
<i>Navicula cincta</i>	59	38	655	289	1041	5.2	4.5
<i>Gyrosigma acuminatum</i>	55	58	668	267	1048	5.2	5.1
<i>Nitzschia frustulum</i>	59	44	717	233	1053	5.3	36.9
<i>Gomphonema parvulum</i>	60	35	714	246	1055	5.3	63.0
<i>Staurosirella pinnata</i>	65	39	708	256	1068	5.3	8.3
<i>Staurosira construens</i>	65	47	706	252	1070	5.4	29.9
<i>Entomoneis paludosa</i>	105	31	723	221	1080	5.4	2.5
<i>Eunotia pectinalis</i> v. <i>undulata</i>	60	30	754	236	1080	5.4	28.0
<i>Cymbella aspera</i>	61	44	755	225	1085	5.5	16.6
<i>Cymbella tumida</i>	62	21	829	180	1092	5.5	7.6
<i>Ctenophora pulchella</i>	27	44	744	277	1092	5.5	3.2
<i>Gomphonema minutum</i>	76	37	730	259	1102	5.5	35.3
<i>Stauroneis anceps</i>	103	29	730	248	1110	5.6	5.1
<i>Planothidium lanceolatum</i>	62	43	763	264	1132	5.7	74.5
<i>Surirella brebissonii</i>	54	41	898	190	1183	5.9	8.0
<i>Meridion circulare</i>	66	43	838	239	1186	5.9	50.3
<i>Cocconeis placentula</i> f. <i>lineata</i>	72	46	801	270	1189	6.0	70.3
<i>Nitzschia vermicularis</i>	83	90	643	380	1196	6.0	5.2
<i>Nitzschia dissipata</i>	76	46	817	263	1202	6.0	53.0
<i>Luticola goeppertiana</i>	84	34	923	173	1214	6.1	5.1
<i>Navicula rhynchocephala</i>	79	30	843	265	1217	6.1	29.3
<i>Cymbopleura naviculiformis</i>	73	34	954	158	1219	6.1	7.6
<i>Nitzschia linearis</i>	53	47	870	258	1228	6.2	46.9
<i>Rhoicosphemia abbreviata</i>	61	44	862	286	1253	6.3	38.2
<i>Ulnaria ulna</i>	80	49	852	286	1267	6.4	77.8
<i>Nitzschia amphibia</i>	73	49	912	238	1272	6.4	6.4
<i>Amphora ovalis</i>	46	61	913	273	1293	6.5	5.6
<i>Cocconeis pediculus</i>	67	67	857	304	1295	6.5	8.9
<i>Stauroneis phoenicenteron</i>	106	45	865	289	1305	6.5	8.2
<i>Nitzschia palea</i>	70	42	937	258	1307	6.5	30.6
<i>Navicula gregaria</i>	69	52	929	267	1317	6.6	36.0
<i>Surirella brebissonii</i> v. <i>kuetzingii</i>	51	68	1032	259	1410	7.1	5.7
<i>Cymatopleura solea</i>	55	58	989	316	1418	7.1	8.3
<i>Nitzschia acicularis</i>	81	38	1052	289	1460	7.3	3.8
<i>Nitzschia filiformis</i>	76	50	1084	272	1482	7.4	31.8
<i>Surirella angustata</i>	61	53	1137	267	1518	7.6	16.7
<i>Aulacoseira ambigua</i>	86	57	1110	268	1521	7.6	22.3
<i>Hippodonta capitata</i>	86	57	1110	294	1547	7.7	9.5
<i>Entomoneis alata</i>	74	34	1160	366	1634	8.2	2.5
<i>Bacillaria paradoxa</i>	81	37	1202	332	1652	8.3	4.5
<i>Surirella minuta</i>	53	51	1352	269	1725	8.6	5.6
<i>Surirella ovalis</i>	72	61	1390	251	1774	8.9	3.2
<i>Melosira varians</i>	93	71	1277	358	1799	9.0	42.0
<i>Nitzschia sigmoidea</i>	102	127	1316	455	2000	10.0	5.7

and their presence suggested elevated embeddedness and nutrients in stream channels throughout much of the study area. These

species were common in the lowland agricultural region west of the Cascade Range. Van Dam et. al. (1994) and Potapova and Charles

(2007) also found these species to be associated with eutrophic conditions. In addition, 26% of the taxa were considered motile in soft sediments (Kociolek and Spaulding 2003). In contrast, species with the highest relative abundance typically had the greatest tolerance to variation in the measured parameters, whereas more sensitive species had the lowest tolerance.

The diatom tolerance index (DTI) indicated that keeled and canalled-raphed diatoms were more common in habitats with high EMBI values or highly silted substrates. It is notable that over 50% of the species with high WADJUS had keeled or canalled raphe structures. In addition, these taxa were associated with high TN values. In contrast, araphid and monoraphid species with WADJUS values  $\leq 4.0$  comprised 40% of taxa associated with substrates such as plants, sand, and rocks (Kingston 2004).

Blinn and Ruiter (2013) developed a caddisfly tolerance index (CTI) for caddisfly species collected throughout Washington in the same habitats at the same time as the diatoms. Tolerance values were comparable for the dominant components at both primary (diatom) and secondary (caddisfly) levels of the food chain. Diatom species with high DTI values occupied the same habitats as caddisfly species with high CTI values. For example, *C. placentula v. lineata*, *M. varians*, *Navicula rhynchocephala*, and *P. lanceolatum*, all with a DTI  $\geq 1132$ , made up 33% of the diatom assemblage in Johnson Creek, and *Hydrotilla xera* (CTI = 1097) made up 89% of the caddisfly assemblage. In contrast, *A. alpigena*, *D. balfouriana*, and *T. glans* in Upper Galena Cr. had low DTI values ranging from 162 to 457 (Table 5) and were associated with caddisflies (*Ecclisocosmoecus scylla*, *Hesperophylax alaskensis*, *Ecclisomyia conspersa*, *Ecclisomyia maculosa*) with CTI values  $\leq 133$ .

Table 6 provides a monitoring program utilizing diatom assemblages from 165 aquatic systems in the state of Washington. Diatom assemblages are provided for the 3 condition levels of stream environments (good, fair, and poor condition). Potapova and Charles (2007) found that diatom indices developed in certain parts of the USA are not as effective when used in other regions of the same continent. Therefore, DTI values for individual species derived from streams, rivers, and lakes in

Washington provide a more accurate index for monitoring the status of various watersheds in the state.

*Didymosphenia geminata* was first reported in North America on Vancouver Island, British Columbia, in the late 1800s (Spaulding and Elwell 2007). More recently, Bahls (2004–2006) reported low numbers of *D. geminata* in 11 stream and river periphyton samples collected in Washington. The most suitable habitats for *D. geminata* were predicted to occur in the western United States, in relatively cool sites, and at high elevations with a high base-flow index (Kumar et al. 2009).

Sutherland et al. (2007) reported that substrates colonized by *D. geminata* in the laboratory died or disappeared when positioned in spring-fed creeks in New Zealand, while those placed in rivers showed growth. In addition, they found water in spring-fed creeks had higher nitrate concentrations and specific conductivity than river waters.

Although nitrates were not measured, similar patterns were found for TN and specific conductivity in Washington streams and rivers. The average TN in streams was  $134 \mu\text{g} \cdot \text{L}^{-1}$  (SE 35) compared to  $93 \mu\text{g} \cdot \text{L}^{-1}$  (SE 7.9) in rivers. Furthermore, the average relative frequency of *D. geminata* was 2.2% (SE 0.6) in streams compared to 4.7% (SE 2.1) in rivers. This further suggests that elevated nitrogen has a negative influence on the growth of this invasive alga. Also, higher TN values in streams and rivers on the west side of the Cascades compared to those on the east side may explain the higher percentage of sites with *D. geminata* on the east side of the Cascades.

Eighty-seven percent of the river sites sampled contained *D. geminata* compared to 33% of the stream sites sampled. This likely resulted from higher TN concentrations in streams compared to rivers. Many of the small streams examined were spring-fed with high TN concentrations, while larger rivers had a lower average TN concentration. River sites without *D. geminata* had an average TN concentration of  $531 \mu\text{g} \cdot \text{L}^{-1}$  (SE 157). Twenty-nine river systems contained *D. geminata*, including all forks of the Nooksack River system. The transport of *D. geminata* by fishermen in the Nooksack River drainage may have been responsible for the high occurrence of this invasive species.

TABLE 6. Monitoring program for streams and rivers based on the diatom tolerance index (DTI) developed for the state of Washington. This program includes diatom assemblages associated with good, fair, and poor channel conditions based on collections from 165 stream, river, and pond/lake sites throughout the state. DTI is also presented for each species, as well as the mean relative frequency of occurrence for each assemblage. These assemblages are in general agreement with Bahls (2004–2006), Lowe (1974), Potapova and Charles (2007), and Van Dam et al. (1994).

Good channel conditions		Fair channel conditions		Poor channel conditions	
Species	DTI	Species	DTI	Species	DTI
<i>Aulacoseira alpina</i>	457	<i>Caloneis schumanniana</i>	821	<i>Aulacoseira ambigua</i>	1521
<i>Aulacoseira distans</i>	657	<i>Cymbella affinis</i>	792	<i>Bacillaria paradoxa</i>	1652
<i>Diatoma hyendalis</i>	604	<i>Cymbella timida</i>	1092	<i>Cocconeis pediculus</i>	1295
<i>Diatomella balfouritana</i>	162	<i>Diatoma mesodon</i>	912	<i>Entomoneis alata</i>	1652
<i>Eucocconeis laevis</i>	747	<i>Diatoma vulgare</i>	790	<i>Laticala goeppertiana</i>	1214
<i>Eumotia paludosa</i>	597	<i>Encyonema minutum</i>	946	<i>Melosira varians</i>	1799
<i>Emotia soleirolii</i>	463	<i>Epithemia sorex</i>	1018	<i>Meridion circulare</i>	1186
<i>Fragilaria virescens</i>	615	<i>Gomphonema acuminatum</i>	919	<i>Navicula rhynchocephala</i>	1217
<i>Hannaea arcus</i>	713	<i>Gyrosigma acuminatum</i>	1048	<i>Nitzschia amphibia</i>	1272
<i>Pinnularia borealis</i>	406	<i>Gomphonema parvulum</i>	1055	<i>Nitzschia palea</i>	1307
<i>Pinnularia subcapitata</i>	476	<i>Gomphonema truncatum</i>	862	<i>Nitzschia stigmioidea</i>	2000
<i>Psammolithidium subatomoides</i>	692	<i>Navicula recens</i>	979	<i>Nitzschia vermicularis</i>	1196
<i>Rossthidium petersenii</i>	588	<i>Nitzschia frustulum</i>	1053	<i>Planothidium lanceolatum</i>	1132
<i>Surirella linearis</i>	458	<i>Reimera sinuata</i>	951	<i>Rhoicosphenia abbreviata</i>	1253
<i>Tetracyclus glans</i>	412	<i>Stauronets anceps</i>	1110	<i>Surirella minuta</i>	1725
<i>Tetracyclus rupestris</i>	207	<i>Stauroneta construens</i>	1070	<i>Unaria ulna</i>	1267
Mean relative frequency	11.4% (SE 3%)		22.9% (SE 4.5%)		25.5% (SE 6.3%)

The absence of *D. geminata* in the Elwa River is likely due to the abrasive high suspended solids in this river. Similar patterns were observed in Glacier and Barr creeks, both with high suspended solid loads (Appendix 1). Although the Elwa River on the Olympic Peninsula had relatively low TN concentration, the abrasive nature of a high suspended sediment load may have restricted the occurrence of this invasive alga in this system (Appendix 1). However, Reid and Torres (2014) recently reported increases in soluble reactive phosphorus uptake within *D. geminata* biomass associated with fine sediments because of altered redox conditions within the algal biomass. *Didymosphenia geminata* ranked in the top 25% of the diatom tolerance index with a value of 707 (Table 5). The negative correlation to altitude may have resulted from limited dispersal vectors to isolated, high-altitude sites.

Studies of streams in New Zealand and North America have demonstrated that *D. geminata* populations decrease with high numbers of Ephemeroptera, Plecoptera, and Trichoptera insect trophic groups, and increase with dipterans, crustaceans, and oligochaetes (Gillis and Chalifour 2010, James et al. 2010, Larned and Kilroy 2014). This has a major effect on the aquatic food web in streams and rivers.

Future studies are needed in the Columbia Unglaciaded freshwater ecoregion that runs along the southern border of Washington and into Oregon and Idaho in the Blue Mountains of the Umatilla National Forest (Abell et al. 2000). This region escaped glaciation during the Pleistocene and may yield endemic diatoms not included in this study or in Bahls's (2009) checklist of species in northwestern USA. This may be especially true for seeps and springs in the region.

#### ACKNOWLEDGMENTS

I thank Ashley Rawhouser and Dr. Leo Bodensteiner for collections in the North Cascades National Park. I also thank Joan Vandersypen and Dr. Robin Matthews at the Institute for Watershed Studies at Western Washington University for nutrient analyses. Dr. Matthews also provided multilevel hierarchical clustering models and Kendall's  $\tau$  rank-based correlations between species richness and abiotic factors.

#### LITERATURE CITED

- ABELL, R.A., D.M. OLSON, E. DINERSTEIN, P.T. HURLEY, J.T. DIGGS, W. EICHBAUM, S. WALTER, W. WETTENGEL, T. ALLNUTT, C.L. LOUCK, AND P. HEDAO. 2000. Freshwater ecoregions of North America: a conservation assessment. Inland Press, Washington, DC. 319 pp.
- [APHA] AMERICAN PUBLIC HEALTH ASSOCIATION. 2005. Standard methods for the examination of water and wastewater. 21st edition. American Public Health Association, American Water Works Association, and Water Environment Federation, Washington, DC.
- BAHLS, L.L. 2004–2006. Northwest diatoms: a photographic catalogue of species in the Montana Diatom Collection, with ecological optima, associates, and distribution records for nine northwestern United States. 3 volumes. Hannaea, Helena, MT.
- \_\_\_\_\_. 2009. A checklist of diatoms from inland waters of the northwestern United States. Proceedings of the Academy of Natural Sciences of Philadelphia 158: 1–35.
- BARBOUR, M.T., J.B. STRIBLING, AND J.R. KARR. 1995. Multi-metric approach for establishing biocriteria and measuring biological condition. Pages 63–77 in W.S. Davis and T.P. Simon, editors, Biological assessment and criteria: tools for water resource planning and decision making. Lewis Publishers, Boca Raton, FL.
- BEN-HUR, A., AND I. GUYON. 2003. Detecting stable clusters, using principal components analysis. Pages 159–182 in M.J. Brounsterin and A. Kohodursdy, editors, Methods in molecular biology. Humana Press, New York, NY.
- BENNETT, W.A.G. 1962. Saline lake deposits in Washington. Bulletin No. 49, Division of Mines and Geology. 129 pp.
- BLINN, D.W. 1993. Diatom community structure along physicochemical gradients in saline lakes. Ecology 74:1246–1263.
- BLINN, D.W., AND D.E. RUITER. 2006. Tolerance values of stream caddisflies (Trichoptera) in the Lower Colorado River Basin, USA. Southwestern Naturalist 51: 326–337.
- \_\_\_\_\_. 2013. Tolerance values and effects of selected environmental determinants of caddisfly (Trichoptera) distribution in northwest and north central Washington, USA. Western North American Naturalist 73: 270–294.
- COOKE, S.E., AND E.E. PREPAS. 1998. Stream phosphorus and nitrogen export from agricultural and forested watersheds on the Boreal Plain. Canadian Journal of Fisheries and Aquatic Sciences 10:2292–2299.
- CRAWLEY, M.J. 2007. The R book. Wiley Online Library. <http://dx.doi.org/10.1002/9780470515075>
- ENACHE, M.D., M. POTAPOVA, R. SHEIBLEY, AND P.W. MORAN. 2013. Three new *Psammothidium* species from lakes of Olympic and Cascade Mountains in Washington State, USA. Phytotaxa 127:49–57.
- ENGER, E., AND B. SMITH. 2009. Environmental science: a study of interrelationships. McGraw Hill, New York, NY. 512 pp.
- FORE, L.S., AND C. GRAFE. 2002. Using diatoms to assess the biological condition of large rivers in Idaho (U.S.A.). Freshwater Biology 47:2015–2037.
- GILLIS, C., AND M. CHALIFOUR. 2010. Changes in the macrobenthic community structure following the introduction of the invasive algae *Didymosphenia geminata*

- in the Matapedia River (Quebec, Canada). *Hydrobiologia* 647:63.
- HAWKINS, C.P., AND R.N. NORRIS. 2000. Performance of different landscape classifications for aquatic bioassessments: introduction to the series. *Journal of the North American Benthological Society* 19:367–369.
- HOLLINGSHEAD, K.D. 2012. Diatoms in Castor Lake (north-central Washington, USA): proxies of climate and hydrologic variation. Master's thesis, University of Western Ontario, London, Canada. 119 pp.
- JAMES, D.A., S.H. RANNEY, S.R. CHIPPS, AND B.D. SPINDLER. 2010. Invertebrate composition and abundance associated with *Didymosphenia geminata* in a montane stream. *Journal of Freshwater Ecology* 25: 235–241.
- JONGMAN, R.H.G., C.J.F. TER BRAAK, AND O.F.R. VAN TONGEREN, EDITORS. 1995. Data analysis in community and landscape ecology. Cambridge University Press. 212 pp.
- KAUFFMAN, J.B., AND W.C. KRUEGER. 1984. Livestock impacts on riparian ecosystems and streamside management implications: a review. *Journal of Range Management* 37:430–438.
- KINGSTON, J.C. 2004. Araphid and monoraphid diatoms. Pages 595–636 in J.D. Wehr and R.C. Sheath, editors, *Freshwater algae of North America: ecology and classification*. Elsevier Science (USA).
- KOCIOLEK, J.P. 2006. Some thoughts on the development of a diatom flora for freshwater ecosystems in the continental United States and a listing of recent taxa described from U.S. freshwaters. *Proceedings of the California Academy of Sciences* 57(21):561–586.
- KOCIOLEK, J.P., AND S.A. SPAULDING. 2003. Symmetrical naviculoid diatoms. Pages 637–653 in J.D. Wehr and R.C. Sheath, editors, *Freshwater algae of North America: ecology and classification*.
- KRAMMER, K., AND H. LANGE-BERTALOT. 1986. Bacillariophyceae. Teil 1. Naviculaceae. In: H. Ettl, J. Gerloff, H. Heynig, and D. Mollenhauer, editors, *Süßwasserflora von Mitteleuropa*. Band 2/1. Gustav Fischer Verlag, Stuttgart, Germany. 876 pp.
- \_\_\_\_\_. 1988. Bacillariophyceae. Teil 2. Bacillariaceae, Epithemiaceae, Surirellaceae. In: H. Ettl, J. Gerloff, H. Heynig, and D. Mollenhauer, editors, *Süßwasserflora von Mitteleuropa*. Band 2/2. Gustav Fischer, Stuttgart, Germany. 596 pp.
- \_\_\_\_\_. 1991a. Bacillariophyceae. Teil 3. Centrales, Fragilariaceae, Eunotiaceae. In: H. Ettl, J. Gerloff, H. Heynig, and D. Mollenhauer, editors, *Süßwasserflora von Mitteleuropa*, Band 2/3. Gustav Fischer Verlag, Stuttgart, Germany. 576 pp.
- \_\_\_\_\_. 1991b. Bacillariophyceae. Teil 4. Achnantheaceae. Kritische Ergänzungen zu *Navicula* und *Gomphonema*. In: H. Ettl, J. Gerloff, H. Heynig, and D. Mollenhauer, editors, *Süßwasserflora von Mitteleuropa*. Band 2/4. Gustav Fischer Verlag, Stuttgart, Jena, Germany. 437 pp.
- \_\_\_\_\_. 2000. Bacillariophyceae. Part 5. English and French translation of the keys. *Süßwasserflora von Mitteleuropa*. Spektrum Akademischer Verlag, Heidelberg/Berlin, Germany. 311 pp.
- KUMAR, S., S.A. SPAULDING, T.J. STOHLGREN, K.A. HERMANN, T.S. SCHMIDT, AND L.L. BAHLS. 2009. Potential habitat distribution for the freshwater diatom *Didymosphenia geminata* in the continental US. *Frontiers in Ecology and the Environment* 7: 415–420.
- LARNED, S.T., AND C. KILROY. 2014. Effects of *Didymosphenia geminata* removal on river macroinvertebrate communities. *Journal of Freshwater Ecology* 29:345–362.
- LAWS, E.A. 2000. *Aquatic pollution: an introductory text*. 3rd edition. John Wiley & Sons, New York, NY. 672 pp.
- LEE, C.J., AND A.C. ZIEGLER. 2010. Effects of urbanization, construction activity, management practices and impoundments on suspended-sediment transport in Johnson County, northeast Kansas, February 2006 through November 2008. U.S. Geological Survey Scientific Investigations Report 2010-5128. 54 pp.
- LOWE, R.L. 1974. Environmental requirements and pollution tolerance of freshwater diatoms. EPA-670/4-74-005. Cincinnati, OH.
- MACDONALD, L.H., A.W. SMART, AND R.C. WISSMAR. 1991. Monitoring guidelines to evaluate effects of forestry activities on streams in the Pacific Northwest and Alaska. EPA 910/9-91-001, U.S. Environmental Protection Agency, Seattle, WA. 166 pp.
- PLATTS, W.S., W.F. MEGAHAN, AND G.W. MINSHALL. 1983. Methods for evaluating stream, riparian, and biotic conditions. General Technical Report INT-138, USDA Forest Service, Odgen, UT.
- POTAPOVA, M., AND D.F. CHARLES. 2007. Diatom metrics for monitoring eutrophication in rivers of the United States. *Ecological Indicators* 7:48–70.
- R DEVELOPMENT CORE TEAM. 2011. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-9090051-07-0. <http://www.R-project.org>
- REID, B., AND R. TORRES. 2014. *Didymosphenia geminata* invasion in South America: ecosystem impacts and potential biogeochemical state change in Patagonian rivers. *Acta Oecologica* 54:101–109.
- ROSENFELD, C. 1985. Landforms and geology. Page 40 in A.J. Kimerling and P.L. Jackson, editors, *Atlas of the Pacific Northwest*.
- SHEIBLEY, R.W., M. ENACHE, P.W. SWARZENSKI, P.W. MORAN, AND J.R. FORMEMAN. 2014. Nitrogen deposition effects on diatom communities in lakes from three National Parks in Washington State. *Water, Air, and Soil Pollution* 225:1–23.
- SOVEREIGN, H.E. 1963. New and rare diatoms from Oregon and Washington. *Proceedings of the California Academy of Sciences* 31:349–368.
- SPAULDING, S.A., AND L. ELWELL. 2007. Increase in nuisance blooms and geographic expansion of the freshwater diatom *Didymosphenia geminata*. U.S. Geological Survey Open-File Report 2007-1425, U.S. Department of the Interior and U.S. Geological Survey. 38 pp.
- STEVENSON, R.J. 2014. Ecological assessments with algae: a review and synthesis. *Journal of Phycology* 50: 437–461.
- STEVENSON, R.J., Y. PAN, K.M. MANOYLOV, C.A. PARKER, D.P. LARSEN, AND A.T. HERLIHY. 2008. Development of diatom indicators of ecological conditions for streams of the western US. *Journal of the North American Benthological Society* 27:1000–1016.
- STOCKNER, J.G., AND W.W. BENSON. 1967. The succession of diatom assemblages in the recent sediments of Lake Washington. *Limnology and Oceanography* 12: 513–532.
- SUTHERLAND, S., M. RODWAY, C. KILROY, B. JARVIE, AND G. HUGHES. 2007. The survival of *Didymosphenia*

- geminata* in the rivers and associated spring-fed tributaries in the South Island of New Zealand. MAF Biosecurity New Zealand. 38 pp.
- TYNNI, R. 1986. Observations of diatom on the coast of the state of Washington. Geological Survey of Finland. Report of Investigation 75. 25 pp + 32 plates.
- UNITED STATES CENSUS BUREAU. 2013. State and country quick facts. U.S. Department of Commerce; [accessed 6 January 2013]. <http://quickfacts.census.gov>.
- VAN DAM, H., A. MERTENS, AND J. SINKELDAM. 1994. A coded checklist and ecological indicator values of freshwater diatoms from the Netherlands. Netherlands Journal of Aquatic Ecology 28:117–133.
- WALLACE, A.R., D.G. FRANK, AND A. FOUNIE. 2006. Freshwater diatomite deposits in the western United States. Fact Sheet 2006-3044, U.S. Geological Survey.

*Received 12 August 2014*  
*Accepted 25 December 2014*

Appendix 1 on page 30

Appendix 2 on page 36



APPENDIX 1. Site, location, altitude (ALTI), maximum water temperature (MTEM), total phosphorus (TP), total nitrogen diatom richness (RICH), and H' for 165 aquatic habitats in northwestern Washington, USA, including North Cascades listed in order of increasing altitude. Letters for 6 clusters in Fig. 1 are provided. Counties: CH = Chelan, CLA = Whatcom. NA = not available; NCNP = North Cascades National Park. Lakes and ponds are in boldface type. Woolley, WA. An asterisk (\*) indicates that *Didymosphenia geminata* was collected. (Data taken in part from Blinn and

Cluster letter	SITE	LOCATION	ALTI (m)	MTEM (°C)
*A	Nooksack River on Slater Rd. (WH)	N48.818168 W122.5803	3	20
A	California Cr.; Knickerville Rd. (WH)	N48.947139 W122.7044	4	27
C <sup>1</sup>	Squalicum Cr.; Bellingham (WH)	N48.765213 W122.5022	5	19
A	California Cr.; Valley View Rd. (WH)	N48.920695 W122.6601	7	19
A	Sammamish River (KIN)	N47.66036 W122.12372	7	17
A	Terrell Cr. on Jackson Rd. (WH)	N48.89690 W122.74884	10	19
A	Johnson Cr. in Sumas, WA (WH)	N48.99828 W122.26665	13	20
C <sup>1</sup>	Oyster Cr.; Chuckanut Drive (SK)	N48.61770 W122.43954	13	17
D	Hoko River on Hoko Ozette Rd. (CLA)	N48.25867 W124/35221	15	15
C <sup>1</sup>	Chuckanut Cr.; Fairhaven (WH)	N48.71569 W122.49484	15	19
A	Tenmile Cr. on Tenmile Rd. (WH)	N48.86994 W122.46700	22	19
A	Fourmile Cr. on Tenmile Rd. (WH)	N48.86980 W122.48189	22	21
*C <sup>1</sup>	Green Cr. on Hwy. 112 (CLA)	N48.17839 W124.20638	23	16
C <sup>1</sup>	Fever Cr.; Bellingham (WH)	N48.77181 W122.44167	24	18
*B	Dungeness River on Ward Rd. (CLA)	N48.11823 W123.14843	24	14
A	<b>Scudder Pond, Bellingham (WH)</b>	<b>N48.76162 W122.42091</b>	<b>25</b>	<b>25</b>
A	Elwa River on Elwa Rd. (CLA)	N48.11376 W123.55392	25	16
C <sup>1</sup>	Breckenridge Cr. Goodwin Rd. (WH)	N48.88900 W122.28790	25	19
*C <sup>1</sup>	White Cr. (Mi. 60.6) Hwy. 530 (SK)	N48.39765 W121.55106	99	19
C <sup>1</sup>	Whatcom Cr.; Bellingham (WH)	N48.75329 W122.43503	25	22
*A	Nooksack River at Everson (WH)	N48.91771 W122.34920	26	18
B	Snoqualamie River in Fall City (KIN)	N47.56819 W121.88284	26	15
C <sup>1</sup>	Unknown stream to Dakota Cr. (WH)	N48.96522 W122.66006	27	19
A	Fish Trap Cr. (Hwy. 546) (WH)	N48.96427 W122.43164	33	20
A	Sumas River on Lindsay Rd. (WH)	N48.94922 W122.30856	33	20
*A	Nooksack River at Cedarville (WH)	N48.84179 W122.29380	33	22
*B	Stilliquamish River on Strotz Rd. (SN)	N48.19859 W122.20468	34	16
C <sup>1</sup>	Bear Cr.; Cain Lk. Rd. (SK)	N48.62792 W122.36263	40	18
*B	Bogachiel River on Hwy. 101 (CLA)	N47.89416 W124.35709	44	17
B	Olney Cr. 9 Mi. 26.5 on Hwy. 2 (SN)	N47.87764 W121.71800	44	18
*A	Baker River in Concrete (SK)	N48.53707 W121.74216	48	19
D	Anderson Cr. on Oil City Rd. (JEF)	N47.77206 W124.32293	48	14
B	Swift Cr. bridge on Goodwin Rd. (WH)	N48.90595 W122.28746	50	21
D	Lost Cr. on Hwy. 101 (JEF)	N47.78170 W124.27018	52	15
*C <sup>2</sup>	Grandy Cr. (Mi. 82.7) Hwy. 20 (SK)	N48.53285 W121.88413	53	25
*D	Smith Cr. (Mi. 11.4) Hwy. 542 (WH)	N48.84145 W122.26037	54	16
*C <sup>1</sup>	Jackman Cr. (Mi. 91) Hwy. 20 (SK)	N48.53079 W121.71985	55	21
A	<b>Wiser Lake near Lynden (WH)</b>	<b>N48.90516 W122.48491</b>	<b>57</b>	<b>27</b>
*B	Calawah River on Hwy. 101 (CLA)	N47.95981 W124.39283	60	17
A	<b>Lake Terrell (WH)</b>	<b>N48.86054 W122.68965</b>	<b>65</b>	<b>28</b>
*B	Nooksack River (Hwy. 9 bridge) (WH)	N48.86054 W122.89650	69	16
*D	Bacon Cr. (Mi. 110.8) Hwy. 20 (SK)	N48.58796 W121.39524	70	16
A	<b>Lake Padden, Bellingham (WH)</b>	<b>N48.70138 W122.44709</b>	<b>74</b>	<b>20</b>
C <sup>1</sup>	Padden Cr.; Padden Cr. Park (WH)	N48.70277 W122.24465	75	18
A	<b>Sammish Lake, Bellingham WA (WH)</b>	<b>N48.67045 W122.38134</b>	<b>75</b>	<b>21</b>
*B	Skagit River at Rockport (SK)	N48.48464 W121.59299	76	16
D	Stream/waterfall at Mi. 245 on I-5 (WH)	N48.67373 W122.38669	80	13
C <sup>1</sup>	Samish River (old Hwy. 99) (SK)	N48.52577 W122.34008	82	16
*B	N. F. Stillaquamish Hwy. 530 (SN)	N48.27748 W121.80850	83	16
C <sup>1</sup>	Friday Cr.; Friday Cr. Rd. (SK)	N48.57403 W122.33828	90	19
A	<b>Lake Whatcom; Agate Bay (WH)</b>	<b>N48.75664 W122.35585</b>	<b>93</b>	<b>19</b>
*B	Sauk River (Mi. 61.6) Hwy. 530 (SK)	N48.40871 W121.55528	93	19
D	Unknown stream by Whatcom Lk. (WH)	N48.70716 W122.29792	96	15
*B	Skagit River at Marblemount WA (SK)	N48.52704 W121.42845	96	14
D	Carpenter Cr.; Bellingham (WH)	N48.75664 W122.35486	96	15
D	Wall Seep at Mi. 71.3 Hwy. 9 (WH)	N48.69127 W122.19295	98	16
*B	N. Fork Nooksack on Mosquito Rd. (WH)	N48.78476 W122.11269	98	15
C <sup>1</sup>	Anderson Cr. bridge on Hwy. 542 (WH)	N48.79027 W122.42083	103	18

(TN), specific conductance (SPCO), canopy cover (CAN), embeddedness (EMBE), suspended sediment (SUSE), National Park, Olympic Peninsula, and Mt. Baker–Snoqualmie, Okanogan, and Wenatchee National Forests. Sites are Clallam, JEF = Jefferson, KIN = King, KIT = Kittitas, OK = Okanogan, SK = Skagit, SN = Snohomish, WH = Information and collections by National Park Service and North Cascades National Park Service Complex; Sedro Ruiter 2013).

TP ( $\mu\text{g} \cdot \text{L}^{-1}$ )	TN ( $\mu\text{g} \cdot \text{L}^{-1}$ )	SPCO ( $\text{mS} \cdot \text{cm}^{-1}$ )	CAN (%)	EMBE (%)	SUSE ( $\text{mg} \cdot \text{L}^{-1}$ )	RICH	H'
58	190	0.110	5	95	16	53	4.68
30	2359	26.20	5	95	4	46	5.04
25	578	0.247	30	55	5	53	3.74
181	3238	0.381	95	95	2	38	4.54
17	437	0.105	10	90	2	69	4.76
209	1209	0.490	95	95	1	45	4.92
84	5615	0.264	85	85	5	53	4.25
4	1420	1.187	85	10	2	40	4.02
15	225	0.085	85	10	1	28	2.95
11	951	0.243	90	15	2	29	3.49
52	817	0.340	5	95	3	40	4.58
85	3157	0.386	5	95	5	37	4.37
15	329	0.087	75	35	1	35	3.94
19	527	0.262	60	40	1	36	3.48
6	94	0.102	25	20	1	45	4.45
99	787	<b>0.182</b>	<b>0</b>	<b>95</b>	1	<b>41</b>	<b>4.20</b>
112	121	0.084	20	75	279	14	2.21
19	401	0.244	90	15	3	42	3.85
5	66	0.060	80	15	1	57	3.56
8	440	0.226	20	20	4	57	3.56
64	138	0.123	5	90	14	25	4.06
23	432	0.052	5	85	3	24	3.47
30	1021	0.180	85	20	2	29	2.74
24	3429	0.303	45	85	5	39	3.32
54	974	0.360	5	95	12	31	3.40
20	105	0.080	5	80	156	38	3.33
6	188	0.045	5	25	10	24	3.48
4	609	0.102	80	25	2	33	3.19
4	108	0.091	20	25	2	34	3.44
26	221	0.038	10	45	35	42	4.27
25	110	0.045	5	85	15	55	3.42
14	235	0.068	90	15	1	28	3.51
10	390	0.240	5	95	4	14	1.78
17	88	0.093	95	20	1	14	0.93
11	217	0.200	70	45	5	27	3.40
7	275	0.099	85	35	1	17	1.72
5	158	0.090	60	40	5	23	2.91
<b>86</b>	<b>1633</b>	<b>0.341</b>	<b>0</b>	<b>95</b>	<b>1</b>	<b>33</b>	<b>3.38</b>
8	278	0.093	20	30	2	33	2.71
<b>36</b>	<b>1149</b>	<b>0.098</b>	<b>0</b>	<b>99</b>	<b>2</b>	<b>44</b>	<b>4.40</b>
20	105	0.080	5	95	15	30	2.96
4	68	0.040	50	20	1	29	3.45
<b>11</b>	<b>471</b>	<b>0.055</b>	<b>5</b>	<b>99</b>	<b>1</b>	<b>34</b>	<b>4.12</b>
10	638	0.183	95	10	1	42	4.63
<b>19</b>	<b>370</b>	<b>0.068</b>	<b>10</b>	<b>40</b>	<b>2</b>	<b>24</b>	<b>3.53</b>
16	35	0.088	50	10	10	10	3.77
7	436	0.061	90	30	1	25	2.65
9	757	0.128	60	40	4	37	3.77
4	123	0.068	5	15	5	24	3.48
8	572	0.135	80	15	2	64	5.06
<b>4</b>	<b>455</b>	<b>0.063</b>	<b>5</b>	<b>80</b>	<b>5</b>	<b>28</b>	<b>3.33</b>
28	31	0.050	5	20	150	34	3.17
4	830	0.099	85	80	1	18	2.48
7	71	0.040	10	90	5	46	4.05
22	660	0.119	95	25	1	27	2.72
31	363	0.085	75	20	1	25	2.95
7	64	0.095	10	65	14	29	3.77
18	400	0.126	85	20	1	38	3.89

## APPENDIX I. Continued.

Cluster letter	SITE	LOCATION	ALTI (m)	MTEM (°C)
*C <sup>2</sup>	Canyon Cr. on Mosquito Lk. Rd. (WH)	N48.83365 W122.13649	105	17
C <sup>2</sup>	Bell Cr. (Mi. 17.4) Hwy. 542 (WH)	N48.84871 W122.16121	107	17
*D	S. Fork Nooksack on Saxon Rd. (WH)	N48.67807 W122.16582	108	16
C <sup>1</sup>	Olsen Cr.; N. Shore Dr. Bellingham (WH)	N48.89768 W122.14376	114	20
D	Kendell Cr. Nooksack Hatchery (WH)	N48.89768 W122.14376	124	16
*B	Hoh River at Hoh Ox Box camp (JEF)	N47.81239 W124.25117	125	16
*C <sup>2</sup>	Rocky Cr. (Mi. 102.6) Hwy. 20 (SK)	N48.52644 W121.45381	133	18
*B	Squire Cr. at Squire Cr. campground (SN)	N48.27023 W121.67094	134	16
*D	Proctor Cr. (Mi. 31.2) Hwy. 2 (SN)	N47.83384 W121.64453	134	17
C <sup>1</sup>	Beaver Cr. on Hwy. 113 (CLA)	N48.11969 W124.21705	135	17
A	<b>Mirror Lake (WH)</b>	<b>N48.66279 W122.21809</b>	<b>135</b>	<b>22</b>
D	Fir Cr.; Sudden Valley (WH)	N48.67346 W122.26760	138	16
D	Brannian Cr.; Sudden Valley (WH)	N48.66903 W122.28041	139	16
D	Austin Cr.; Sudden Valley (WH)	N48.71297 W122.33139	147	17
*D	Porter Cr. on Mosquito Lk. Rd. (WH)	N48.79416 W122.11621	149	17
*B	Middle F Nooksack; Mosquito Rd. (WH)	N48.78483 W122.11246	152	16
*D	Damnation Cr. (Mi. 115) Hwy. 20 (SK)	N48.62717 W121.33821	157	16
A	<b>Silver Lake (WH)</b>	<b>N48.97137 W122.06909</b>	<b>157</b>	<b>25</b>
B	<b>Bog pond (Mi. 6.1); Mosquito Rd. (WH)</b>	<b>N48.78022 W122.11444</b>	<b>161</b>	<b>17</b>
C <sup>1</sup>	Maple Cr. (Mi. 26.1) Hwy. 542 (WH)	N48.92134 W122.07188	165	16
*B	N. Fork Nooksack (Mi. 27) (WH)	N48.92027 W122.06338	168	15
D	Hutchinson Cr.; Mosquito Rd. (WH)	N48.74097 W122.12128	174	15
C <sup>1</sup>	Saar Cr. on South Pass Rd. (WH)	N48.96306 W122.18544	183	18
*C <sup>2</sup>	Boulder Cr. (Mi. 26.6) Hwy. 542 (WH)	N48.92772 W122.03107	194	15
D	Stream on Mosquito Lk. Rd. (WH)	N48.74098 W122.12155	201	16
A	<b>Mosquito Lake Mosquito Lk. Rd. (WH)</b>	<b>N48.76881 W122.11814</b>	<b>203</b>	<b>17</b>
A	<b>Toad Lake, Bellingham WA (WH)</b>	<b>N48.79067 W122.39366</b>	<b>217</b>	<b>22</b>
*B	Sol Duc River near Fish Hatchery (CLA)	N48.06066 W124.12282	233	15
*A	Columbia River (CH)	N48.53549 W120.29815	248	21
*A	Okanogan River (OK)	N48.35345 W119.59373	260	23
*E	Cornell Cr. (Mi. 32.5) Hwy. 542 (WH)	N48.89208 W121.96208	263	15
D	Sulphur Cr. (Mi. 13.3) Baker Lk. Rd. (WH)	N48.65962 W121.71178	269	15
B	Glacier Cr. on Hwy. 542 (WH)	N48.88889 W121.94097	272	14
*D	Gallop Cr. on Hwy. 542 (WH)	N48.88921 W121.94331	274	16
D	Bear Cr. (Mi. 9.5) Baker Lk. Rd. (SK)	N48.62027 W121.74832	287	17
*B	N. Fork Nooksack Hwy. 542 (WH)	N48.90183 W121.91176	292	14
*B	Bechler River on FS 65 River RD (CH)	N47.72789 W121.33824	304	16
*D	Thompson Cr. on FS 39 (WH)	N48.87905 W121.91206	324	16
E	Wall seep (Mi. 125.1) Hwy. 20 (WH)	N48.69190 W121.22426	337	15
*C <sup>2</sup>	Stream (Mi. 123.5) Hwy. 20 (SK)	48°69085N,121°22611W	341	16
E	Stream/falls (Mi. 124) Hwy. 20 (WH)	48°69102N,121°22563W	353	16
*D	Falls Cr. on FS38 (WH)	N48.75477 W121.97300	369	15
D	Clearwater Cr. on FS 38 (WH)	N48.74640 W121.94625	373	15
*B	Cascade River 0.9 mi. on FS 1550 (SK)	N48.50134 W121.25027	377	16
D	Wall seep on FS Rd. 1550 (SK)	N48.50134 W121.25027	377	16
A	<b>Pond on Baker Lake Rd. (WH)</b>	<b>N48.72526 W121.73040</b>	<b>381</b>	<b>24</b>
*B	Skykomish River (Mi. 54.8) Hwy. 2 (KIN)	N47.71184 W121.31284	385	16
*C <sup>2</sup>	Mill Cr. Hwy. 97 (CH)	N47.51102 W120.63229	421	19
*C <sup>2</sup>	Wenatchee River (Mi. 96.5) Hwy. 2 (CH)	N47.59482 W120.71292	422	16
*C <sup>2</sup>	Monogram Cr. on Cascade Hwy. (SK)	N48.53565 W121.27194	429	9
D	N. Fork Nooksack River at Falls (WH)	N48.90561 W121.80847	458	15
*D	Trib near Nooksack Falls (WH)	N48.90657 W121.80682	469	16
*C <sup>2</sup>	Stream (Mi. 40.1) Hwy. 542 (WH)	N48.90785 W121.81145	474	15
*C <sup>2</sup>	Twisp River near Twisp (OK)	N48.36965 W120.14859	500	18
*C <sup>2</sup>	Wenatchee River (Mi. 90.5) Hwy. 2 (CH)	N47.76895 W120.80300	506	16
D	Talapus Cr.; NSF RD 9030 (KIN)	N47.39747 W121.53539	522	16
E	Stream (Mi. 40.9) Hwy. 542 (WH)	48°90793N;121°80636W	522	15
*C <sup>2</sup>	Chewuch River in Winthrop WA (OK)	N48.48010 W120.18144	537	16
C <sup>1</sup>	Beaver Cr. (Mi. 205.9) Hwy. 20 (OK)	N48.34903 W120.04261	541	18
E	Stream/falls (Mi. 41.6) Hwy. 542 (WH)	N48.90879 W121.80725	558	14
*C <sup>2</sup>	Confluence of Ruby & Granite Cr. (WH)	N48.70691 W120.91772	573	15
E	Stream (Mi. 42.8) Hwy. 542 (WH)	N48.91127 W121.79332	574	16

TP ( $\mu\text{g} \cdot \text{L}^{-1}$ )	TN ( $\mu\text{g} \cdot \text{L}^{-1}$ )	SPCO ( $\text{mS} \cdot \text{cm}^{-1}$ )	CAN (%)	EMBE (%)	SUSE ( $\text{mg} \cdot \text{L}^{-1}$ )	RICH	H'
4	73	0.100	70	15	1	23	2.84
5	255	0.065	80	10	2	16	2.37
5	120	0.073	20	20	5	36	4.16
16	460	0.137	75	20	2	34	3.87
7	586	0.203	25	80	2	38	4.75
17	77	0.070	5	40	2	31	3.52
4	79	0.100	75	35	1	24	3.10
4	92	0.032	85	15	1	29	2.36
15	145	0.016	90	5	8	25	2.96
14	239	0.087	35	25	1	58	4.14
<b>12</b>	<b>343</b>	<b>0.055</b>	<b>10</b>	<b>95</b>	<b>1</b>	<b>37</b>	<b>4.45</b>
18	956	0.069	85	55	1	37	4.45
14	1016	0.050	90	55	1	40	1.97
12	444	0.149	90	10	1	30	3.23
4	188	0.050	15	10	1	28	3.93
7	89	0.100	10	10	50	20	3.36
4	94	0.040	20	20	5	36	2.86
<b>9</b>	<b>225</b>	<b>0.144</b>	<b>5</b>	<b>80</b>	<b>1</b>	<b>42</b>	<b>4.40</b>
<b>48</b>	<b>551</b>	<b>0.127</b>	<b>15</b>	<b>95</b>	<b>5</b>	<b>37</b>	<b>3.49</b>
4	855	0.192	10	10	7	62	4.31
11	48	0.060	5	20	18	25	3.24
11	268	0.092	95	10	1	21	2.79
6	605	0.152	85	20	1	33	3.67
7	160	0.110	5	5	15	34	2.28
12	437	0.085	85	25	1	32	3.44
<b>48</b>	<b>551</b>	<b>0.128</b>	<b>5</b>	<b>95</b>	<b>1</b>	<b>29</b>	<b>2.70</b>
<b>9</b>	<b>934</b>	<b>0.114</b>	<b>10</b>	<b>80</b>	<b>2</b>	<b>47</b>	<b>4.37</b>
5	74	0.105	30	25	1	38	4.13
16	149	0.137	0	90	10	83	4.76
23	217	0.230	0	85	25	75	4.03
4	113	0.050	60	5	2	26	3.30
29	190	0.090	90	15	1	41	4.17
38	61	0.065	5	15	325	31	2.70
11	301	0.060	45	15	1	22	2.38
5	137	0.070	75	25	2	30	3.69
22	89	0.070	40	20	16	34	4.12
18	30	0.027	30	5	7	34	3.91
29	408	0.080	95	10	1	26	3.83
4	64	0.066	50	5	1	17	2.44
4	46	0.050	50	20	1	31	3.24
4	75	0.062	20	10	1	18	2.62
9	231	0.070	95	5	1	16	2.25
7	68	0.064	5	5	5	16	2.67
41	63	0.027	20	10	5	20	3.10
25	14	0.169	90	25	1	14	1.85
<b>22</b>	<b>646</b>	<b>0.058</b>	<b>20</b>	<b>90</b>	<b>1</b>	<b>25</b>	<b>3.63</b>
16	35	0.025	5	10	5	22	3.56
9	19	0.147	15	10	1	42	4.12
19	94	0.026	5	10	16	43	4.17
5	273	0.033	95	10	1	37	3.75
22	64	0.036	20	65	5	32	3.72
20	40	0.080	85	20	1	36	2.93
7	67	0.130	80	20	1	36	3.24
8	173	0.144	40	40	5	67	4.98
18	90	0.027	5	10	20	38	3.17
8	99	0.040	95	5	1	32	3.43
5	51	0.074	90	15	19	23	3.24
7	98	0.120	25	15	1	66	4.93
53	240	0.133	35	30	8	32	4.30
4	22	0.060	15	10	1	20	2.83
4	29	0.026	65	5	1	32	3.38
4	8	0.131	10	5	1	24	3.15

## APPENDIX 1. Continued.

Cluster letter	SITE	LOCATION	ALTI (m)	MTEM (°C)
*B	Chiwaukum Cr. (Mi. 89.9) Hwy. 2 (CH)	N47.72320 W120.73655	574	16
B	Skinny Cr. (Mi. 88.5) Hwy. 2 (CH)	N47.73061 W120.73897	574	17
*B	N. Fork Nooksack Hwy. 542 (WH)	N48.91236 W121.78424	592	14
*C <sup>2</sup>	Cle Elum River I-90 (KIT)	N47.18411 W121.00428	599	17
E	N. Fork Nooksack; Hannegan Pass (WH)	N48.90588 W121.69389	608	14
E	Swamp Cr.; Hannegan Pass Rd. (WH)	N48.90561 W121.68723	609	14
E	Stream (Mi. 46.2) Hwy. 542 (WH)	N48.91100 W121.76732	610	14
*C <sup>1</sup>	Teaway River (Hwy. 970) (KIT)	N47.19562 W120.78508	614	23
*C <sup>2</sup>	Big Cr. on Nelson Siding Rd. (KIT)	N47.20397 W121.11251	618	16
*C <sup>2</sup>	Yakima River (Mi. 79.1) I-90 (KIT)	N47.18580 W121.04352	620	17
C <sup>2</sup>	Stream 1.2 mi. on Hannegan Pass (WH)	N48.90304 W121.67	634	14
*C <sup>2</sup>	Nason Cr. (Mi. 81.4) Hwy. 2 (CH)	N47.76910 W120.80270	644	15
C <sup>2</sup>	Lower Goat Cr.; Goat Cr. Rd. (OK)	N48.58104 W120.37939	660	16
*C <sup>2</sup>	Methow River at Mazama WA (OK)	N48.59052 W120.40689	664	16
C <sup>1</sup>	Loup Loup Cr. (Mi. 222.2) Hwy. 20 (OK)	N48.36698 W119.72832	664	15
*C <sup>2</sup>	Early Winter Cr. Hwy. 20 (OK)	N48.59790 W120.44338	669	15
*E	Varden Cr. (Mi. 173.1) Hwy. 20 (OK)	N48.58836 W120.49825	675	14
*C <sup>2</sup>	Upper Goat Cr. (FS 52) (OK)	N48.59038 W120.37012	686	15
*C <sup>2</sup>	Boulder Cr. on FS37 near Winthrop (OK)	N48.58300 W120.14850	694	16
C <sup>2-</sup>	Stream (Mi. 48.1) Hwy. 542 (WH)	N48.88998 W121.67530	712	15
B	Barr Cr. (Mi. 4.4) FS33 (WH)	N48.86704; W121.76761	715	13
E	Wells Cr. (Mi. 4.4) FS33 (WH)	N48.86718; W121.76630	718	13
E	Wall seep; Wells Cr. (Mi. 4.4) F33 (WH)	N48.86736; W121.76635	729	13
E	Wall seep (Mi. 47.6) Hwy. 542 (WH)	N48.88854; W121.67356	734	15
E	Wall seep (Mi. 48.6) Hwy. 542 (WH)	N48.88758 W121.67236	741	15
E	Stream/falls (Mi. 144.3) Hwy. 20 (SK)	N48.68110 W120.88167	763	14
E	Bagley Cr. (Mi. 49.1) Hwy. 542 (WH)	N48.87708 W121.67064	822	14
C <sup>1</sup>	Frazer Cr. (Mi. 210.3) Hwy. 20 (OK)	N48.36466 W120.01267	848	15
E	Lower Galena Cr. Hwy. 542 (WH)	N48.87028 W121.6651	921	14
E	Commonwealth Cr. (KIT)	N47.42945 W121.42017	922	16
E	Ruth Cr. at Ruth Cr. campground (WH)	N48.90943 W121.59166	934	13
C <sup>2</sup>	Swauk Cr. (Mi. 159.7) Hwy. 97 (KIT)	N47.24250 W120.69739	944	15
E	Wall seep (Mi. 52.1) Hwy. 542 (WH)	N48.85164 W121.68526	1051	14
E	Jack Cr. (Mi. 213.9) Hwy. 20 (OK)	N48.36783 W120.00336	1117	14
<b>B</b>	<b>Pond (Mi. 53.1) Hwy. 542 (WH)</b>	<b>48°51953N, 121°39965W</b>	<b>1167</b>	<b>14</b>
E	Wall seep (Mi. 54) Hwy. 542 (WH)	N48.86222 W121.67215	1229	14
B	Highwood Lake; Mt. Baker (WH)	N48.86513 W121.67473	1238	21
E	Cutthroat Cr. Mi. 167 Hwy. 20 bridge (OK)	N48.54965 W120.66954	1242	12
<b>B</b>	<b>Picture Lake; Mt. Baker (WH)</b>	<b>N48.86565 W121.67627</b>	<b>1317</b>	<b>19</b>
E	Stream on Fire & Ice trail (WH)	N48.85586 W121.68795	1329	13
E	Stream near Heather Info Center (WH)	N48.85507 W121.68295	1343	16
<b>B</b>	<b>Terminal Lake; Mt. Baker (WH)</b>	<b>N48.85315, 121°68473W</b>	<b>1344</b>	<b>18</b>
E	Wall seep at Artist Point Hwy. (WH)	N48.85177 W121.68473	1381	13
E	Early Winter Cr. Headwaters (WH)	N48.52873 W120.64003	1410	13
E	Bridge Cr. (Mi. 158.7) Hwy. 20 (CH)	N48°30.29 W120°43.16	1414	13

TP ( $\mu\text{g} \cdot \text{L}^{-1}$ )	TN ( $\mu\text{g} \cdot \text{L}^{-1}$ )	SPCO ( $\text{mS} \cdot \text{cm}^{-1}$ )	CAN (%)	EMBE (%)	SUSE ( $\text{mg} \cdot \text{L}^{-1}$ )	RICH	H'
29	157	0.029	45	5	1	37	2.90
5	101	0.208	0	35	16	28	4.26
29	67	0.033	10	15	15	37	4.27
10	61	0.051	5	10	5	39	4.44
20	53	0.029	5	5	1	26	4.00
4	55	0.076	10	35	5	19	3.38
6	10	0.141	5	5	1	20	3.11
51	207	0.162	5	20	5	41	1.98
16	35	0.090	90	20	1	33	4.11
16	108	0.065	35	10	10	57	4.20
4	46	0.062	5	5	1	23	2.79
15	35	0.038	25	10	5	51	4.28
5	25	0.099	10	5	5	40	4.58
4	28	0.090	25	5	5	44	4.15
61	166	0.188	30	15	1	49	3.47
4	29	0.070	10	5	5	57	4.09
4	17	0.042	5	5	1	27	3.12
5	20	0.061	5	5	1	28	4.31
6	78	0.080	10	5	1	47	3.92
4	19	0.066	75	5	1	38	3.80
12	83	0.147	0	15	785	15	2.56
9	44	0.060	5	10	1	26	2.46
4	83	0.205	75	20	1	30	2.99
4	83	0.104	5	5	1	19	2.53
4	38	0.168	20	10	1	13	1.57
4	51	0.034	20	5	1	27	3.13
8	106	0.033	95	5	1	30	3.11
51	180	0.161	95	15	3	63	4.74
4	56	0.023	80	20	1	23	3.20
21	123	0.283	75	20	1	33	3.29
4	91	0.022	75	5	1	20	3.19
38	28	0.246	95	25	5	43	3.99
8	73	0.034	50	75	1	11	2.13
30	194	0.108	95	5	1	29	3.22
<b>20</b>	<b>248</b>	<b>0.152</b>	<b>10</b>	<b>90</b>	<b>1</b>	<b>36</b>	<b>4.53</b>
5	117	0.016	25	5	1	21	2.47
15	163	0.037	0	95	1	31	3.90
4	25	0.010	90	5	1	24	3.11
8	<b>164</b>	<b>0.062</b>	<b>0</b>	<b>90</b>	<b>1</b>	<b>36</b>	<b>3.95</b>
11	30	0.025	0	5	1	37	4.00
4	0.8	0.006	0	5	1	21	3.00
<b>13</b>	<b>130</b>	<b>0.013</b>	<b>0</b>	<b>75</b>	<b>NA</b>	<b>31</b>	<b>3.83</b>
10	21	0.004	0	5	0	12	1.99
8	19	0.009	0	5	0	15	3.15
4	42	0.030	10	5	1	26	3.43

APPENDIX 2. Diatom species collected in northwestern Washington, including the Olympic Peninsula. Total = 415 species; 352 river/stream; 213 pond and lake.

Species	Stream/river	Lake and pond
<i>Achnanthes grischuna</i> Wuthrich	x	
<i>Achnanthes nitidiformis</i> Lange-Bertalot		x
<i>Achnanthes rupestroides</i> Hohn		x
<i>Achnanthes rupestris</i> Krasske	x	
<i>Achnanthes stolidia</i> Krasske		x
<i>Achnanthes subsalsa</i> Petersen	x	
<i>Achnanthidium affine</i> (Grun.) Czar.	x	
<i>Achnanthidium biasolettianum</i> (Grun.) Round & Bukht.	x	x
<i>Achnanthidium exiguum</i> (Grun.) Czar.	x	
<i>Achnanthidium exile</i> (Kütz.) Heiberg	x	
<i>Achnanthidium gracillimum</i> (Meister) Lange-Bertalot		x
<i>Achnanthidium jackii</i> Rabh.	x	
<i>Achnanthidium minutissimum</i> (Kütz.) Czar.		x
<i>Adlafia bryophila</i> (Petersen) Moser, Lange-Bertalot & Metzeltin		x
<i>Adlafia miniscula</i> (Grun.) Lange-Bertalot	x	
<i>Adlafia miniscula</i> v. <i>muralis</i> (Grun.) Lange-Bertalot	x	
<i>Amphipleura lindheimeri</i> Grun.	x	
<i>Amphipleura pellucida</i> (Kütz.) Kütz.	x	
<i>Amphora ovalis</i> (Kütz.) Kütz.		x
<i>Amphora pediculus</i> (Kütz.) Grun. ex A. Schmidt		x
<i>Anomoeoneis brachysira</i> (Bréb) Grun.		x
<i>Anomoeoneis sphaerophora</i> E. Pfitzer	x	
<i>Anomoeoneis styriaca</i> (Grun.) Hust.	x	
<i>Anomoeoneis vitrea</i> (Grun.) Ross		x
<i>Asterionella formosa</i> Hass.	x	
<i>Aulacoseira alpigina</i> (Grun.) Krammer	x	x
<i>Aulacoseira ambigua</i> (Grun.) Simonsen		x
<i>Aulacoseira distans</i> (Ehr.) Simonsen	x	x
<i>Aulacoseira granulata</i> (Ehr.) Simonsen	x	x
<i>Aulacoseira islandica</i> (Müller) Simonsen	x	x
<i>Aulacoseira italica</i> (Ehr.) Simonsen	x	
<i>Aulacoseira italica</i> v. <i>tenuissima</i> (Grun.) Simonsen	x	
<i>Aulacoseira valida</i> (Grun.) Krammer		x
<i>Bacillaria paradoxa</i> Gmelin		x
<i>Biddulphia laevis</i> Ehr.	x	
<i>Brachysira brebissonii</i> Ross	x	
<i>Caloneis alpestris</i> (Grun.) Cl.	x	
<i>Caloneis bacillum</i> (Grun.) Cl.	x	x
<i>Caloneis latiuscula</i> (Kütz.) Cl.		x
<i>Caloneis molaris</i> (Grun.) Krammer	x	
<i>Caloneis permagna</i> (Bailey) Cl.	x	
<i>Caloneis pulchra</i> Messikommer	x	
<i>Caloneis schumanniana</i> (Grun.) Cl.	x	
<i>Caloneis silicula</i> (Ehr.) Cl.	x	x
<i>Caloneis sublinearis</i> (Grun.) Krammer	x	
<i>Caloneis tenuis</i> (Greg.) Krammer	x	
<i>Caloneis undulata</i> (Greg.) Krammer	x	
<i>Caloneis westii</i> (W. Sm.) Hendy	x	
<i>Campylodiscus clypeus</i> (Ehr.) Ehr. ex Kütz.	x	
<i>Cocconeis neodiminuta</i> Krammer	x	
<i>Cocconeis pediculus</i> Ehr.	x	
<i>Cocconeis placentula</i> Ehr.	x	x
<i>Cocconeis placentula</i> v. <i>euglypta</i> (Ehr.) Grun.	x	
<i>Cocconeis placentula</i> f. <i>lineata</i> (Ehr.) Van Heurck	x	
<i>Cocconeis scutellum</i> Ehr.	x	
<i>Craticula cuspidata</i> (Kütz.) Mann		x
<i>Craticula halophila</i> (Grun.) Mann		x
<i>Ctenophora pulchella</i> (Ralfs. ex Kütz.) Williams & Round		x
<i>Cyclotella atomus</i> Hust.		x
<i>Cyclotella distinguenda</i> Hust.	x	
<i>Cyclotella hakanssoniae</i> Wendker	x	
<i>Cyclotella menghiniana</i> Kütz.	x	x

## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Cyclotella michiganana</i> Skvortzow		x
<i>Cyclotella planctonica</i> Brunthaler	x	
<i>Cymatopleura solea</i> (Bréb.) W. Sm.	x	
<i>Cymbella aequalis</i> W. Sm.		x
<i>Cymbella affinis</i> Kütz.	x	x
<i>Cymbella aspera</i> (Ehr.) Cl.	x	x
<i>Cymbella austriaca</i> Grun.	x	x
<i>Cymbella caespitosa</i> (Kütz.) Brun		x
<i>Cymbella cesatii</i> (Rabh.) Grun.	x	
<i>Cymbella cistula</i> (Hemprich & Ehr.) Kirchner	x	x
<i>Cymbella cuspidata</i> Kütz.	x	
<i>Cymbella cymbiformis</i> C. Ag.	x	x
<i>Cymbella cymbiformis</i> v. <i>nonpunctata</i> Fontell	x	
<i>Cymbella delicatula</i> (Kütz.) Krammer	x	
<i>Cymbella ehrenbergii</i> Kütz.	x	
<i>Cymbella hauckii</i> Van Heurck	x	
<i>Cymbella helvetica</i> Kütz.	x	x
<i>Cymbella hungarica</i> (Grun.) Pantocsek	x	
<i>Cymbella hybrida</i> Grun. ex Cl.		x
<i>Cymbella incerta</i> (Grun.) Krammer		x
<i>Cymbella incerta</i> v. <i>crassipunctata</i> Krammer		x
<i>Cymbella janischii</i> (A. Schmidt) De Toni.	x	
<i>Cymbella laevis</i> Nägeli	x	x
<i>Cymbella lata</i> Grun. ex Cl.	x	
<i>Cymbella mesiana</i> Cholnoky	x	
<i>Cymbella mexicana</i> (Ehr.) Cl.	x	
<i>Cymbella naviculiformis</i> (Auerswald) Cl.	x	x
<i>Cymbella norvegica</i> Grun.	x	
<i>Cymbella proxima</i> Reimer		x
<i>Cymbella rainierensis</i> Sov.	x	
<i>Cymbella reinhardtii</i> Grun.	x	x
<i>Cymbella schimanskii</i> Krammer	x	
<i>Cymbella tumida</i> (Bréb.) Van Heurck	x	
<i>Cymbopleura amphicephala</i> (Nägeli) Krammer		x
<i>Cymbopleura naviculiformis</i> (Auerswald ex Heiberg) Krammer	x	
<i>Decussata placenta</i> (Ehr.) Lange-Bertalot	x	
<i>Denticula elegans</i> Kütz.	x	x
<i>Denticula kuetzingii</i> Grun.	x	
<i>Denticula subtilis</i> Grun.	x	
<i>Diatoma anceps</i> (Ehr.) Kirchner	x	x
<i>Diatoma hyemalis</i> (Roth) Heiberg	x	
<i>Diatoma mesodon</i> (Ehr.) Kütz.	x	x
<i>Diatoma tenue</i> C. Ag.	x	
<i>Diatoma vulgare</i> Bory	x	x
<i>Diatomella balfouriana</i> Grev.	x	
<i>Didymosphaenia geminata</i> (Lyngbye) M. Schmidt	x	x
<i>Diploneis elliptica</i> (Kütz.) Cl.	x	x
<i>Diploneis modica</i> Hust.		x
<i>Diploneis oblongella</i> (Nägeli ex Kütz.) CL-EU	x	
<i>Diploneis ovalis</i> (Hilse) Cl.	x	x
<i>Diploneis pseudovalis</i> Hust.	x	
<i>Diploneis puella</i> (Schumann) Cl.	x	
<i>Discotella stelligera</i> (Cl. & Grun.) Houk & Klea	x	x
<i>Encyonema gracile</i> Rabh.	x	x
<i>Encyonema minutum</i> (Hilse) Mann	x	x
<i>Encyonema muelleri</i> Hust.	x	
<i>Encyonema prostratum</i> (Berkeley) Kütz.	x	
<i>Encyonema silesiacum</i> (Bleisch) Mann	x	x
<i>Encyonopsis cesatii</i> (Rabh.) Krammer	x	x
<i>Encyonopsis descripta</i> (Hust.) Krammer		x
<i>Encyonopsis falaisensis</i> (Grun.) Krammer	x	
<i>Encyonopsis microcephala</i> (Grun.) Krammer		x
<i>Entomoneis alata</i> (Ehr.) Reimer	x	



## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Entomoneis paludosa</i> (W. Sm.) Reimer	x	
<i>Epithemia adnata</i> (Kütz.) Bréb.	x	x
<i>Epithemia alpestris</i> (W. Sm.)	x	
<i>Epithemia argus</i> (Ehr.) Kütz.	x	x
<i>Epithemia sorex</i> Kütz.	x	x
<i>Epithemia turgida</i> (Ehr.) Kütz.	x	x
<i>Epithemia turgida</i> v. <i>granulata</i> (Ehr.) Brun	x	x
<i>Epithemia turgida</i> v. <i>westermanni</i> (Ehr.) Grun.	x	x
<i>Eucocconeis flexella</i> (Kütz.) Cl.	x	
<i>Eucocconeis laevis</i> (Østrup) Lange-Bertalot	x	
<i>Eumotia arcus</i> Ehr.	x	x
<i>Eumotia bilunaris</i> (Ehr.) Schaarschmidt	x	x
<i>Eumotia bilunaris</i> v. <i>mucophila</i> (Lange-Bertalot & Norpel-Schempp) Lange-Bertalot	x	
<i>Eumotia diodon</i> Ehr.	x	x
<i>Eumotia exigua</i> (Bréb.) Rabh.	x	
<i>Eumotia formica</i> (Ehr.)	x	
<i>Eumotia implicata</i> Nörpel, Lange-Bertalot & Alles	x	x
<i>Eumotia incisa</i> Smith ex Greg.	x	x
<i>Eumotia monodon</i> Ehr.	x	
<i>Eumotia muscicola</i> Krasske	x	
<i>Eumotia muscicola</i> v. <i>tridentula</i> Nörpel & Lange-Bertalot	x	
<i>Eumotia pectinalis</i> (Kütz.) Rabh.	x	x
<i>Eumotia pectinalis</i> v. <i>undulata</i> (Ralfs.) Rabh.	x	x
<i>Eumotia praerupta</i> Ehr.	x	
<i>Eumotia serra</i> v. <i>diadema</i> (Ehr.) Patrick		x
<i>Eumotia serra</i> v. <i>tetraodon</i> (Ehr.) CL-EU	x	
<i>Eumotia soleirolii</i> (Kütz.) Rabh.	x	
<i>Fallacia pygmaea</i> (Kütz.) Stickle & Mann	x	
<i>Fragilaria capucina</i> Desm.	x	x
<i>Fragilaria constricta</i> Ehr.	x	
<i>Fragilaria crotonensis</i> Kitton	x	x
<i>Fragilaria delicatissima</i> (W.Sm.) Lange-Bertalot	x	
<i>Fragilaria famolica</i> (Kütz.) Lange-Bertalot		x
<i>Fragilaria mazamaensis</i> (Sov.) Lange-Bertalot	x	
<i>Fragilaria nanana</i> Lange-Bertalot		x
<i>Fragilaria pseudoconstruens</i> Marciniak	x	
<i>Fragilaria tenera</i> (W. Sm.) Lange-Bertalot	x	x
<i>Fragilaria vaucheriae</i> (Kütz.) Perersen	x	x
<i>Fragilaria virescens</i> (Ralfs) Williams & Round	x	
<i>Frustulia crassinervia</i> (Bréb.) Lange-Bertalot & Krammer	x	x
<i>Frustulia krammeri</i> Lange-Bertalot & Metzeltin	x	x
<i>Frustulia saxonica</i> Rabh.		x
<i>Frustulia spicula</i> Amossé		x
<i>Frustulia vulgaris</i> (Thwaites) De Toni	x	x
<i>Geisslera decussis</i> (Østrup) Lange-Bertalot & Metzeltin	x	
<i>Geisslera ignota</i> (Krasske) Lange-Bertalot & Metzeltin	x	
<i>Geisslera schoenfeldii</i> (Hust.) Lange-Bertalot & Metzeltin	x	x
<i>Gomphoneis erianse</i> v. <i>varabilis</i> Kociolek & Stoermer	x	
<i>Gomphoneis herculeana</i> (Ehr.) Cl.	x	
<i>Gomphoneis minuta</i> (Stone) Kociolek and Stoermer	x	
<i>Gomphonema accuminatum</i> Ehr.	x	x
<i>Gomphonema affine</i> Kütz.	x	x
<i>Gomphonema angustatum</i> (Kütz.) Rabh.	x	x
<i>Gomphonema angustum</i> C. Ag.	x	
<i>Gomphonema auger</i> v. <i>sphaerophorum</i> (Ehr.) Grun.	x	
<i>Gomphonema clavatum</i> Ehr.	x	x
<i>Gomphonema clevei</i> Fricke	x	x
<i>Gomphonema gracile</i> Ehr.	x	
<i>Gomphonema hebridense</i> W. Greg.	x	
<i>Gomphonema kobayashii</i> Kociolek & Kingston	x	
<i>Gomphonema minutum</i> (C. Ag.) C. Ag.	x	x
<i>Gomphonema olivaceum</i> (Hornemann) Ehr.	x	

## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Gomphonema olivaceum</i> f. <i>fonticola</i> Hust.	x	
<i>Gomphonema parvulum</i> (Kütz.) Kütz.	x	x
<i>Gomphonema subtile</i> Ehr.	x	x
<i>Gomphonema tenellum</i> Kütz.	x	
<i>Gomphonema truncatum</i> Ehr.	x	x
<i>Gomphonema truncatum</i> v. <i>turgidum</i> (Ehr.) Patr.	x	
<i>Gyrosigma acuminatum</i> (Kütz.) Rabh.		x
<i>Gyrosigma attenuatum</i> (Kütz.) Rabh.	x	
<i>Gyrosigma spencerii</i> (W. Sm) Griffith & Henfrey	x	
<i>Halamphora coffeaeformis</i> (Ag.) Levkov.	x	
<i>Halamphora veneta</i> (Kütz.) Levkov		x
<i>Hannaea arcus</i> (Ehr.) Patr.	x	x
<i>Hantzschia amphioxys</i> (Ehr.) Grun.	x	
<i>Hippodonta capitata</i> (Ehr.) Lange-Bertalot, Metzeltin & Witkowski	x	
<i>Karayevia oblongella</i> (Østrup) M. Aboal		x
<i>Karayevia suchlandtii</i> (Hust.) Bukhtiyarova	x	x
<i>Lemnicola hungarica</i> (Grun.) Round & Basson	x	
<i>Luticola cohnii</i> (Hilse) Mann	x	
<i>Luticola goeppertiana</i> (Bleisch) Mann	x	x
<i>Luticola mutica</i> (Kütz.) Mann	x	
<i>Mastogloia braunii</i> Grun.	x	
<i>Mastogloia pumila</i> (Cl. & Möller, Grun.) Cl.	x	
<i>Melosira varians</i> C. Ag.	x	
<i>Meridion circulare</i> (Grev.) C. Ag.	x	x
<i>Meridion circulare</i> v. <i>constrictum</i> (Ralfs) Van Heurck	x	
<i>Navicula aboensis</i> (Cl.) Cl.	x	
<i>Navicula abiskoensis</i> Hust.	x	
<i>Navicula angusta</i> Grun.	x	x
<i>Navicula cari</i> Ehr.	x	
<i>Navicula cincta</i> (Ehr.) Ralfs.	x	x
<i>Navicula cocconeiformis</i> Greg. ex Greville		x
<i>Navicula concentrica</i> Carter & Bailey-Watts		x
<i>Navicula crucicula</i> (W. Sm.) Donkin	x	
<i>Navicula crucicula</i> v. <i>cruciculoides</i> (Brockman) Lange-Bertalot	x	
<i>Navicula cryptocephala</i> Kütz.	x	x
<i>Navicula cryptonella</i> Lange-Bertalot	x	
<i>Navicula dentata</i> Hust.	x	x
<i>Navicula expecta</i> VanLandingham	x	
<i>Navicula gottlandica</i> Grun.	x	x
<i>Navicula gregaria</i> Donkin	x	x
<i>Navicula hambergii</i> Hust.	x	
<i>Navicula hasta</i> Pantocsek	x	
<i>Navicula heimansii</i> Van Dam & Kooyman	x	x
<i>Navicula insociabilis</i> Krasske	x	
<i>Navicula jaernefeltii</i> Hust.		x
<i>Navicula lacustris</i> W. Greg.	x	
<i>Navicula lanceolata</i> Ehr.	x	
<i>Navicula leptostriata</i> Jørgensen		x
<i>Navicula libonensis</i> Schoeman	x	
<i>Navicula margalithii</i> Lange-Bertalot	x	
<i>Navicula menisculus</i> Schumann	x	x
<i>Navicula molestiformis</i> Hust.	x	
<i>Navicula ordinaria</i> Hust.		x
<i>Navicula peregrina</i> (Ehr.) Kütz.	x	
<i>Navicula phyllepta</i> Kütz.	x	
<i>Navicula placentula</i> (Ehr.) Grun.	x	
<i>Navicula pseudoscutiformis</i> Hust.	x	x
<i>Navicula pseudoventralis</i> Hust.		x
<i>Navicula pusilla</i> v. <i>incognita</i> (Krasske) Lange-Bertalot	x	
<i>Navicula pusio</i> Cl.		x
<i>Navicula radiosa</i> Kütz.	x	x
<i>Navicula recens</i> (Lange-Bertalot) Lange-Bertalot	x	x
<i>Navicula reinhardtii</i> (Grun.) Grun.	x	

## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Navicula rhynchocephala</i> Kütz.	x	x
<i>Navicula rostellata</i> Kütz.		x
<i>Navicula saxophila</i> W. Bock ex Hust.	x	
<i>Navicula schroeteri</i> Meister	x	
<i>Navicula scutiformis</i> Grun. ex A. Schmidt		x
<i>Navicula semen</i> Ehr.	x	
<i>Navicula similis</i> Krasske		x
<i>Navicula striolata</i> (Grun.) Lange-Bertalot	x	
<i>Navicula stroemii</i> Hust.	x	x
<i>Navicula subminuscula</i> Manguin		x
<i>Navicula submuralis</i> Hust.	x	
<i>Navicula subrhynchocephala</i> Hust.	x	
<i>Navicula tenelloides</i> Hust.	x	
<i>Navicula tridentula</i> Krasske		x
<i>Navicula tripunctata</i> (Müller) Bory	x	
<i>Navicula variostrata</i> Krasske	x	
<i>Navicula veneta</i> Kütz.	x	x
<i>Navicula viridula</i> (Kütz.) Kütz.	x	
<i>Navicula culpina</i> Kütz.	x	x
<i>Neidium affine</i> (Ehr.) Pfitzer	x	
<i>Neidium affine</i> v. <i>longiceps</i> (Greg.) Cl.	x	x
<i>Neidium alpinum</i> Hust.		x
<i>Neidium ampliatum</i> (Ehr.) Krammer		x
<i>Neidium bisulcatum</i> (Lagerstedt) Cl.	x	x
<i>Neidium dubium</i> (Ehr.) Cl.	x	
<i>Neidium iridis</i> (Ehr.) Cl.	x	x
<i>Neidium productum</i> (Sm.) Cl.	x	
<i>Nitzschia acicularis</i> (Kütz.) W. Sm.	x	x
<i>Nitzschia acuminata</i> (W. Sm.) Grun.	x	
<i>Nitzschia alpina</i> Hust.	x	x
<i>Nitzschia amphibia</i> Grun.	x	x
<i>Nitzschia angustata</i> (W. Sm.) Grun.	x	x
<i>Nitzschia clausii</i> Hantz.	x	
<i>Nitzschia communis</i> Rabh.	x	x
<i>Nitzschia commutata</i> Grun.	x	
<i>Nitzschia desortorum</i> Hust.		x
<i>Nitzschia dissipata</i> (Kütz.) Grun.	x	x
<i>Nitzschia dissipata</i> v. <i>media</i> (Hantz.) Grun.	x	
<i>Nitzschia dubia</i> W. Sm.	x	
<i>Nitzschia filiformis</i> (W. Sm.) Hust.	x	x
<i>Nitzschia flexa</i> Schumann	x	
<i>Nitzschia flexioides</i> Geitler	x	
<i>Nitzschia fonticola</i> (Grtun.) Grun.	x	
<i>Nitzschia frustulum</i> (Kütz.) Grun.	x	x
<i>Nitzschia hantzschiana</i> Rabh.	x	
<i>Nitzschia heufleriana</i> Grun.	x	
<i>Nitzschia hybrida</i> Grun.	x	
<i>Nitzschia inconspicua</i> Grun.	x	
<i>Nitzschia intermedia</i> Hantz. ex Cl. & Grun.		x
<i>Nitzschia linearis</i> West	x	x
<i>Nitzschia linearis</i> v. <i>tenuis</i> (W. Sm.) Grun.	x	
<i>Nitzschia microcephala</i> Grun.	x	
<i>Nitzschia palea</i> (Kütz.) W. Sm.	x	x
<i>Nitzschia paleacea</i> Grun.	x	x
<i>Nitzschia pellucida</i> Grun.	x	
<i>Nitzschia perminuta</i> (Grun.) M. Peragallo	x	x
<i>Nitzschia plana</i> W. Sm.	x	
<i>Nitzschia pusilla</i> Grun.	x	
<i>Nitzschia sigma</i> (Kütz.) W. Sm.	x	
<i>Nitzschia sigmoidea</i> (Nitzsch) W. Sm.	x	
<i>Nitzschia sublinearis</i> Hust.	x	
<i>Nitzschia umbonata</i> (Ehr.) Lange-Bertalot	x	
<i>Nitzschia vermicularis</i> (Kütz.) Hantz.	x	x

## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Orthoseira dendroteres</i> (Ehr.) Round, Crawford, Mann	x	
<i>Orthoseira roeseana</i> (Rabh.) O'Meara	x	x
<i>Pinnularia appendiculata</i> (C. Ag.) Cl.		x
<i>Pinnularia borealis</i> Ehr.	x	x
<i>Pinnularia brevicostata</i> Cl.	x	
<i>Pinnularia dactylus</i> Ehr.	x	
<i>Pinnularia divergens</i> W. Sm.	x	x
<i>Pinnularia gibba</i> Ehr.	x	x
<i>Pinnularia gibba</i> v. <i>linearis</i> Hust.	x	
<i>Pinnularia hemiptera</i> (Kütz.) Cl.		x
<i>Pinnularia intermedia</i> (Lagerstedt) Cl.		x
<i>Pinnularia interrupta</i> W. Sm.	x	x
<i>Pinnularia karelica</i> Cl.	x	
<i>Pinnularia lata</i> (Bréb.) W. Sm.	x	x
<i>Pinnularia maior</i> (Kütz.) Cl.	x	
<i>Pinnularia microstauron</i> (Ehr.) Cl.	x	x
<i>Pinnularia nobilis</i> (Ehr.) Ehr.	x	
<i>Pinnularia obscura</i> Krasske	x	x
<i>Pinnularia stomatophora</i> (Grun.) Cl.	x	
<i>Pinnularia streptoraphe</i> Cl.	x	x
<i>Pinnularia subcapitata</i> W. Greg.	x	x
<i>Pinnularia subrostrata</i> Lohman & Andrews	x	
<i>Pinnularia sudetica</i> (Hilse) Peragallo	x	
<i>Pinnularia viridis</i> (Nitzsch.) Ehr.	x	x
<i>Placoneis clematis</i> (Grun.) Cox	x	
<i>Placoneis elginensis</i> (Greg.) Cox	x	
<i>Placoneis explanata</i> (Hust.) Lange-Bertalot	x	
<i>Planothidium delicatulum</i> (Kütz.) Round & Bukht.	x	x
<i>Planothidium dubium</i> (Grun.) Round & Bukht.	x	
<i>Planothidium lanceolatum</i> (Bréb. ex Kütz.) Round & Bukht.	x	x
<i>Planothidium oestrupii</i> (Cl.-Euler) Round & Bukht.	x	
<i>Planothidium rostratum</i> (Østrup) Lange-Bertalot	x	
<i>Pleurosigma delicatulum</i> W. Sm.	x	
<i>Pleurosira laevis</i> (Ehr.) Comprère	x	
<i>Psammothidium bioretii</i> (Germain) Bukht. & Round	x	x
<i>Psammothidium didymum</i> (Hust.) Bukht. & Round		x
<i>Psammothidium grischumum</i> f. <i>daonensis</i> (Lange-Bertalot) Bukht. & Round	x	
<i>Psammothidium helveticum</i> (Hust.) Bukht. & Round	x	x
<i>Psammothidium levanderi</i> (Hust.) Bukht. & Round	x	x
<i>Psammothidium marginulata</i> (Grun.) Bukht. & Round		x
<i>Psammothidium rossi</i> (Hust.) Bukht. & Round		x
<i>Psammothidium subatomoides</i> (Hust.) Bukht. & Round		x
<i>Pseudostaurosira brevistriata</i> (Grun.) Williams & Round	x	x
<i>Pseudostaurosira parasitica</i> (W. Sm.) Morales	x	x
<i>Pseudostaurosira robusta</i> (Fusey) Williams & Round		x
<i>Puncticulata bodanica</i> (Eulenstein ex Grun.) Håkansson	x	
<i>Reimeria sinutata</i> (Greg.) Kociolek & Stoermer	x	x
<i>Rhoicosphenia abbreviata</i> (C. Ag.) Lange-Bertalot B	x	
<i>Rhopalodia gibba</i> (Ehr.) O. Müll.	x	x
<i>Rhopalodia gibba</i> v. <i>minuta</i> Krammer	x	
<i>Rhopalodia gibberula</i> (Ehr.) O. Müll.	x	x
<i>Rossithidium linearis</i> (W. Sm.) Round & Bukht.	x	
<i>Rossithidium nodosum</i> (Cl.) M. Aboal	x	x
<i>Rossithidium petersenii</i> (Hust.) Round & Bukht.	x	x
<i>Rossithidium pusillum</i> (Grun.) Round & Bukht.	x	
<i>Sellaphora americana</i> (Ehr.) Mann	x	
<i>Sellaphora laevissima</i> (Kütz.) Mann		x
<i>Sellaphora pupula</i> (Kütz.) Mereschkowsky	x	x
<i>Sellaphora seminulum</i> (Grun.) Mann	x	
<i>Stauroneis anceps</i> Ehr.	x	x
<i>Stauroneis lundii</i> Hust.	x	
<i>Stauroneis phoenicenteron</i> (Nitz.) Ehr	x	x
<i>Stauroneis producta</i> Grun.	x	

## APPENDIX 2. Continued.

Species	Stream/river	Lake and pond
<i>Stauroneis undata</i> Hilse	x	
<i>Staurosira construens</i> Ehr.	x	x
<i>Staurosira construens</i> v. <i>binodus</i> (Ehr.) Hamilton		x
<i>Staurosira construens</i> f. <i>center</i> (Ehr.) Hamilton	x	x
<i>Staurosirella lapponica</i> (Grun.) Williams & Round	x	
<i>Staurosirella leptostauron</i> (Ehr.) Williams & Round	x	x
<i>Staurosirella pinnata</i> (Ehr.) Williams & Round	x	x
<i>Stenopterobia curvula</i> (W. Sm.) Krammer		x
<i>Stenopterobia densestriata</i> (Hust.) Krammer		x
<i>Stephanodiscus niagarae</i> Ehr.	x	x
<i>Stephanodiscus parvus</i> Stoermer & Håkansson		x
<i>Surirella amphioxys</i> W. Sm.	x	
<i>Surirella angustata</i> Kütz.	x	
<i>Surirella biseriata</i> Bréb.		x
<i>Surirella brebissonii</i> Krammer & Lange-Bertalot	x	
<i>Surirella brebissonii</i> v. <i>kuetzingii</i> Krammer & Lange-Bertalot	x	
<i>Surirella brightwellii</i> W. Sm.	x	
<i>Surirella elegans</i> Ehr.	x	
<i>Surirella linearis</i> W. Sm.	x	x
<i>Surirella minuta</i> Bréb.	x	
<i>Surirella ovalis</i> Bréb.	x	
<i>Surirella robusta</i> Ehr.		x
<i>Surirella splendida</i> (Ehr.) Kütz.	x	x
<i>Surirella subsalsa</i> W. Sm.	x	
<i>Surirella tenera</i> W. Greg.	x	
<i>Tabellaria binalis</i> (Ehr.) Grun.	x	
<i>Tabellaria fenestra</i> (Lyngbya) Kütz.	x	x
<i>Tabellaria flocculosa</i> (Roth) Kütz.	x	x
<i>Tetracyclus glans</i> (Ehr.) Mills	x	
<i>Tetracyclus rupestris</i> (Braun) Grun.	x	
<i>Tryblionella apiculata</i> Greg.	x	
<i>Tryblionella calida</i> (Grun.) Mann	x	
<i>Tryblionella compressa</i> (Bailey) M. Poulin	x	
<i>Tryblionella corctata</i> (Grun.) Mann	x	
<i>Tryblionella hungarica</i> (Grun.) Frenguelli	x	
<i>Tryblionella levidensis</i> W. Sm.	x	
<i>Tryblionella littoralis</i> (Grun.) Mann	x	
<i>Ulnaria ulna</i> (Nitz.) P. Compère	x	x
<i>Ulnaria ulna</i> v. <i>acus</i> (Kütz) Lange-Bertalot	x	