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A hydrothermal after-ripening time model for seed dormancy loss in *Bromus tectorum* L.

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Abstract

After-ripening, the loss of dormancy under dry conditions, is associated with a decrease in mean base water potential for germination of *Bromus tectorum* L. seeds. After-ripening rate is a linear function of temperature above a base temperature, so that dormancy loss can be quantified using a thermal after-ripening time (TAR) model. To incorporate storage water potential into TAR, we created a hydrothermal after-ripening time (HTAR) model. Seeds from two *B. tectorum* populations were stored under controlled temperatures (20 or 30 °C) and water potentials (–400 to –40 MPa). Subsamples were periodically removed from each storage treatment and incubated at 15 or 25 °C to determine germination time courses. Dormancy status (mean base water potential) was calculated from each time course using hydrothermal time equations developed for each seed collection. Seeds stored at –400 MPa did not after-ripen. At water potentials from –400 to –150 MPa, the rate of after-ripening increased approximately linearly with increasing water potential. Between –150 and –80 MPa, there was no further increase in after-ripening rate, while at –40 MPa seeds did not after-ripen and showed loss of vigour. These results suggest that the concept of critical water potential thresholds, previously shown to be associated with metabolic activity and desiccation damage in partially hydrated seeds, is also relevant to the process of after-ripening. The HTAR model generally improved field predictions of dormancy loss when the soil was very dry. Reduced after-ripening rate under such conditions provides an ecologically relevant explanation of how seeds prolong dormancy at high summer soil temperatures.

Keywords: after-ripening, *Bromus tectorum*, dormancy loss, hydrothermal after-ripening time, hydrothermal time, modelling, water potential

Introduction

Seed germination is strongly influenced by temperature and water potential, and can be described by models based on hydrothermal time. Hydrothermal concepts underlie many recent efforts to predict seed germination, as well as dormancy loss (reviewed by Allen, 2003; Bradford, 2005). Hydrothermal time was first proposed by Gummerson (1986) and further developed by Bradford (1990, 1995). The hydrothermal time equation for a given germination fraction is:

$$\theta_{HT} = (\Psi - \Psi_b(g))(T - T_b)t_g \quad (1)$$

where θ_{HT} is the amount of hydrothermal time (i.e. MPa°d) required for germination to occur, Ψ is the water potential of the incubation medium, $\Psi_b(g)$ is the base water potential below which germination will not occur for fraction g , T is the incubation temperature, T_b is the base (minimum) temperature for germination, and t_g is the actual time to germination for fraction g . In order to extend equation (1) to describe germination for a seed population, Gummerson (1986) assumed that the distribution of mean base water potentials within a population was normal. Probit transformation, which linearizes a cumulative normal distribution curve, could then be incorporated to predict germination for all seed fractions, as follows:

$$\text{Probit}(g/g_m) = [\Psi - \Psi_b(50) - \theta_{HT}/((T - T_b)t_g)]/\sigma_{\Psi_b} \quad (2)$$

where g_m is the fraction of viable seeds in the population, $\Psi_b(50)$ is the mean (median) base water potential of the population, and σ_{Ψ_b} is the standard deviation of base water potentials within the population.

While the assumptions underlying hydrothermal time have been questioned (Phelps and Finch-Savage, 1997; Hardegree *et al.*, 1999; Kebreab and Murdoch,

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1999), hydrothermal models do provide a quantitative description of seed germination rates based on actual Ψ and T conditions. Modelling efforts based on hydrothermal time have provided many insights into germination-related phenomena, and serve as a useful tool in predicting germination outcomes (Bradford, 2002). For example, treatments that cause an upward shift in $\Psi_b(50)$ lead to a decreased germination rate and, if sufficient, a decrease in germination percentage as well (Bradford, 1990, 1995; Christensen, *et al.*, 1996). Similarly, germination rate and percentage at a given incubation water potential are increased by more negative $\Psi_b(50)$ values. Shifts in $\Psi_b(50)$ above the water potential of the imbibition medium can explain germination behaviours such as dormancy cycling (Bradford, 2002), as well as the delay and inhibition of seed germination in the supra-optimal temperature range (Bauer *et al.*, 1998; Shrestha *et al.*, 1999; Meyer *et al.*, 2000; Alvarado and Bradford, 2002; Rowse and Finch-Savage, 2003).

The discovery that after-ripening rate is a linear function of storage temperature above a specific base temperature led to the use of thermal time to describe rates of dormancy loss in *Bromus tectorum* seeds (Christensen *et al.*, 1996), a winter annual that has invaded a wide variety of habitats in western North America (Mack, 1981). In order to predict dormancy loss in the field, Bauer *et al.* (1998) used the hydrothermal time parameter mean base water potential [$\Psi_b(50)$] as an index of dormancy status. Thermal after-ripening time (TAR) models successfully predicted dormancy loss in dry soil for *B. tectorum* seeds in the field as long as mean daily soil water potential remained above -150 MPa.

In earlier studies using seeds of wild oat (*Avena fatua*; Foley, 1994) and red rice (*Oryza sativa* L.; Leopold *et al.*, 1988), after-ripening was slowed or prevented at very low seed water contents. Citing unpublished data, Leopold and Vertucci (1989) reported that after-ripening of red rice seeds did not occur at water contents below $0.05 \text{ g H}_2\text{O} (\text{g dry weight})^{-1}$ and was also retarded at water contents above $0.15 \text{ g H}_2\text{O g}^{-1}$. Walters *et al.* (2001) suggested that specific reactions associated with water content thresholds in partially hydrated seeds could apply to a variety of developmental processes. For example, below *c.* -150 MPa, enzyme-mediated catabolic activities in seeds are restricted, while certain reactions leading to free radical production are increased (Leopold and Vertucci, 1989; Vertucci and Farrant, 1995).

The objectives of the present study were: (1) to determine how storage at low water potentials affects after-ripening rate for *B. tectorum* seeds; (2) to expand the TAR model to account for seed water potential, i.e. to develop a hydrothermal after-ripening time (HTAR) model to describe seed dormancy loss; and (3) to create and test a field simulation model using

HTAR concepts and measured values for soil water potential and temperature to predict dormancy loss under very dry seed-zone conditions.

The research was carried out in two phases. In the first phase we performed preliminary laboratory experiments on the effect of storage water potential on after-ripening. We used this provisional data set to construct a simulation model for dormancy loss in field seed zones with widely fluctuating water potentials, and tested this model with data from a field seed retrieval experiment. In the second phase, we conducted much more detailed laboratory experiments on the effect of water potential on dormancy loss, and developed a conceptual model to describe dormancy loss rates as a function of water potential over the range -400 to -40 MPa.

Materials and methods

Laboratory experiment – 1994

Mature florets (hereafter referred to as seeds) of *B. tectorum* were collected from two semi-arid Great Basin sites (Whiterocks, Utah, a salt desert shrub site, and Hobblercreek Canyon, Utah, a mountain brush site) in June 1994. Seeds were air-dried to a water content of $0.08\text{--}0.10 \text{ g H}_2\text{O} (\text{g dry weight})^{-1}$, cleaned by rubbing and fanning, and hand-examined to ensure fill.

Seeds were stored at factorial combinations of temperature (20 or 30°C) and water potential (-300 , -150 or -80 MPa). Storage water potentials were obtained by equilibrating seeds above saturated salt solutions (LiCl , MgCl_2 and CaNO_3 , respectively) in sealed containers (Winston and Bates, 1960; Schneider and Schneider, 1972). Subsamples of seeds were removed for evaluation of dormancy status at intervals from 0 to 14 weeks. Four replications of 25 seeds were used for each incubation treatment. Seeds were placed in 100×15 mm Petri dishes on two layers of blue blotter paper (Anchor Paper, St. Paul, Minnesota, USA) saturated with water and incubated at each of two alternating temperature regimes ($10/20^\circ\text{C}$ and $20/30^\circ\text{C}$, 12h:12h, with cool white fluorescent light during the warmer part of the temperature cycle). Dishes were stacked in clear plastic bags, with a water-saturated paper towel placed at the bottom of each bag to reduce evaporative loss. Germinated seeds (radicle emergence ≥ 1 mm) were counted and removed on days 1, 2, 4, 7, 11, 14, 21 and 28. Viability of the remaining ungerminated seeds was then determined using a cut test (e.g. Meyer *et al.*, 2000).

Field after-ripening experiment – 1994

A field retrieval study using the 1994 seed collections from Whiterocks and Hobblercreek was conducted,

starting within a week of seed harvest in June 1994 at Point of the Mountain, Utah, a sagebrush/grass site with sandy loam soil. Seeds were air-dried, placed inside nylon mesh bags and buried approximately 5 mm below the soil surface. Each bag contained approximately 200 seeds, as estimated by weighing. The bags were placed in four rows of 25 bags each, and four bags of each collection (one bag per row) were retrieved weekly from the experimental site. Seeds were transported from the field to the laboratory (about 30 min transit time) in plastic bags to minimize changes in water content. There was no precipitation and, consequently, no germination during the first 7 weeks of the retrieval experiment, the period over which we tracked dormancy loss in the field. The after-ripening portion of the field retrieval experiment was completed with the first major rains of late summer.

Seeds in each bag were divided into two approximately equal groups. The first group of seeds was incubated at 10/20°C and the second at 20/30°C (12 h:12 h with fluorescent light during the warm part of the cycle; hereafter reported as 15 and 25°C). Germination time courses were obtained as described previously. Temperature and water potential of the seed zone (approximately the top 1 cm of soil) at the field site were measured using thermistor (Omnidata, Logan, Utah, USA) and Aquatel sensors (Automata Inc., Grass Valley, California, USA), respectively. Measurements were recorded hourly, as an average of six 10-min readings, using a data logger (Omnidata Easylogger 900, Logan, Utah, USA). Aquatel sensors measure capacitance of the soil, which varies as a function of water content and soil characteristics. Laboratory calibrations were performed to determine water content values corresponding to soil capacitance readings. Corresponding water potential values were determined using a soil water release curve for this soil (Hanks, 1992).

A second method used to predict water potential in the field involved estimating seed-zone water potential, based upon measured temperature at the soil surface, as well as temperature and relative humidity at 1 m above the surface, and then solving for soil water potential according to the following equation:

$$\Psi_a = (RT/V_m)(\ln e/e^\circ) \quad (3)$$

where Ψ_a is the atmospheric water potential (in megapascals, MPa), R is the gas constant (8.2 MPa cm³ mol⁻¹ K⁻¹), T is the temperature of the soil in degrees Kelvin, V_m is the molar volume of water (18 cm³ mol⁻¹), and $\ln e/e^\circ$ is the natural logarithm for actual water vapour pressure divided by the saturating vapour pressure for that temperature ($\ln e/e^\circ \times 100 =$ relative humidity). This method assumes that the water vapour in the air and soil are the same, an assumption that is valid only in soils dry enough for the water content below the seed zone to be negligible (i.e. there is

no significant movement of water vapour to the seed zone from greater soil depths).

Laboratory experiment – 2002

Mature seeds of *B. tectorum* were hand-collected from Whiterocks and Hobblecreek Utah in June of 2002 and cleaned as described previously. Seeds were stored at factorial combinations of temperature (20 or 30°C) and water potential (–400, –350, –300, –200, –150, –80 or –40 MPa) in the dark. Storage water potentials were obtained by equilibrating seeds above saturated salt solutions [ZnCl₂, KOH, LiCl, (glycerol, see below), MgCl₂, CaNO₃, and NaCl, respectively; Winston and Bates, 1960; Schneider and Schneider, 1972] or glycerol solutions (–200 and –150 MPa; Forney and Brandl, 1992). A –200 MPa glycerol solution was used because the appropriate saturated salt solution proved unstable, and a –150 MPa glycerol solution was included to verify that glycerol and salt solutions produced similar results. Water potential over each salt solution varied slightly as a function of storage temperature; mean values were used for data analysis and presentation, except for the seed water content data. Seed water content was determined gravimetrically for subsamples of seeds equilibrated for 8 weeks at each storage temperature–water potential combination, as described by Copeland and McDonald (2001).

For evaluation of dormancy status, seeds were removed from storage at intervals ranging from 0 to 73 weeks and placed in 15 and 25°C incubation treatments in water, as described previously, except that seeds were incubated in the dark, with fluorescent light only during germination scoring. The use of constant temperature regimes for 2002 (i.e. 15 and 25°C incubation, instead of alternating 10/20 and 20/30°C) facilitated germinating seeds at several different water potentials throughout after-ripening, which would not have been practical with alternating temperature regimes. The timing of seed transfer from storage to incubation was based on how rapidly seeds lost dormancy. Seeds were stored at the lowest water potentials for up to 73 weeks, while seeds at less negative water potentials were stored for as little as 18 weeks. Germination time-course data were obtained for each treatment combination, as described previously.

Data analysis

In order to determine hydrothermal time parameters for each seed collection in both 1994 and 2002, fully after-ripened seeds were incubated at two temperatures (15 and 25°C) at each of four water potentials (0, –0.5, –1.0 and –1.5 MPa). Blotters were

saturated with water or solutions of polyethylene glycol (PEG) 8000 at the desired water potential. PEG was mixed according to Michel (1983). Germination time courses obtained, as described above, from all incubation water potential and temperature combinations for each seed collection were combined for analysis using repeated probit regression. The hydrothermal time analysis consisted of regressing probit (g) on $\Psi_b(g)$, calculated as $[\Psi - \theta_{HT}/(T - T_b)t_g]$ (Bauer *et al.*, 1998), adjusting the value of θ_{HT} until the highest R^2 value for the regression was obtained. From the regression line with the best fit, probit (g/g_m) = $m(\Psi_b(g)) + b$, mean base water potential ($\Psi_b(50)$) and standard deviation for base water potentials (σ_{Ψ_b}) were determined according to the following relationships: $\Psi_b(50) = -b/m$ and $\sigma_{\Psi_b} = 1/m$ (Christensen *et al.*, 1996).

Next, $\Psi_b(50)$ values were calculated from the germination time courses for seeds stored at each temperature–water potential–storage duration–incubation temperature combination. Once the hydrothermal parameters θ_{HT} and σ_{Ψ_b} had been determined for a seed collection, the $\Psi_b(50)$ characterizing each germination curve could be calculated from the relationship:

$$\Psi_b(50) = -\theta_{HT}/(T(t_{50})). \quad (4)$$

This equation can be derived from equation (1) by defining $T_b = 0^\circ\text{C}$ (the base temperature for germination in this species; Christensen *et al.*, 1996) and $\Psi = 0$ MPa. Estimation of $\Psi_b(50)$ for highly dormant seed samples (i.e. final germination < 50%) is described in detail in Bauer *et al.* (1998). These calculated $\Psi_b(50)$ values served as measures of the dormancy status of seed subsamples after storage for given intervals at each water potential–temperature combination and incubation at each temperature.

Characterizing decreases in $\Psi_b(50)$ (i.e. dormancy loss) through time at two storage temperatures makes it possible to determine the thermal time required for after-ripening. The TAR equation is:

$$\theta_{AT} = (T_s - T_1)t_{ar} \quad (5)$$

where θ_{AT} is the thermal time required for after-ripening, T_s is the storage temperature, T_1 is the base storage temperature (below which after-ripening does not occur), and t_{ar} is the actual time in storage required for completion of after-ripening [the time required for $\Psi_b(50)$ to change from its starting value to its final value; Bauer *et al.*, 1998]. T_1 for *B. tectorum* seeds was assumed to be 0°C , based on Bauer *et al.* (1998).

To expand TAR to include the effects of water potential, $\Psi_b(50)$ values calculated from the germination time course curves for each storage interval were regressed on thermal after-ripening time. A separate regression was performed for each storage water potential–storage temperature–incu-

bation temperature combination. The resulting lines are described by the equation:

$$\Psi_b(50) = m[(T_s - T_1)t_{ar}] + b \quad (6)$$

where b is the initial value of $\Psi_b(50)$ before any thermal time is acquired, and the slope (m) is the decrease in $\Psi_b(50)$ per unit thermal time (i.e. MPa/ $^\circ\text{C}$ weeks). The values for these slopes (dormancy loss rates) were then plotted against storage water potential to describe the influence of water potential on the rate of after-ripening.

In performing regressions to determine after-ripening rates, data obtained from storage from 0 to 240 degree-weeks were used. The decision to omit later values was based primarily on the observation that most after-ripening had occurred by this time, the response was nearly linear over this range, and a linear slope for each storage Ψ was easier to fit into a hydrothermal after-ripening model. Seeds stored at -40 MPa, which rapidly lost viability, were not included in further analyses.

Simulation model development

We used hourly seed-zone temperature and water potential values from the Point of the Mountain study site as driver variables for a simulation model to predict changes in $\Psi_b(50)$ during after-ripening in the field. Initial and final $\Psi_b(50)$ values from laboratory data were used as boundary values for starting and ending each simulation. The models were created using Microsoft Excel (Microsoft Works, Seattle, Washington, USA). They were similar in structure to our field simulation model for after-ripening as a function of thermal time (Bauer *et al.*, 1998), but explicitly incorporated the effect of water potential on after-ripening rate. For each hourly time step, the program calculated the expected decrement in $\Psi_b(50)$, based on thermal time alone, as in the TAR model. If the measured water potential was -150 MPa or greater for that hour, the thermal time-based decrement was applied. If the measured water potential was -375 MPa or lower, a decrement of zero was applied (i.e. we assumed no after-ripening at or below this water potential threshold). If the measured water potential fell in the range from -375 to -150 MPa, the decrement was calculated by subtracting the measured water potential from -150 MPa, then dividing by 225 MPa (the interval between -375 and -150 MPa) to give a proportion that was then multiplied by the decrement based on thermal time alone, to correct for water potential (see Fig. 1). Hydrothermal time equations for the two seed collections were used to calculate observed $\Psi_b(50)$ values for seeds incubated at two temperatures following each weekly retrieval. Predicted $\Psi_b(50)$ values after each time period in the

field were then compared with observed values (i.e. obtained by incubating subsamples retrieved from the field). For each seed collection and incubation temperature, three simulations were performed: one based on TAR alone, one based on HTAR using water potential estimates from the Aquatel sensor, and one based on HTAR using estimates based on relative humidity measurements.

Results

Hydrothermal time parameters

All four seed populations were at least partially dormant when recently harvested (Table 1). Recently collected seeds incubated at 25°C had germination percentages that were 13–51% lower than seeds incubated at 15°C, indicating that $\Psi_b(50)$ was higher at the warmer temperature before seeds after-ripened. For fully after-ripened seeds, $\Psi_b(50)$ was the same at both temperatures. The θ_{HT} values for the Hobblecreek collection were similar both years; however, θ_{HT} for the 1994 Whiterocks collection was nearly three times greater than for the 2002 Whiterocks collection. The low θ_{HT} value for the Whiterocks 2002 collection was offset by a high $\Psi_b(50)$ value, which resulted in a similar germination rate in water for both collections. The σ_{Ψ_b} values, which indicate germination uniformity, were nearly double for Hobblecreek

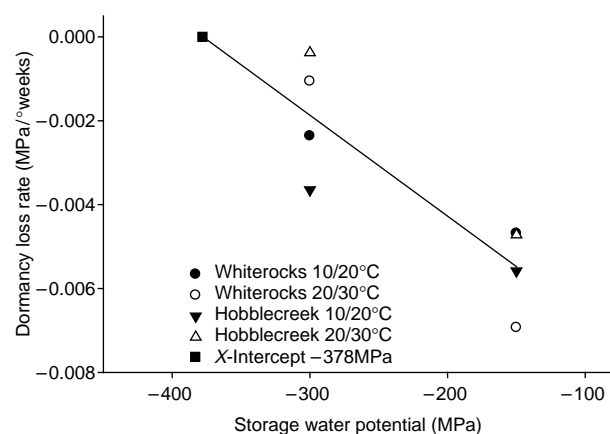


Figure 1. Dormancy loss rate for 1994 *Bromus tectorum* seeds, as indicated by changes in $\Psi_b(50)$ per unit thermal time, plotted as a function of storage water potential. Regressions for the Whiterocks and Hobblecreek seed collections were determined (not shown); but because these slopes are not significantly different, the equation based on the common slope (shown on the figure) was used as the best estimate of dormancy loss rate. The x -intercept represents the extrapolated water potential estimate below which after-ripening does not take place. Dormancy loss rate = 0.0000241 (Storage Water potential) - 0.0091 $R^2 = 0.731$; D.F. = 6; $P < 0.01$.

seeds collected in 2002 relative to the other three collections. R^2 values from probit regressions ranged from 0.84 to 0.91, which were reasonably high and similar to those reported previously (Bauer *et al.*, 1998).

Hydrothermal after-ripening time model – 1994

Seeds collected in 1994 and stored at -150 MPa after-ripened more quickly than those stored at -300 MPa (Fig. 1). Seeds stored at -80 MPa after-ripened at a rate essentially equivalent to the rate at -150 MPa (data not shown). Because of limited data (i.e. only two water potentials), we were forced to assume that the relationship between dormancy loss rate and water potential over the range -150 to -300 MPa was linear. In order to get the best estimate for the slope of this relationship, we combined data from both seed collections and both incubation temperatures into a single regression (Fig. 1). This regression also provides an extrapolated estimate of the water potential where after-ripening would be halted completely, namely the x -intercept, -375 MPa.

Field after-ripening simulation – 1994

Field seed-zone water potential for the week of 7–14 July 1994 was representative of the hot, dry weather that summer (Fig. 2). Estimated soil seed-zone water potentials showed wide diurnal fluctuation between $c. -150$ and -800 MPa (Fig. 2). Measured seed-zone temperatures fluctuated between 12 and 57°C during this same time period. Water potential estimates based on Aquatel capacitance readings showed less overall fluctuation (-150 to -600 MPa) than did estimates based on relative humidity measurements (-150 to -800 MPa), probably because the Aquatel sensor had reached its lower limit of detection. Both methods recorded a similar pattern of wide fluctuation in soil water potential values, and both consistently estimated very dry soil water conditions (almost never above -150 MPa).

Thermal time (TAR) predictions of dormancy loss in the field during this exceptionally hot, dry summer were consistently the most rapid, nearly always faster than observed values for change in $\Psi_b(50)$ (Fig. 3). For the two HTAR models, the model based on soil water potential estimated from atmospheric humidity yielded the slowest predicted rate of after-ripening, and the model based on Aquatel sensor readings resulted in intermediate predictions. Three out of four observed plots of actual rates of change in $\Psi_b(50)$ fell at or between values predicted by one of the two HTAR approaches. For Whiterocks seeds incubated at 10/20°C, the TAR model made the best prediction of dormancy loss, as indicated by decreased $\Psi_b(50)$, especially for longer durations of after-ripening.

Table 1. Germination percentages for recently harvested and fully after-ripened *Bromus tectorum* seeds, and hydrothermal time parameters for fully after-ripened seeds. Seeds were collected at two locations during June of 1994 and 2002

	Year*	Germination (%)				θ_{HT} (MPa°d)	$\Psi_b(50)$ (MPa)	σ_{Ψ_b} (MPa)	R^2
		At harvest		After-ripened					
		15°C	25°C	15°C	25°C				
Whiterocks	1994	69	56	97	95	42	-1.22	0.31	0.89
Whiterocks	2002	32	15	95	98	16	-0.80	0.24	0.84
Hobblecreek	1994	83	32	100	99	37	-1.17	0.31	0.91
Hobblecreek	2002	31	18	92	94	31	-1.22	0.56	0.85

*Incubation regimes in 1994 were actually 10/20°C and 15/25°C alternating (12h/12h) temperature regimes; mean temperatures of the regime are reported above.

Hydrothermal after-ripening time model – 2002

As expected, 2002 seed collections stored at more negative water potentials equilibrated at lower water contents; water content decreased more or less exponentially with decreasing water potential for both collections (Fig. 4). Under the range of storage conditions included in this study, water contents

ranged from 0.02 g H₂O (g dry weight)⁻¹ at -400 MPa to 0.19 g H₂O (g dry weight)⁻¹ at -40 MPa. Water contents at a given water potential were similar for the two seed collections, but were slightly higher at the lower storage temperature, as discussed by Walters (1998).

After-ripening, as indicated by decreasing $\Psi_b(50)$ over time, was essentially prohibited at the most negative water potential (-400 MPa; Fig. 5). To verify that seeds had not been killed by this treatment, subsets of seeds stored at -400 MPa for 73 weeks were transferred to storage at 20 or 30°C and -150 MPa for 8 weeks. Seeds were then incubated at 15 or 25°C in water for 28 d. Germination averaged 85% after 4 d and 91% after 28 d of incubation. In contrast, seeds stored at -40 MPa began to lose viability after only 8 weeks of storage, which led to spurious $\Psi_b(50)$ values.

Over the water potential range from -350 to -150 MPa, storage at progressively less negative water potentials resulted in correspondingly increased rates of after-ripening (Fig. 6). As in the 1994 data, no further acceleration of after-ripening occurred with storage at -80 MPa. The influence of storage water potential on after-ripening rate was similar for both collections and incubation temperatures. Near -400 MPa, or above -150 MPa, there was little change in after-ripening rate as a function of storage water potential. Between -150 and -350 MPa, the rate of after-ripening progressively increased (i.e. the dormancy loss rate became more negative, indicating a steeper rate of decrease in dormancy), as storage water potential became less negative. Therefore, the results of the 2002 experiments confirmed in broad outline the preliminary model developed from the 1994 data. Dormancy loss rates at -150 and -300 MPa were also similar in the two experiments (Figs 1, 6), resulting in similar slopes as a function of water potential over this range.

At water potentials between -300 and -80 MPa, after-ripening occurred more rapidly during the first 240 degree-weeks of storage (Fig. 5). Over this water potential range, the rate of after-ripening slowed as seeds neared completion of after-ripening. At

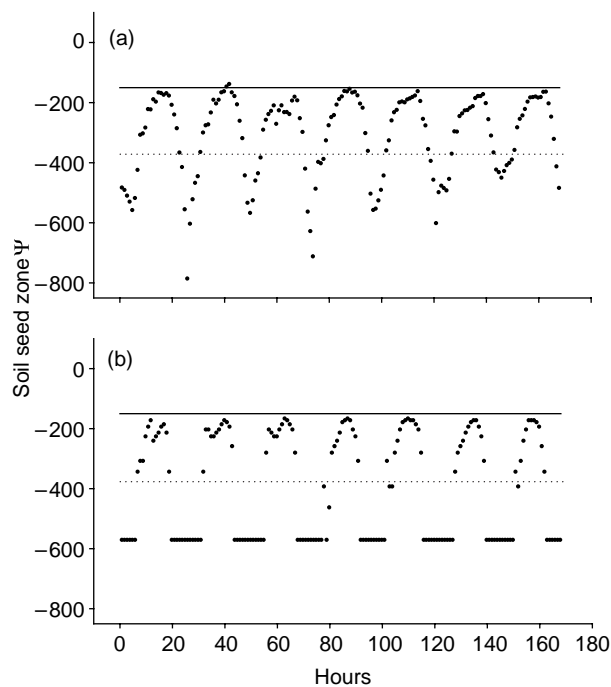


Figure 2. Field seed-zone water potential hourly estimates for the week of 7–14 July 1994 at Point of the Mountain, Utah, based on (a) soil capacitance readings and (b) measured atmospheric temperature and humidity, corrected for seed-zone temperature. The solid lines indicate the approximate water potential (-150 MPa) below which after-ripening is progressively reduced, while the dotted lines indicate the approximate water potential (-375 MPa) where after-ripening is completely halted, based on the equation for 1994 seeds in Fig. 1.

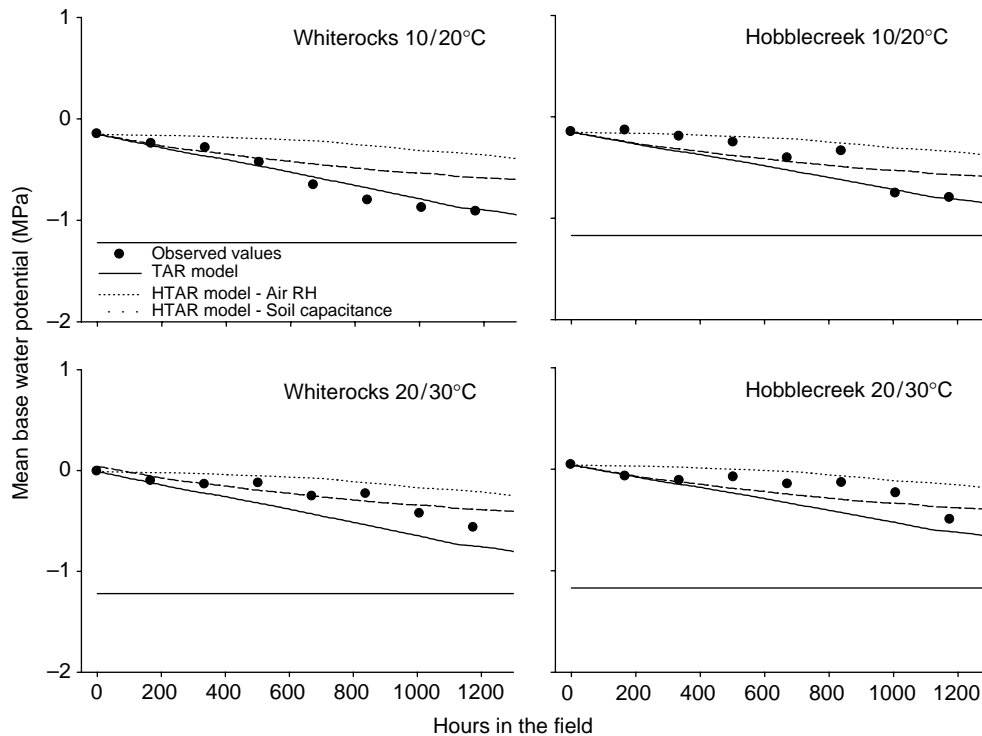


Figure 3. Predicted and observed changes in $\Psi_b(50)$ during field after-ripening at Point of the Mountain, Utah for two 1994 collections of *Bromus tectorum* seeds. The horizontal line in each graph represents $\Psi_b(50)$ for fully after-ripened seeds.

– 350 MPa, the decrease in $\Psi_b(50)$ during storage was approximately linear.

Hobblecreek seeds reached a much lower final $\Psi_b(50)$ value than did Whiterocks seeds (Table 1), but $\Psi_b(50)$ also decreased more rapidly during storage, especially at less negative water potentials. The slope of the regression line relating dormancy loss rate to water potential below –150 MPa was much steeper for the Hobblecreek collection (Figs 5, 6). The net result was that seeds from both 2002 collections required approximately the same amount of thermal time for completion of after-ripening at any given water potential.

Discussion

Physical and chemical reactions that occur in partially hydrated seeds appear to be limited by water potential thresholds, resulting in qualitative changes in the types of reactions that dominate at different levels of hydration (e.g. Vertucci and Farrant, 1995; Walters, 1998). It is interesting that Vertucci and Farrant’s proposed critical moisture level of –150 MPa, which these authors suggest as a discrete threshold for changes in metabolic activity between ‘Hydration Level 1’ and ‘Hydration Level 2’ (Fig. 1 in Vertucci and Farrant, 1995), appears to be identical to the threshold

where after-ripening *B. tectorum* seeds can best be explained by TAR or HTAR. While our data do not provide a physiological explanation for why after-ripening rate progressively declines below –150 MPa, Vertucci and Farrant discuss Hydration Level 2 as containing water with glassy characteristics believed to have strong interactions with both polar surfaces of macromolecules and hydroxyl groups of solutes. In contrast, Hydration Level 1 is associated with water that binds to macromolecules as a structural component. As water is progressively removed from seed tissues, remaining water is increasingly bound more tightly to macromolecules, influencing the type of reactions allowed, as well as their kinetics (Walters, 1998). Mechanistic studies of low seed moisture to date, addressing almost exclusively questions related to seed ageing, have shown that loss of viability is promoted at very low moisture contents. We emphasize here, and discuss later, that the very low water potentials experienced by *B. tectorum* seeds in 1994 resulted in no apparent harm under field conditions.

We propose a conceptual framework to describe the influence of water potential on after-ripening in *B. tectorum* seeds (Fig. 7). The diagram includes four important ranges of seed water potential, with associated thresholds that determine which model (TAR versus HTAR versus no after-ripening) best

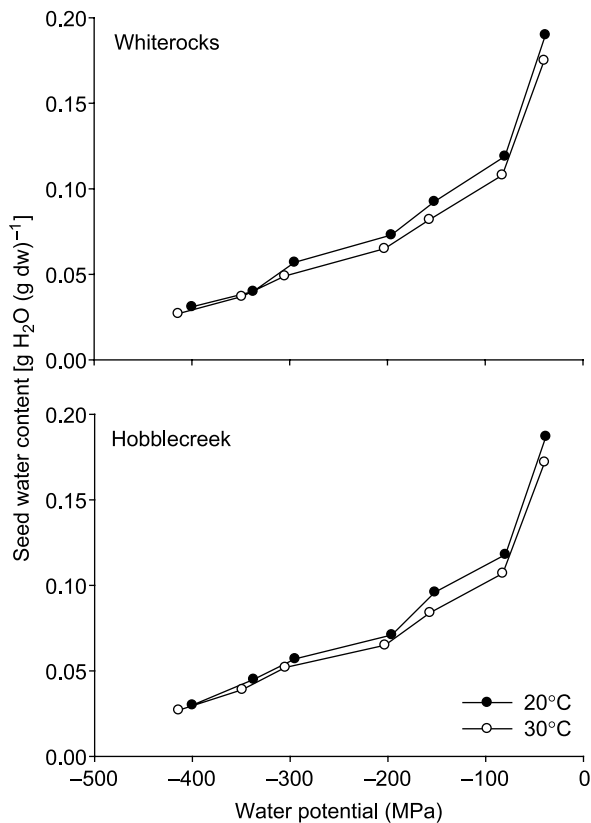


Figure 4. Water content for Whiterocks and Hobbblecreek collections of *Bromus tectorum* seeds stored at 20 or 30°C after 8 weeks of storage over saturated salt solutions at a range of water potentials. Standard errors for each value are all smaller than symbols.

predicts dormancy loss. These ranges are as follows: (1) seeds stored below approximately -375 MPa do not after-ripen; (2) seeds stored between -375 and -150 MPa after-ripen as a function of both temperature and water potential, and dormancy loss can be explained by HTAR; (3) seeds stored at or above -150 MPa experience after-ripening as a linear function of temperature alone (TAR); and (4) seeds stored above -40 MPa are too wet for after-ripening to occur (they deteriorate, slowly progress toward germination or remain imbibed but dormant). In the widely fluctuating temperature and water potential environment that characterizes the soil seed-zone of semi-arid habitats, seeds likely cycle repeatedly through two or more of these ranges, depending on the specific water potential conditions both in and below the seed zone. The ability to successfully predict the rate of dormancy loss in the field suggests that seeds indeed lose dormancy progressively by integrating the effects of fluctuating temperature and water potential. Additional support for the HTAR model is provided by successful field predictions of

after-ripening for seeds of the perennial bunchgrass *Elymus elymoides* (Bair, 2004).

Seeds stored at very low water contents have previously been reported to experience negligible after-ripening (Leopold *et al.*, 1988; Foley, 1994; Steadman *et al.*, 2003), but this finding with *B. tectorum* provides an ecologically relevant explanation for how seeds can be prevented from losing dormancy too rapidly. *B. tectorum* seeds mature in early summer, and typically experience weeks to months of hot, dry conditions that are not conducive to successful seedling establishment. If after-ripening were based solely upon thermal time, seeds would lose dormancy very quickly, and precocious germination (e.g. following summer thunderstorms) could result, reducing the probability of seedling survival. Seeds are prevented from losing dormancy at these times by an inhibited rate of after-ripening at very low water potentials, because high soil temperatures occur when the soil is also very dry.

While long-term storage at very low water potentials can damage seeds (e.g. Walters, 1998), we have not found this to be the case for wild plants adapted to desert and semi-desert habitats (e.g. Bauer *et al.*, 1998; Meyer *et al.*, 2000; Bair, 2004). Low water content probably does not affect viability of seeds in the field, possibly because fluctuating soil water potentials either prevent the accumulation of reactions that favour seed ageing and subsequent loss of viability, or they allow for repair processes to occur when seeds temporarily become hydrated. Given the high summer temperatures and low humidity in desert and semi-desert seed regions, the ability to survive as seeds under these conditions would clearly be advantageous.

The second range of storage water potentials proposed in Fig. 7 predicts dormancy loss according to a hydrothermal after-ripening time model. Model development often requires many simplifying assumptions. One decision made earlier in the development of TAR (Bauer *et al.*, 1998) was to ignore the influence of soil water potential. The assumption was based on the fact that soil water potential values averaged > -150 MPa during the year of study (1995), and TAR alone was a good predictor of actual after-ripening rates. However, in 1994, a particularly dry year, the TAR model overestimated the rate of after-ripening for field data. By including the combined influence of seed-zone water potential and temperature on after-ripening rate, the HTAR model generally predicted dormancy loss more accurately than TAR. We do not have an explanation for the single exception to superiority of the HTAR over the TAR model (Whiterocks seeds tested at 10/20°C). Because the HTAR model also produced better results than did

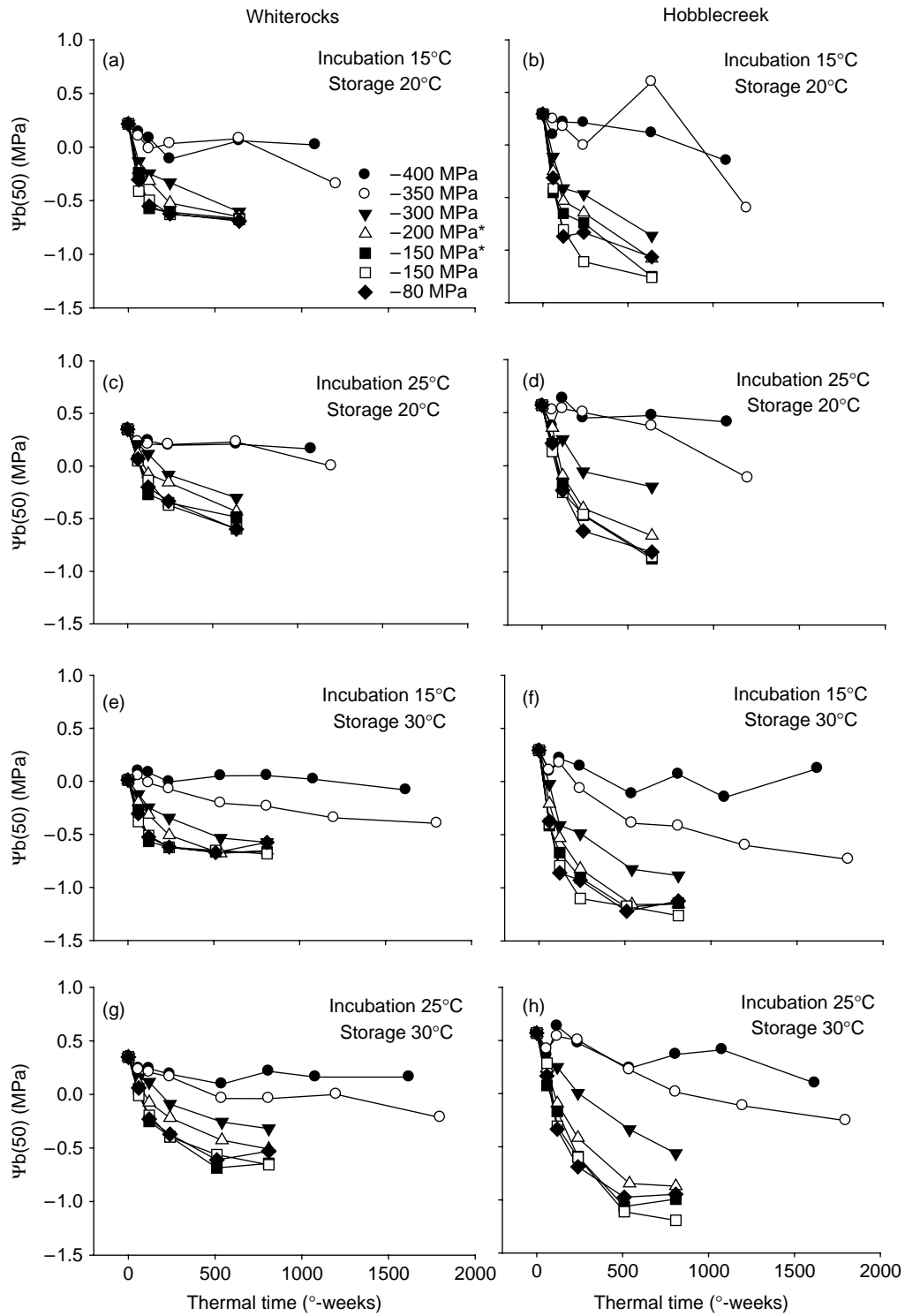


Figure 5. Dormancy status as indicated by $\Psi_b(50)$ for 2002-collected *Bromus tectorum* seeds from Whiterocks and Hobblecreek, as influenced by storage water potential, storage temperature, storage duration and incubation temperature. Water potentials marked with an asterisk indicate seeds were equilibrated above glycerol solutions; all other water potentials were achieved above saturated salt solutions. Values for -40 MPa are not included because seeds rapidly lost viability in this treatment, resulting in spurious $\Psi_b(50)$ values.

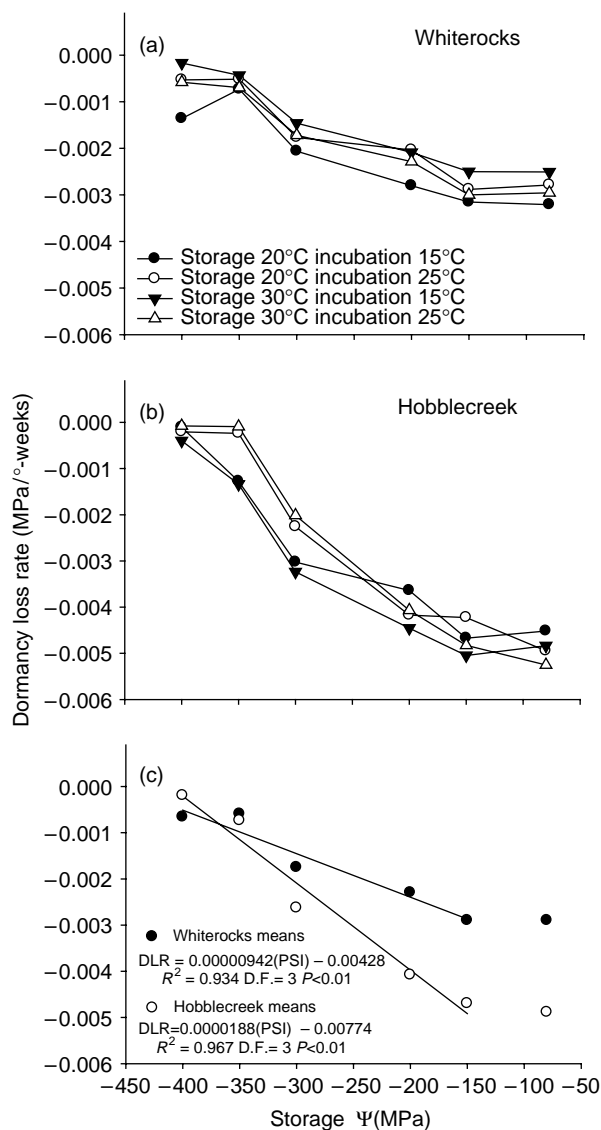


Figure 6. Dormancy loss rate [change in $\Psi_b(50)$ per unit thermal time] for 2002-collected *Bromus tectorum* seeds stored at 20°C or 30°C and incubated at 15°C or 25°C, plotted as a function of storage water potential: (a) Whiterocks and (b) Hobblecreek. Values for 0 to 240 degree-weeks of storage from Fig. 5 were used in regressions to obtain the slope of dormancy loss for each storage temperature–storage water potential–incubation temperature combination. (c) Dormancy loss rates for each seed population were averaged across storage temperature and incubation temperature at each storage water potential; mean dormancy loss rates were then regressed on storage water potential over the range of -150 to -400 MPa. The mean dormancy loss rate at -80 MPa is shown to demonstrate the lack of change as a function of water potential at water potentials > -150 MPa. The x -intercept predicted for Hobblecreek is -412 MPa and for Whiterocks is -454 MPa.

TAR for seeds of *E. elymoides* (two populations) included in this same experiment (Bair, 2004), the HTAR model should consistently provide a better overall model for use in very dry soils.

We believe that simulation modelling of dormancy loss for *B. tectorum* is ultimately limited by the ability to estimate seed water potentials accurately in the field. When *B. tectorum* seeds were alternated between -150 and -300 MPa on a diurnal basis, seed water content on average was much closer to the equilibrium value for -150 than for -300 MPa (P. Allen, unpublished data). This suggests that seed water potential does not necessarily reflect soil water potential values under wide diurnal fluctuations. In addition, studies with barley (*Hordeum vulgare*) suggest that water moves preferentially to the embryo from other seed tissues when previously imbibed seeds dry (Allen *et al.*, 2000). Thus, whole-seed measurements or estimates of water status may be unreliable under a diurnal cycle characterized by wide fluctuations in temperature and water potential.

The after-ripening rate of seeds stored between -80 and -300 MPa was approximately linear initially, followed by a progressively slower rate that often led to an overall curvilinear response. In some storage treatments, seeds failed to complete dormancy loss, which is partially a result of the experiment being terminated before a fully after-ripened state was attained. Seeds stored at -350 , -300 and -200 MPa after-ripened more slowly with time, and even began to level off at less negative $\Psi_b(50)$ values. If the underlying mechanisms associated with after-ripening include complex processes involving multiple reactions that occur at different rates, it is possible that some reactions might be slowed or prevented at water potentials above -400 MPa (Vertucci and Farrant, 1995). In addition, if a primary reaction controls the rate of initial after-ripening and a secondary reaction (i.e. with a slower rate) takes longer to complete, the combined result would be a curvilinear response. Gianinetti and Cohn (unpublished data, 2005) used a log transformation to linearize the negative curvilinear relationship between $\Psi_b(50)$ and thermal time, rather than just using the linear initial phase, to better describe dormancy loss in red rice seeds. However, for the purpose of predicting dormancy loss in the field, using the linear initial slope appeared to be sufficiently accurate, and simplified model development.

When water potentials were above -150 MPa and the soil was still relatively dry (the third region identified in Fig. 7), TAR was sufficient to predict after-ripening (Bauer *et al.*, 1998). For the TAR portion of the model, we assumed that the relationship between storage temperature and after-ripening rate

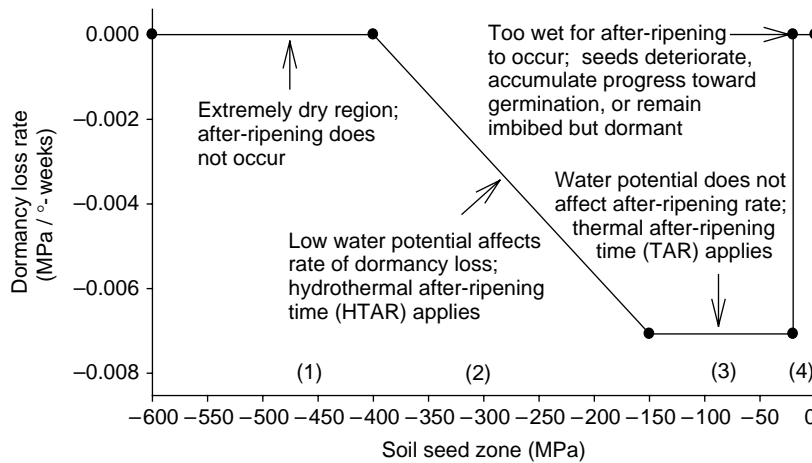


Figure 7. Conceptual diagram proposing how storage water potential influences after-ripening in *Bromus tectorum* seeds. The four regions and the associated threshold water potentials are discussed in the text.

was the same for both incubation temperatures. This implies that after-ripening rates would be approximately equal for all incubation temperatures, with only initial and possibly final $\Psi_b(50)$ values varying with incubation temperature (Meyer *et al.*, 2000). The 2002 data set generally supported this assumption. Failure to include this simplifying assumption would greatly complicate the simulation model, because in prediction of field germination based on dormancy status, incubation temperature is also a continuous and fluctuating variable, not a defined laboratory regime. An adequate test of the assumption that after-ripening rates are uniform across incubation temperatures would require multiple incubation temperature regimes that were not included in this study.

The fourth region in our conceptual after-ripening model (Fig. 7) is too wet for after-ripening to occur. At these water potentials, seeds deteriorate, accumulate progress toward germination or remain imbibed but dormant. In any case, storing seeds at constant water potentials above -40 MPa, but below the threshold for radicle emergence, creates experimental difficulties that prevent meaningful interpretation of after-ripening results.

Results from -40 MPa storage treatments were difficult to interpret due to rapid loss of viability. As seeds age they lose vigour, take longer to germinate (Ellis and Roberts, 1980, 1981) and eventually lose the ability to germinate (Walters, 1998). Extended exposure to water potentials near -40 MPa does not typically occur in semi-arid environments. Fluctuating temperatures, evaporation of water from the soil and precipitation all contribute to widely fluctuating water potential cycles. Although *B. tectorum* seeds regularly experienced water potentials above -40 MPa in the field during the summer of 1995 (Bauer *et al.*, 1998), it was generally just a few hours

before seeds were either close to 0 MPa or much drier. As with seeds frequently exposed to very dry soil conditions, seeds of *B. tectorum* that encountered moderately moist soil conditions did not lose viability in the field.

We propose a model that defines four ranges of water potential that influence the rate of after-ripening. The first range involves seeds that are experiencing very dry soil water potential conditions, where negligible after-ripening occurs. In the intermediate range, decreasing water potentials progressively inhibit after-ripening rate. The third range can be explained by thermal after-ripening time alone, and the wettest range fails to promote after-ripening (i.e. 'dry after-ripening' does not occur in wet seeds). Incorporating water potential into models that predict dormancy loss through after-ripening provides more accurate field predictions of dormancy loss than TAR alone. The model is consistent with both empirical and theoretical literature on the physiology of incompletely hydrated seeds (Vertucci and Farrant, 1995; Walters, 1998), although this literature is directed toward answering questions other than dormancy loss *per se*. The model has potential for making better predictions of dormancy loss under the widely fluctuating soil water potential conditions that occur in the field. Linking the hydrothermal after-ripening time with hydrothermal time for germination will be an important step in creating a combined model to account for both dormancy and germination under fluctuating water potential and temperature conditions.

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