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A list of arthropods of medical importance which occur in Utah with a review of arthropod-borne diseases endemic in the state

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Brigham Young University Science Bulletin

A LIST OF ARTHROPODS OF MEDICAL IMPORTANCE WHICH OCCUR IN UTAH WITH A REVIEW OF ARTHROPOD-BORNE DISEASES ENDEMIC IN THE STATE

by

Vernon J. Tipton and Robert C. Saunders



BIOLOGICAL SERIES — VOLUME XV, NUMBER 2

AUGUST 1971

BRIGHAM YOUNG UNIVERSITY SCIENCE BULLETIN BIOLOGICAL SERIES

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INTRODUCTION

The information contained in this paper has been assembled for the express purpose of demonstrating lacunae in current knowledge of health problems associated with arthropods in Utah. Obviously it is not definitive, nor is it intended to be, but rather it should serve as a base for future research investigations. It will be modified as additional information becomes available.

Not all of the arthropods in this list are of proven medical importance—in fact many of them are of doubtful importance—but they are included in the list because they are close relatives of species which are of medical significance in other areas and their potential for affecting the health of man has not yet been fully investigated.

Some difficulties are inherent in a review of the history of arthropod-borne diseases, particularly in rural areas. Records are incomplete and their accuracy is suspect. Some arthropod-borne diseases are not included on the standard state report form and thus in many instances are not reported. In rural areas, where many arthropodborne diseases occur, people may not seek medical care because of tradition or because no care is available. Diagnostic techniques are slow to reach rural areas and receive broad acceptance. Unless the index of suspicion is high, physicians are prone to ignore diagnostic tests necessary for specific discrimination. However, the history of tularemia in Utah provides ample evidence that rural medicine does not necessarily mean archaic methods and techniques. Dr. Richard A. Pearse, a Brigham City physician, published a clinical description of tularemia in humans which is considered to be the first account in the English language (Jellison, 1971). Many of the early epidemiological investigations of tularemia were conducted in the rural community of Delta and

were prompted by the astute observations of local physicians.

COLORADO TICK FEVER:

The work of Becker (1926, 1930) demonstrated that Colorado tick fever (CTF) is a disease entity distinct from Rocky Mountain spotted fever (RMSF) but it was not until 1940 that Topping, Cullyford, and Davis (1940) provided the first detailed clinical description of CTF. Consequently, accurate data on the incidence of CTF prior to 1940 are not available. Records of cases of CTF in Utah from 1940 to 1959 maintained by the Rocky Mountain Laboratory in Hamilton, Montana, are probably the most reliable but may be incomplete because information is based on CTF virus isolated from the blood of Utah residents at Hamilton and undoubtedly there were patients who were hospitalized elsewhere or not at all. Prior to 1960 the accuracy of records of CTF is in question because diagnostic techniques for the arboviruses were in their infancy. Comparatively simple but reliable tests are available but physicians may not utilize them either because of inconvenience or they may not be aware that the CTF virus persists for approximately 90 days after onset. Because of their epidemiological and clinical similarities CTF has been confused with RMSF. In children CTF may cause encephalitislike symptoms and even death (Eklund, Kennedy, and Casey, 1961) but probably there are few inapparent infections in a population. It is possible that many mild cases have escaped detection and have not been reported. According to Pratt and Rice (1969) there were only 96 cases of CTF reported in Utah during the period from 1956 to 1969 compared with 1,717 cases in Colorado during the same period. However, the low incidence of CTF in Utah may not be a

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true measure of its significance. There is no immunization available and treatment consists of supportive care.

Dermacentor andersoni Stiles is considered to be the most important vector in the epidemiological pattern involving man. However, CTF virus has been isolated from ticks of several other species indicating that they may play an important role in the cycle in nature. Ecological studies aimed at determining epidemiological patterns in nature and particularly the identity of reservoir animals are needed.

THE ENCEPHALITIDES:

Cases of western encephalitis (WE) and perhaps other encephalitides have probably occurred in humans and horses in Utah prior to the outbreak of 1933. However, the etiology and epidemiology of the encephalitides had not yet been elucidated, and one can only speculate about the prevalence of arthropod-borne viral agents in horses and humans prior to that date. In the Register of Deaths at the Salt Lake City Health Department, Bureau of Vital Statistics, brain fever was listed as the cause of death in several instances during the 1847-1865 period. It is possible that at least some of these deaths were due to infections with arboviruses.

The 1933 episode described by Madsen (1934) occurred in two waves, both of which began in the West Point area of Weber County. The first wave began about July, reached a peak about 10 August, and had almost completely subsided by 1 September. Madsen estimated that 1,139 sick horses were involved in the first wave, of which 43.9 percent died. The second wave, which began the middle of September and subsided about 1 November, involved 2,819 horses, of which about 53.2 percent died. The main focus of infection was in Salt Lake, Davis, Box Elder, Cache, and Weber counties in Utah and Franklin County in Idaho, but there were isolated eases in Rich, Summit, Morgan, Utah, and Tooele counties. An outbreak among humans occurred twenty-five years later (in 1958) in the same general locality in northern Utah and represents the largest number of eases reported in Utah for a single year (Jenkins and Donath, 1959). Serological studies revealed a high number of inapparent infections. Thomas and Smith (1959) conducted a survey on infection rates in mosquitoes, birds, and mammals and found that the highest infection rates in chickens and mosquitoes occurred in the geographical center of the human outbreak. In connection with the 1958 outbreak, Rees et al. (1959) concluded:

(1) There was in Utah in 1958 a recognized outbreak of Western Equine Encephalitis in man; (2) It was accompanied by a tremendous increase in the numbers of *C. tarsalis* mosquitoes; (3) *Culex tarsalis* mosquitoes were avidly feeding on man in considerable numbers during this period; and (4) Some *C. tarsalis* mosquitoes were harboring the Western equine strain of encephalitis virus.

Local physicians reported an extensive outbreak of human encephalitis of unconfirmed etiology in Box Elder County in 1936. In Weber County in 1956 there was a serologically confirmed case of St. Louis encephalitis and in 1957 a fatal case of SLE plus two cases of WE, one of which was fatal. There were 525 cases of equine encephalitis from 1955 to 1969, including 244 cases in the outbreak of 1958. There is no evidence of arbovirus activity in Weber County from 1933 to 1955. During recent years along the Wasatch front there have been occasional eases of a disease, difficult to diagnose but typical of western encephalitis. Physicians do not routinely submit acute and convalescent sera necessary for definitive diagnostic tests. Some additional information is needed on feeding preferences of mosquitoes and the role of passerine birds in the epidemiology of WE in areas where there have been active foei in the past. The fate of arboviruses during periods between epidemies is a perennial problem requiring investigation.

Other arboviruses which have been isolated in Utah include Hart Park-like isolates from Culex tarsalis, California encephalitis group isolates from Anopheles freeborni, Culiseta inornata, Aedes dorsalis, A. nigromaculis, Culex erythrothorax, Psorophora signipennis, and Culex tarsalis and Cache Valley isolates from Culiseta inornata and Anopheles freeborni (Holden and Hess, 1959; Crane et al., 1970; and Elbel et al., 1971).

ROCKY MOUNTAIN SPOTTED FEVER:

Beck (1955) reviewed the history of Rocky Mountain spotted fever (RMSF) in Utah. It is not certain when RMSF first occurred in the state, but probably the disease was prevalent among early settlers. Beck (loc cit.) reproduced a newspaper article, published in 1941, in which Dr. William M. McKay, acting commissioner of health for the state of Utah during that period, speculates that Brigham Young was afflicted with RSMF at the time he entered Salt Lake Valley. Some support for this view is supplied by Byington (in Beck, 1955) who believed that "mountain fever" was the same as RMSF. In the Register of Deaths at the Salt Lake City Health Department, Bureau of Vital Statistics,

several deaths are recorded for which the cause is listed as "mountain fever." Although there is insufficient evidence to establish a definitive relationship between the two, the season during which the deaths occurred is consistent with the epidemiology of RMSF. We do not overlook the possibility that "mountain fever" could be Colorado tick fever which also occurs in Utah under similar circumstances. The deaths for which "mountain fever" was listed as the cause occurred as follows: two in July and one in August of 1849; one in June and five in July of 1850; two in early November of 1854; and one in late May and two in September of 1855. In at least one instance the cause of death was given as "intermittent mountain fever." In the majority of cases the victims were adult males.

Dermacentor andersoni Stiles is considered to be the most important vector involved in human cases of RMSF in Utah although there may be other species important in the perpetuation of the disease in nature. Beck (1955) and Coffee (1953) gave data on seasonal and altitudinal distribution as well as life cycles of tick vectors of RMSF.

Jellison (1945) considers the cottontail rabbit, Sylvilagus nuttalli Bachman to be an important component of the RMSF biocenose. It is a vagile animal with a fairly high degree of ecological tolerance. Studies of the population dynamics of the ectoparasites of the cottontail rabbit may suggest ecological patterns which help to perpetuate the disease in nature.

From 1915 through 1969 the Utah State Department of Health, Bureau of Vital Statistics, recorded 496 cases of RMSF in Utah among both residents and nonresidents. Cases occurred most frequently during the months of June, July, and August, although others were reported as early as April and as late as November. Among arthropod-borne diseases with endemic foci in Utah, RMSF is second only to tularemia in total number of cases reported in the state.

PLAGUE:

The recorded history of plague in Utah is not dramatic, but nonetheless, plague represents a disease of great potential significance because of the widespread distribution of capable vectors and reservoirs in the state and the increasing number of fishermen and campers who invade the plague biocenose each year. There have been only two confirmed human cases (in 1936 and 1966) and one doubtful case (in 1939) reported in the state.

According to Allred (1952) and Beck (1955), capable vectors of plague are indige-

nous to every county in the state, and there are 41 proven reservoirs of plague in Utah, of which 36 are species of rodents. Stark lists 43 species of fleas which are classified as "capable (natural)" or "potential (experimental)" vectors. Xenopsylla cheopis (Rothschild), the most important vector of plague on a worldwide basis, has not been collected in large numbers in Utah. Diamanus mantanus (Baker) has been considered the most significant vector of plague in Utah, but Parker (1971) obtained Pasteurella pestis isolates repeatedly from Malaraeus sinomus (Jordan), Opisodasys keeni (Baker), Monopsyllus eumolpi (Rothschild), and Epitedia stanfordi Traub. The number of isolates was highest from specimens of M. sinomus and O. keeni associated with species of Peromyscus. Parker believes that plague is not limited to squirrel-flea complexes or to particular vegetative associations and topographic patterns. It is evident that vector efficiency varies considerably and is influenced by several environmental factors. Thus, there may be several species of rodent fleas with the potential to function effectively as vectors of plague as environmental conditions change and meet the requirements for transmission by a particular vector species. A study extending over several years relating population fluctuations of fleas on ground squirrels, on wood rats, and in their nests to environmental changes may be helpful in understanding vector and reservoir capabilities.

TULAREMIA:

Tularemia is a zoonotic disease which has probably been present in the wild fauna of Utah for hundreds of years. Thus it is tempting to speculate about its importance among the pioneers during the last half of the nineteenth century. A large segment of the male population in pioneer Utah was engaged in agricultural pursuits, particularly clearing of land, which suggests an invasion of an ecosystem in which transmission of tularemia was likely of common occurrence. Before the advent of white settlers, Indians may have been victims of tularemia inasmuch as rabbits, proven reservoirs of tularemia, probably constituted a significant part of their diet.

However, recorded history of tularemia in Utah begins about 1908 as indicated by Francis (1925):

There has existed in Utah, at least since 1908, a human disease known locally as deer-fly fever. What I believe to be the first clinical reference to human cases of tularemia is contained in a paper read before the Utah State Medical Association, Salt Lake City, October

3, 1910, by R. A. Pearse, Brigham City, Utah. Dr. Pearse refers to six cases, which occurred in the month of August, caused by the bite of a fly, on the exposed parts of the body (neck, ear, cheek, wrist, ankle, and hand). After an incubation period of from two to five days . . In 1919 and 1920, I studied seven eases of deer-fly lever near Fillmore, Millard County, Utah, and found them positive for tularemia, clinically, culturally, and serologically. The cases occurred in June, July and August during the seasonal prevalence of the fly Chrysops discalis. The sites of the fly bites were the neck, temple, ear, and posterior surface of the lower third of the thigh. In all cases, suppuration occurred in the glands draining the bitten area. All patients had fever; one died on the twenty-sixth day of illness. I heard of perhaps two dozen other cases in the general community in which I worked, From seventeen jackrabbits, sick or dead, in the community I isolated Bacterium tularense, thus establishing the great reservoir of infection.

In an earlier publication, Francis (1922) gives a more specific location of tularemia foci in Utah.

So far as known there have been but two foci of infection in Utah. The focus here reported is in Millard County, 5 miles (8 kilometers) west of Holden, 5 miles northwest of Fillmore, 25 miles (40 kilometers) southeast of Delta, and 120 miles (193 kilometers) south of Salt Lake City. The other focus has received clinical confirmation and is located near Brigham, a town 20 miles (32 kilometers) north of Ogden in Box Elder County. Both foci have probably existed for at least fifteen years.

Although tularemia does not usually occur in epidemic form, Hillman and Morgan (1937) reported an outbreak of 26 cases among a group of 170 enrollees of a Civilian Conservation Corps camp "located on the treeless plains north of Great Salt Lake." They suggested that the epidemiological evidence available pointed to deer flies as the vectors and jackrabbits as the reservoirs. The cases were diagnosed between 11 and 30 July 1935. There was a noticeable increase in the population of deer flies the week before the onset of the first case. Several men in the camp experienced multiple bites, and lesions on tularemia victims were on uncovered portions of the body. Jackrabbits were numerous; many were dead and several were lethargie.

Locomotive Springs, the site of the Civilian Conservation Corps camp, is in the general area of Tremonton where Pearse had seen cases in 1908 and 1910.

Russian workers have proposed subspecifie designations for the causative agents of tularemia which have been accepted by most North American workers. Francisella tularensis tularensis of North America is usually associated with rabbits and arthropods while the more cosmopolitan form, Francisella tularensis palearctica appears to be transmitted independent of arthropods and has been isolated from aquatic or semiaquatic rodents. An organism isolated from a water sample collected in Utah was given the name Francisella novicida (Larson, Wicht, and Jellison, 1955). All three forms have been found in Utah. Francisella tularensis tularensis is the principal cause of human tularemia but Francisella tularensis palearctica, isolated from muskrats, should be mentioned because of its importance in Utah.

According to the records of the Utah State Department of Public Health, Bureau of Vital Statistics, there have been 986 eases of tularemia in the state during the 45-year period from 1925 through 1969. Approximately three-fourths of these cases occurred during the twenty-year

period from 1935 to 1954.

Chrysops discalis has been shown to be an efficient experimental vector of tularemia; it has been known to bite man (Jellison, 1950). For these reasons it has been suspected of being the most important deer fly vector of tularemia in Utah. However, Cox (1965) found C. discalis to be less abundant than C. fulvaster and C. aestuans in study areas near Utah Lake. Moreover, he isolated F. tularensis from three of 73 pools of deer flies. Two isolates were obtained from two pools of C. fulvaster and one isolate from one pool of C. aestuans.

There is a particular need for investigation of seasonal and geographic distribution of species of *Chrysops* and the animals on which they feed, the duration of infection in reservoirs and vector species, and serological surveys of human populations in areas where there are high density populations of deer flies.

MALARIA:

Most cases of malaria which have occurred in the state were contracted elsewhere, but Marshall and Rees (1948), in their excellent review of malaria in Utah, have provided substantial evidence that local transmission has taken place, particularly in southern Utah. They point out that most of the early Utah settlers came from the Mississippi Valley where malaria was prevalent. Contact with the outside world was maintained through continuing immigration, returning missionaries, and settlers passing through on their way to California or Oregon. Perhaps the only case of malaria in Utah sufficiently well documented to be consid-

ered autochthonous is cited by them as follows: "In April, 1947, a vivax infection was reported in a two-year-old child of that area who had never been out of the state." The "area" referred to is southern Utah. Anopheles freeborni Aitken is widespread throughout the state and Anopheles franciscanus McKracken is widespread throughout the southern half of the state (Nielsen, 1968). Both are considered to be efficient vectors of malaria, especially the former.

In the Register of Deaths at the Salt Lake City Health Department, Bureau of Vital Statistics, there are several entries in which "malignant fever" and "bilious fever" are listed as the cause of death. Early physicians made the distinction among "fever," "mountain fever," "malignant fever," "bilious fever," and "typhoid fever," and although it would be inaccurate to associate malignant fever or bilious fever with malaria, there is a possibility that a persistent fever occurring during the summer months could be malaria.

In a five-year period from 1943 through 1947, 723 cases of malaria were reported in Utah and reflect the impact of returning servicemen on the incidence of disease within the state. There was another less dramatic rise in the incidence of malaria in Utah, associated with the Korean War, during the period from 1951 to 1955 when 75 cases were reported. Nevertheless, with an adequate reservoir of infection, capable vectors, and a susceptible resident population, malaria has not become established in Utah. Rapid diagnosis and treatment of servicemen, improved mosquito control, and an informed public are the principal factors which mitigate the importance of malaria in Utah. The feeding habits of the mosquito vectors in a rural setting may be another factor of some importance. Cattle and horses are the preferred sources of blood meals for some Anopheles species.

MISCELLANEOUS:

Relapsing fever is virtually unknown in Utah, although it has been reported on several occasions from surrounding states. Davis (1939) reported a single case which occurred near Salt Lake City in 1928. Both *Ornithodoros parkeri* and *O. turicata*, proven vectors of relapsing fever, occur in the state although their distribution is not completely known.

Coxiella burnetti, the causative agent of Q fever, has been isolated from rodents (Dipodomys ordii, D. microps, and Peromyscus maniculatus) and a tick (Dermacentor parumapterus) in the Great Salt Lake Desert in Utah. C. burnetti antibodies were demonstrated

serologically in *Lepus californicus*, *Onychomys leucogaster*, and *Eutamias minimus* (Stoenner et al., 1959). There is no record of Q fever in

Cases of Selected Arthropod-Borne Diseases in Utah 1915-1969

Year	CTF	WE	Malaria	RMSF	Tularemia		
1915				35			
1916				34			
1917				15			
1918				5			
1919				10			
1920				9			
1921				8			
1922				18			
1923				15			
1924				10			
1925		2		6	1		
		4		5	4		
1926				7	1		
1927			1	10	1		
1928			1		0		
1929				11			
1930				13	4		
1931				10	2		
1932				22	4		
1933				5	6		
1934			1	12	6		
1935			3	14	37		
1936				8	5		
1937			2	12	41		
1938			10	19	73		
1939		2	I	24	44		
1940	2	2	5	15	55		
1941		2 2 5	0	13	45		
1942		0	5	8	48		
1943		12	313	12	34		
1944		5	157	10	23		
1945		2	112	9	28		
1946		$\frac{2}{4}$	93	5	29		
1947		6	48	5	36		
1948		9	2	4	45		
1949	1 °	2 3	2	14	35		
	1	5	$\vec{0}$	7	40		
1950	1°	2	23	18	30		
1951	10	8	38	9	20		
1952				9	40		
1953	1°	6	7	4	22		
1954	20	6	4		29		
1955	3°	2	3	7			
1956	3	100	0	3	21		
1957	4	300	1	9	30		
1958	0	47	0	0	17		
1959	4	38	2	1	10		
1960	4	23	0	3	19		
1961	2	6	0	1	17		
1962	8	8	0	0	16		
1963	12	12	0	0	3		
1964	12	1	0	1	24		
1965	11	0	1	0	10		
1966	4	1	3	()	3		
1967	11	Ô	3	0	7		
1968	5	0	1	0	5		
1969	16	0	3	2	16		
1909	10						

^{*}Represents isolations from patients hospitalized at Rocky Mountain Laboratory, Hamilton, Montana.

One case of SLE.

humans in Utah, but this may be due to faulty diagnosis or reporting, inasmuch as human cases have occurred in surrounding states.

Mohr (1951), in his paper on the distribution of murine typhus and plague in the United States, gives no records of murine typhus for the state of Utah. The flea index of Xenopsylla cheopis on Rattus sp. apparently has never been very high in Utah, and probably accounts for the absence of the disease in the state.

Armstrong (1922) reported an epidemic of typhus on the San Juan Indian Reservation during the last half of 1920 and the first half of 1921 in which there were 63 cases of typhus with 27 deaths among approximately 7,000 Indians. The San Juan Indian Reservation is 5,884 square miles in the four-corners area of New Mexico, Arizona, and Utah.

One case of dengue was reported in 1942, but it was probably contracted outside the state.

One doubtful case of rickettsialpox has been

reported from Utah (Pratt and Rice, 1969).

In Utah the incidence of bites and stings of arthropods and the number of cases of dermatitis eaused by urticating and vesicating insects is unknown. Scattered eases of archinidism have been reported, including at least one death from the sting of a hymenopterous insect. Latrodectus hesperus Chamberlin and Ivie and several species of Hymenoptera are the most important venomous arthropods in the state.

Tick paralysis, caused by the bite of female ticks, *Dermacentor andersoni* Stiles, occurs most frequently in an area comprising the northern part of Idaho and adjacent portions of Washington and Montana. Isolated cases have been reported in other sections of the Rocky Mountains where *Dermacentor andersoni* occurs (Philip, 1969). Insofar as we are aware there have been no cases of tick paralysis reported in Utah, but the possibility of its occurrence should not be overlooked.

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LIST OF ARTHROPODS

ARACHNIDA

I. Acarina: Ixodides

A. Argasidae

1. Argas

A. cooleyi Kohls and Hoogstraal, 1960 A. giganteus Kohls and Clifford, 1968

A. sanchezi Dugès, 1887

2. Ornithodoros

O. concanensis Cooley and Kohls, 1941 O. eremicus Cooley and Kohls, 1941

O. kelleyi Cooley and Kohls, 1941 O. parkeri (Cooley, 1936)

O. sparnus Kohls and Clifford, 1963 O. talaje (Guérin-Mèneville, 1849)

O. turicata (Dugès, 1876)

3. Otobius

O. megnini (Dugès, 1884)

O. lagophilus Cooley and Kohls, 1940

B. Ixodidae

1. Dermacentor

D. albipictus (Packard, 1869)

D. andersoni Stiles, 1908 D. lunteri Bishopp, 1912 D. parumapertus Neumann, 1901

2. Haemaphysalis

H. leporispalustris (Paekard, 1869)

3. Ixodes

I. angustus Neumann, 1899

I. jellisoni Cooley and Kohls, 1938

I. kingi Bishopp, 1911I. marmotae Cooley and Kohls, 1938

1. muris Bishopp and Smith, 1937

I. ochotonae Gregson, 1941

I. pacificus Cooley and Kohls, 1943

I. sculptus Neumann, 1904 I. soricis Gregson, 1942

1. spinipalpis Hadwen and Nuttall, 1916

I. texanus Banks, 1908 I. woodi Bishopp, 1911

4. Rhipicephalus

R. sanguineus (Latreille, 1806)

I'. Acarina: Mesostigmata

A. Dermanyssidae

1. Dermanyssus

D. gallinae (De Geer, 1778)

2. Hirstionyssus

H. bisetosus Allred, 1957

H. cynomys (Radford, 1941)

H. eutamiae Allred and Beck, 1966

H. femuralis Allred, 1957

II. hilli (Jameson, 1950)

II. incomptus (Eads and Hightower, 1952) H. isabellinus (Oudemans, 1913)

H. latiscutatus (de Meillon and Lavoipierre,

II. longichelae Allred and Beck, 1966

II. ncotomae (Eads and Hightower, 1951)

II. oecidentalis (Ewing, 1923)

H. paraffinis Herrin, 1970 II. perognathi Herrin, 1970

H. staffordi Strandtmann and Hunt, 1951

H. talpae Zemskaya, 1955

II. thomomys Allred and Beek, 1966

H. torus Allred and Beck, 1966

II. triacanthus (Jameson, 1950) II. utahensis Allred and Beck, 1966

3. Liponyssoides

L. becki (Allred, 1957) L. sanguineus (Hirst, 1914)

4. Myonyssus

M. montanus Furman and Tipton, 1955

B. Haemogamasidae

1. Brevisterna

B. montanus (Ewing, 1922) B. utahensis (Ewing, 1922)

2. Eulaelaps

E. stabularis (Koch, 1836)

3. Haemogamasus

H. alaskensis Ewing, 1925

H. barberi (Ewing, 1925) H. nidiformis Bregetova, 1955

II. occidentalis (Keegan, 1951)

H. pontiger (Berlese, 1903)

4. Ischyropoda

I. armatus Keegan, 1951

I. furmani Keegan, 1951

C. Halaraehnidae

1. Zumptiella

Z. bakeri (Furman, 1954)

D. Laelapidae

1. Androlaelaps

A. circularis (Ewing, 1933)

A. crowei Jameson, 1947 A. debilis Jameson, 1950 A. fenilis (Megnin, 1876)

A. geomys Strandtmann, 1949

A. glasgowi (Ewing, 1925)

A. hollisteri (Ewing, 1925)

A. leviculus Eads, 1951

2. Hypoaspis

H. gurabensis (Fox, 1946)

II. Iubrica Oudemans and Voigts, 1904

3. Laelaps

L. incilis Allred and Beck, 1966

L. kochi Oudemans, 1936

L. multispinosus Banks, 1909

L. nuttalli Hirst, 1915

E. Macronyssidae

1. Chrioptomyssus

C. robustipes (Ewing, 1925)

2. Ornithonyssus

O. aridus Furman and Radovsky, 1963

O. bacoti (Hirst, 1913)

O. silviarum (Canestrini and Fanzago, 1877)

3. Steatonyssus

S. antrozoi Radovsky and Furman, 1963

F. Rhinonyssidae

1. Paranconyssus

P. ieteridius Strandtmann and Furman, 1956

G. Spinturnicidae

1. Paraspinturnix

P. globosus Rudnick, 1960

2. Spinturnix

S. orri Rudnick, 1960

I". Aearina: Oribatei Garabodidae

Passalozetes

P. linearis Higgins and Woolley, 1962

I'''. Acarina: Trombidiformes

A. Trombiculidae

1. Acomatacarus

A. arizonensis Ewing, 1942

2. Chatia

C. ochotona (Badford, 1942)

C. sctosa Brennan, 1946

3. Cheladonta

C. crossi Lipovsky, Crossley, and Loomis, 1955

4. Euschoengastia

E. cordiremus Brennan, 1948

E. criceticola Brennan, 1948

E. cynomyicola Crossley and Lipovsky, 1954

E. decipiens Gould, 1956

E. fasolla Brennan and Beck, 1955

E. furmani Gould, 1956 E. hoffmanae Gould, 1956

E. lanceolata Brennan and Beek, 1955

E. lanei Brennan and Beek, 1955

E. lutcodema Brennan, 1948

E. obesa Brennan and Beck, 1955

E. oregonensis (Ewing, 1929)

E. pomerantzi Brennan and Jones, 1954

E. radfordi Brennan and Jones, 1954

E. rotunda Brennan and Beek, 1955

E. sciuricola (Ewing, 1925)

E. soricinus Gould, 1956

5. Euschoengastoides

E. lacerta (Brennan, 1948)

E. hoplai (Loomis, 1954)

E. utahensis (Brennan and Beek, 1955)

6. Eutrombicula

E. belkini (Gould, 1950)

7. Hexidionis

H. allredi (Brennan and Beck, 1955)

H. doremi (Brennan and Beck. 1955)

8. Hyponeocula

II. arenicola (Loomis, 1954)

II. montanensis (Brennan, 1946)

9. Gahrliepia

G. americana (Ewing, 1942)

10. Leeuwenhoekia

L. americana (Ewing, 1942)

11. Leptotrombidium

L. myotis (Ewing, 1929)

L. panamensis (Ewing, 1925) L. potosina (Hoffman, 1950)

12. Miyatrombicula

M. csocnis (Sasa and Ogata, 1953)

M. sargenti (Brennan, 1952

13. Neoschoengastia

N. americana (Hirst, 1921)

14. Neotrombicula

N. californica (Ewing, 1942)

N. harperi (Ewing, 1928)

N. jewetti (Brennan and Wharton, 1950)

N. microti (Ewing, 1928)

N. subsignata (Brennan and Wharton, 1950)

15. Odontacarus

O. hirsutus (Ewing, 1931)

O. linsdalei (Brennan and Jones, 1954)

O. micheneri Greenberg, 1952

16. Trombicula

T. bakeri Ewing, 1946

T. kardosi Loomis, 1954

T. univari Brennan, 1965

17. Whartonia

W. perplexa (Brennan, 1947)

B. Myobiidae

Radfordia

R. bachai Howell and Elzinga, 1962

R. lemina (Koch, 1841)

R. subuliger Ewing, 1938

II. Araneida

A. Loxoscelidae

Loxosceles

L. unicolor Keyserling, 1887

B. Theridiidae

Latrodectus

L. hesperus Chamberlin and Ivie, 1935

III. Seorpionida

A. Buthidae

Centruroides

C. sculpturatus Ewing, 1928

B. Vejovidae

1. Anuroctonus

A. phaiodactylus (Wood, 1863)

II. arizonensis Ewing, 1928

II. spadix, Stahnke, 1940

3. Vejovis

V. becki Gertsch and Allred, 1965

V. boreus (Girard, 1845) V. concusus Stahnke, 1940 V. utahensis Williams, 1968

V. wapatkiensis Stahnke, 1940

INSECTA

I. Anoplura

A. Haematopinidae

Haematopinus

II. asini (Linnaeus, 1758)

II. eurysternus (Nitzsch, 1818)

H. suis (Linnaeus, 1758)

B. Hoplopleuridae

1. Enderleinellus

E. marmotae Ferris, 1919

E. osborni Kellogg and Ferris, 1915

E. paralongiceps Kim, 1966

E. suturalis (Osborn, 1891)

E. tamiasciuri Kim, 1966

2. Fahrenholzia

F. pinnata Kellogg and Ferris, 1915

F. reducta Ferris, 1922

3. Haemodipsus

II. lyriocephalus (Burmeister, 1839)

II. setoni Ewing, 1924

H. ventricosus (Denny, 1842)

4. Hoplopleura

H. acanthopus (Burmeister, 1839)

II. arboricola Kellogg and Ferris, 1915

H. captiosa Johnson, 1960

H. difficilis Kim, 1965

II. erratica (Osborn, 1896)

H. ferrisi Cook and Beer, 1959 II. hesperomydis (Osborn, 1891)

II. onychomydis Cook and Beer, 1959

II. pacifica Ewing, 1924

H. reithrodontomydis Ferris, 1951

H. sciuricola Ferris, 1921

II. trispinosa Kellogg and Ferris, 1915

5. Neohaematopinus

N. citellinus Ferris, 1942

N. inornatus (Kellogg and Ferris, 1915)

N. laeviusculus (Grube, 1851)

N. marmotae Ferris, 1923

N. neotomae Ferris, 1942

N. pacificus Kellogg and Ferris, 1915

N. sciuri Janeke, 1931

N. sciuropteri (Osborn, 1891) N. semifasciatus Ferris, 1916

N. spilosomae Pratt and Stojanovich, 1961

6. Polyplax

P. alaskensis Ewing, 1927

P. auricularis Kellogg and Ferris, 1915

P. borealis Ferris, 1933

P. serrata (Burmeister, 1839)

P. spinulosa (Burmeister, 1839)

C. Linognathidae

1. Linognathus

L. africanus Kellogg and Paine, 1911

L. pedalis (Osborn, 1896)

L. setosus (von Olfers, 1816)

L. stenopsis (Burmeister, 1838)

L. vituli (Linnaeus, 1758)

2. Solenoptes

S. binipilosus (Fahrenholz, 1916)

S. capillatus Enderlein, 1904

S. ferrisi (Fahrenholz, 1916)

D. Pediculidae

1. Microphthirus

M. uncinatus (Ferris, 1916)

2. Pediculus

P. humanus Linnaeus, 1758

P. pubis (Linnaeus, 1758)

II. Orthoptera

A. Blattidae

1. Arenivaga

A. crratica Rehn, 1907

2. Blatta

B. orientalis (Linnaeus, 1758)

3. Blatella

B. germanica (Linnaeus, 1767)

4. Panchlora

P. nivea (Linnaeus, 1758)

5. Periplaneta

P. americana (Linnaeus, 1758)

P. australiasiae (Fabricius, 1775)

6. Suppella

S. longipalpa (Fabricius, 1798)

HI. Colcoptera

A. Leptinidae

1. Leptinillus

L. validus (Horn, 1872)

2. Platypsyllus

P. castoris Ritsema, 1869

B. Meloidae

1. Epicauta

E. fabricii (LeConte, 1853)

E. ferruginea (Say, 1823)

E. normalis Werner, 1945

E. puncticollis (Mannerheim, 1843)

2. Lytta

L. cyanipennis (LeConte, 1851)

3. Nemognatha

N. lurida LeConte, 1853

N. lutea LeConte, 1853

IV. Diptera

A. Calliphoridae

1. Aldrichina

A. grahami (Aldrich, 1930)

2. Bufolucilia

B. silvarum (Meigen, 1826)

3. Calliphora

C. coloradensis Hough, 1899 C. livida Hall, 1948

C. terraenovae Macquart, 1851

C. vicina Robineau-Desvoidy, 1830

C. comitoria (Linnaeus, 1758)

4. Cochliomyia

C. hominivorax (Coqueral, 1858)

C. macellaria (Fabricius, 1775)

5. Cynomyopsis

C. cadaverina (Robineau-Desvoidy, 1830)

6. Eucalliphora

E. lilaea (Walker, 1849)

7. Lucilia

L. illustris (Meigen, 1826)

8. Phaenicia

P. scricata (Meigen, 1826)

9. Phormia

P. regina (Meigen, 1826)

10. Pollenia

P. rudis (Fabricius, 1794)

11. Protocalliphora

P. aenea Shannon and Dobroseky, 1924 P. asiovora Shannon and Dobroseky, 1924

P. cuprina (Hall, 1948)

P. hesperia Shannon and Dobroseky, 1924

P. hirudo Shannon and Dobroscky, 1924

P. hirundo Shannon and Dobroseky, 1924

P. metallica (Townsend, 1919)

P. sialia Shannon and Dobroseky, 1924

12. Protophormia

P. terraenovae (Robineau-Desvoidy, 1830)

B. Ceratopogonidae

1. Culicoides

C. baueri Hoffman, 1925

C. cochisensis Wirth and Blanton, 1967

C. cockerellii (Coquillett, 1901)

C. crepuscularis Malloch, 1915

C. freeborni Wirth and Blanton, 1969

C. haematopotus Malloch, 1915

C. hieroglyphicus Malloch, 1915

C. montanus Wirth and Blanton, 1969

C. obsoletus (Meigen, 1818)

C. palmerae James, 1945

C. stellifer (Coquillett, 1901)

C. usingeri Wirth, 1952 C. utaliensis Fox, 1946

C. variipennis variipennis (Coquillett, 1901)

2. Leptoconops

L. kerteszi Hieffer, 1908

C. Chloropidae

Hippelates

H. microcentrus Coquillett, 1904

H. montanus Sabrosky, 1941

11. pallipes (Loew, 1865)

II. particeps (Beeker, 1912)

H. pusio Loew, 1872

D. Culicidae

1. Aedes

A. atropalpus (Coquillett, 1902)

A. campestris Dyar and Knab, 1907

A. cataphylla Dyar, 1916

A. cinereus Meigen, 1818

A. communis (De Geer, 1776)

A. dorsalis (Meigen, 1830)

A. excrucians (Walker, 1856)

A. fitchii (Felt and Young, 1904) A. flavescens (Muller, 1764) A. hexodontus Dyar, 1916

A. impiger (Walker, 1848)

A. implicatus Vockeroth, 1954

A. increpitus Dyar, 1916

A. intrudens Dyar, 1919

A. melanimon Dyur, 1924

A. nielseni O'Meare and Craig. 1970

A. nigromaculis (Ludlow, 1907)

A. niphadopsis Dyar and Knab, 1918

A. pullatus (Coquillett, 1904)

A. schizopinax Dyar, 1929

A. sierrensis (Ludlow, 1905)

A. spencerii idahoensis (Theobald, 1901).

A. sticticus (Meigen, 1838)

A. trivittatus (Coquillett, 1902)

A. varipalpus (Coquillett, 1902) A. ventrovittis Dyar, 1916

A. vexans (Meigen, 1830)

2. Anopheles

A. earlei Vargas, 1943

A. franciscanus McCracken, 1904

A. freeborni Aitken, 1939

3. Coquillettidia (Mansonia) C. perturbans (Walker, 1856)

4. Culex

C. apicalis Adams, 1903

C. erythrothorax Dvar, 1907

C. pipiens pipiens Linnaeus, 1758

C. pipiens quinquefasciatus Say, 1823

C. restuans (Theobald, 1901)

C. tarsalis Coquillett, 1896

C. territans Walker, 1856 C. thriambus Dyar, 1921

5. Culiseta

C. impatiens (Walker, 1848)

C. incidens Thompson, 1868

C. inornata Williston, 1893

C. morsitans dyari Coquillett, 1902

C. silvestris minnesotae Barr, 1957

6. Orthopodomyia

O. signifera (Coquillett, 1896)

7. Psorophora

P. signipennis (Coquillett, 1896)

E. Cuterebridae

Cuterebra

C. angustifrons Dalmat, 1942

C. approximata Walker, 1866

C. grisea Coquillett, 1904

C. jellisoni Curran, 1942

C. lepusculi Townsend, 1897

C. polita Coquillett, 1898

C. princeps (Austen, 1895) C. ruficrus (Austen, 1933)

C. tenebrosa Coquillett, 1898

F. Gasterophilidae

Gasterophilus

G. haemorrhoidalis (Linnaeus, 1785)

G. intestinalis (De Geer, 1776)

G. nasalis (Linnaeus, 1758)

G. Hippoboscidae

1. Icosta

I. americana (Leach, 1817)

I. hirsuta (Ferris, 1927)

I. nigra (Perty, 1833)

2. Lipoptena

L. depressa (Sav. 1823)

3. Melophagus

M. ovinus (Linnaeus, 1785)

4. Myophthiria

M. fimbriata (Waterhouse, 1887)

5. Neolipoptena

N. ferrisi (Bequaert, 1935)

6. Olfersia

O. sordida Bigot, 1885

7. Ornithomya

O. anchincuria Speiser, 1905

8. Ornithoica

O. vicina (Walker, 1849)

H. Museidae

1. Fannia

F. canicularis (Linnaeus, 1761)

F. scalaris (Fabricius, 1794)

2. Haematobia

H. irritans Linnaeus, 4785

3. Musca

M. autumnalis De Geer, 1776

M. domestica (Linnaeus, 1758)

4. Muscina

M. assimilis (Fallén, 1823)

M. stabulans (Fallén, 1817)

5. Stomoxys

S. calcitrans (Linnaeus, 1758)

1. Nyeteribiidae

Basilia

B. antrozoi (Townsend, 1893)

B. corynorhini (Ferris, 1916)

B. forcipata Ferris, 1924

J. Oestridae

1. Cephenemuja

C. jellisoni Townsend, 1941

C. pratti Hunter, 1916

2. Hypoderma

H. bovis (Linnaeus, 1758)

H. lineatum (Villers, 1789)

3. Oestrus

O. ovis Linnaeus, 1758

K. Piophilidae

Piophila

P. casci (Linnaeus, 1758)

L. Psychodidae

1. Lutzomyia

L. aquilonia (Fairchild and Harwood, 1961)

L. californica (Fairchild and Hertig, 1957)

L. oppidana (Dampf, 1944)

L. stewarti (Mangabeira and Galindo, 1944)

L. vexator (Coquillett, 1907)

2. Psychoda

P. alternata Say, 1824

M. Rhagionidae

Symphoromyia

S. atripes Bigot, 1887

S. fulvipes Bigot, 1887

S. hirta Johnson, 1897

S. inquisitor Aldrich, 1915

S. jolinsoni Coquillett, 1894

S. pachyceras Williston, 1886

N. Sareophagidae

1. Ravinia

R. aeerba (Walker, 1849)

R. derelieta (Walker, 1852)

R. crrabunda (Wułp, 1895)

R. latisetosa Parker, 1914

R. lherminieri (Robineau-Desvoidy, 1830)

R. planifrons (Aldrich, 1916)

R. pusiola (Wulp, 1895)

2. Sarcophaga

S. argurostoma (Robineau-Desvoidy, 1830)

S. bishoppi Aldrich, 1916

S. bullata Parker, 1916

S. cooleyi Parker, 1914

S. haemorrhoidalis (Fallén, 1817)

S. perspicax, Aldrich, 1916

S. sarracenioides Aldrich, 1916

S. shermani Parker, 1923

S. sinuata Meigen, 1826

S. utilis Aldrich, 1915

3. Wohlfahrtia

W. vigil opaca Coquillett, 1897

O. Simuliidae

1. Cnephia

C. jcanae DeFoliart and Peterson, 1960

C. mutata (Malloch, 1914)

C. villosa DeFoliart and Peterson, 1960

2. Prosimulium

P. davicsi Peterson and DeFoliart, 1960

P. exigens Dyar and Shannon, 1927

P. flaviantennum (Stains and Knowlton, 1940)

P. fulvum (Coquillett, 1902)

P. longilobum Peterson and DeFoliart,

P. onychodactylum Dyar and Shannon, 1927

P. shcwelli Peterson and DeFoliart, 1960

P. travisi Stone, 1952

P. uinta Peterson and DeFoliart, 1960

P. unicum (Twinn, 1938)

3. Simulium

S. arcticum Malloch, 1914

S. argus Williston, 1893

S. aureum Fries, 1824

S. bicorne Dorogostajskij, Rubtzov, and Vlasenko, 1935 S. *bivittatum* Malloch, 1914

S. canadense Hearle, 1932

S. canonicola (Dyar and Shannon, 1927)

S. corbis Twinn, 1936

S. decorum Walker, 1848

S. defoliarti Stone and Peterson, 1958

S. griscum Coquillett, 1898

S. hunteri Malloch, 1914

S. jaeumbae Dyar and Shannon, 1927 S. latipes (Meigen, 1804)

S. mediovittatum Knab, 1915

S. meridionale Riley, 1887

S. nigricoxum Stone, 1952 S. petersoni Stone and DeFoliart, 1959

S. piperi Dyar and Shannon, 1927

S. pugetense (Dyar and Shannon, 1927)

S. rugglesi Nicholson and Mickel, 1950

S. trivittatum Malloch, 1914

S. tuberosum (Lundstrom, 1911)

S. venator Dyar and Shannon, 1927

S. venustum Say, 1823

S. virgatum Coquillett, 1902

S. vittatum Zetterstedt, 1838

S. wyomingense Stone and DeFoliart, 1959

T. nova (Dyar and Shannon, 1927)

P. Streblidae

Trichobius

T. corynorhini Cockerell, 1910

T. major Coquillett, 1899

Q. Syrphidae

Eristalis

E. dimidiatas Wiedemann, 1830

E. tenax (Linnaeus, 1758)

R. Tabanidae

1. Atylotus

A. incisuralis var. utahensis Rowe and Knowlton, 1935

2. Chrysops

C. aestuans Wulp, 1867

C. callidus Osten Sacken, 1875

C. carbonarius Walker, 1848

C. coquilletti Hine, 1904

C. discalis Williston, 1880

C. excitans Walker, 1850

C. frigidus Osten Sacken, 1877

fulvaster Osten Sacken, 1877

furcatus Walker, 1848

C. indus Osten Sacken, 1875

C. mitis Osten Saeken, 1875

C. niger Macquart, 1838

C. noctifer noctifer Osten Sacken, 1877

C. noctifer pertinax Williston, 1887

C. pachycerus Williston, 1887

C. sackenii Hine, 1903

C. wileyae Philip, 1955

3. Haematopota

H. americana Osten Sacken, 1875

4. Hybomitra

H. epistates (Osten Sacken, 1878)

H. frontalis (Walker, 1848)

II. opaca (Coquillett, 1904)

H. rhombica (Osten Sacken, 1876)

H. rhombica var. osburni (Hine, 1904)

H. rupestris (McDonnough, 1921)

II. sequax (Williston, 1887)

H. sonomensis var. phaenops (Osten Sacken,

II. tetrica var. hirtula (Bigot, 1892)

5. Pilimas

P. californicus (Bigot, 1892)

6. Silvius

S. quadrivittatus (Say, 1823)

7. Stenotabanus

S. flavidus (Hine, 1904)

S. guttatulus (Townsend, 1893)

8. Tabanus

T. aegrotus Osten Sacken, 1877

T. atratus Fabrieius, 1775

T. dorsifer Walker, 1860

T. gilanus Townsend, 1897 T. laticeps Hine, 1904

T. lincola Fabricius, 1794

T. productus Hine, 1904

T. pumilus Maequart, 1838

T. punctifer Osten Sacken, 1876

T. stonei Philip, 1941

Hemiptera

A. Cimicidae

C. lectularius Linnaeus, 1758

C. pilosellus (Horvath, 1910)

2. Occiacus

O. vicarius Horvath, 1912

B. Reduviidae

I. Reduvius

R. personatus (Linnaeus, 1758)

R. vanduzeci Wygodzinsky and Usinger,

2. Triatoma

T. protracta (Uhler, 1894)

Hymenoptera

A. Apidae

1. Apis

A. mellifera Linnaeus, 1758

2. Bombus

B. appositus Cresson, 1878

B. bifarius Cresson, 1878

B. centralis Cresson, 1864 B. edwardsii Cresson, 1878

B. flavifrons Cresson, 1863

B. griscocollis (De Geer, 1773) B. hunti Greene, 1860

B. morrisoni Cresson, 1878

B. nevadensis nevadensis Cresson, 1874

B. occidentalis occidentalis Greene, 1858

B. rufocinctus Cresson, 1863

B. Formicidae

I. Pogonomyrmex

P. barbatus barbatus (F. Smith, 1856)

P. californicus (Buckley, 1867)

P. imberbiculus W.M. Wheeler, 1902

P. occidentalis (Cresson, 1865)

P. rugosus Emery, 1895

2. Crematogaster

C. mormonum Emery, 1895

C. vermiculata Emery, 1895

3. Solenopsis

S. molesta molesta (Say, 1836)

S. molesta validiuscula Emery, 1895

S. salina W.M. Wheeler, 1908

S. xyloni McCook, 1879

C. Mutillidae

1. Chyphotes

C. epedaphus Buzicky, 1941

C. similis Baker, 1905

2. Dasymutilla

D. californica (Radoszkowski, 1861)

D. canco (Blake, 1879)

D. fulvohirta (Cresson, 1865)

D. gloriosa (Saussure, 1867)

D. klugii (Gary, 1872)

D. monticola (Cresson, 1865)

D. phaon phaon (Fox, 1899)

D. phaon var. fimbrialis Mickel, 1928

D. scitula Miekel, 1928

D. ursula (Cresson, 1875)

D. vesta vesta Cresson, 1865

3. Dilophotopsis

D. concolor concolor (Cresson, 1865)

4. Odontophotopsis

O. erchus (Melander, 1903)

O. inconspicua (Blake, 1886)

O. melicausa (Blake, 1871)

O. venusta (Blake, 1886)

5. Pseudomethoca

P. contumax (Cresson, 1865)

P. contumeliosa Mickel, 1935

P. manca Mickel, 1924

P. propinqua (Cresson, 1865)

P. toumcyi (Fox, 1894)

6. Sphaeropthalma

S. abdominalis (Blaker, 1886)

S. ceres (Fox, 1899)

S. dirce (Fox, 1899)

S. marpesia (Blake, 1879) S. unicolo (Cresson, 1865)

7. Timulla

T. grotei (Blake, 1871)

8. Typhoctes

T. peculiaris (Cresson, 1875)

D. Pompilidae

Pepsis

P. angustimarginata Viereck, 1908

P. mildei Stol, 1857

P. pallidolimbata pallidolimbata Lucas, 1895

P. thisbe Lucas, 1895

E. Sphecidae

1. Astata

A. bicolor Say, 1823

A. nevadica Cresson, 1881

A. nubecula Cresson, 1865

A. occidentalis Cresson, 1881

2. Bembix

B. americana comata Parker, 1917

B. americana spinolae Lepeletier, 1845

B. amoena Handlirsch, 1893

B. occidentalis W.J. Fox, 1893

B. rugosa Parker, 1917

3. Cerceris

C. conifrons Mickel, 1916

C. convergeus Viereck and Cockerell, 1904

C. finitima Cresson, 1865

C. nigrescens Smith, 1856

4. Chypeadon

C. laticinetus (Cresson, 1865)

Didineis

D. nodosa Fox, 1894

6. Mimesa

M. cressonii Packard, 1867

7. Philanthus

P. gibbosus (Fabricius, 1775)

8. Prionyz

P. atratus (Lepeletier, 1845)

P. parkeri Bohart and Menke, 1963

F. Vespidae

1. Ancistrocerus

A. antilope antilope (Panzer, 1798)

A. catskill albophaleratus (Saussure, 1855)

A. catskill eatskill (Saussure, 1853)

A. lineativentris fulvicarpus Cameron, 1908

A. neocallosus neocallosus Bequaert, 1943

A. spilogaster Cameron, 1905

A. tigris tigris (Saussure, 1857)

A. tuberculiceps sutterianus (Saussure, 1875)

A. tuberculiceps tuberculiceps (Saussure,

2. Enodynerus

E. annulatus annulatus (Say, 1824)

E. annulatus sulphureus (Saussure, 1858)

E. auranus (Cameron, 1906)

E. boscii boscii (Lepeletier, 1841) E. exoglyphus alborittatus (R. Bohart, 1939)

E. exoglyphus exoglyphus (R. Bohart, 1939) E. foraminatus acqualus (Cameron, 1906) E. fusus fusus (Cresson, 1872)

E. hidalgo hidalgo (Saussure, 1857)

E. martini (R. Bohart, 1942)

E. pratensis pratensis (Saussure, 1870)

E. russatus (R. Bohart, 1942)

3. Eumenes

E. bollii bollii Cresson, 1872

E. iturbide pedalis Fox, 1894

E. sculleni R. Bohart, 1950

E. verticalis tricinctus Isely, 1917

4. Leptochilus

L. erubescens (R. Bohart, 1940)

L. republicanus (Dalla Torre, 1889)

L. rubicundulus (R. Bohart, 1940)

L. rufinodus (Cresson, 1868)

5. Mischocyttarus

M. flavitarsis flavitarsis (Saussure, 1854)

M. flavitarsis idahoensis Bequaert, 1933

6. Odynerus

O. cinnabarinus R. Bohart, 1939

O. margaretellus Rohwer, 1915

P. canadensis var. kaibabensis Hayward,

P. flavus Cresson, 1868

P. fuscatus centralis 11ayward, 1933

P. fuscatus utahensis Hayward, 1933

8. Pscudomasaris

P. edwardsii (Cresson, 1872)

P. zonalis (Cresson, 1864)

9. Pterocheilus

P. laticeps Cresson, 1872

P. micheneri R. Bohart, 1940

P. pedicellatus R. Bohart, 1940

P. provancheri (Huard, 1895)

10. Stenodynerus

S. apache R. Bohart, 1949

S. blandoides blandoides R. Bohart, 1943

S. blandus blandus (Saussure, 1870)

S. cochisensis (Viereck, 1908)

S. minimoferus R. Bohart, 1949

S. noticeps noticeps R. Bohart, 1948

S. percampanulatus (Viereck, 1906)

S. toltecus (Saussure, 1857)

S. valliceps R. Bohart, 1948

Symmorphus

S. meridionalis (Viereck, 1903)

12. Vespula

V. artica Rohwer, 1916

V. arenaria (Fabricius, 1775)

V. atropilosa (Sladen, 1918)

V. austriaca (Panzer, 1799)

V. consobrina (Saussure, 1864)

V. maculata (Linnaeus, 1763)

V. norvegicoides Sladen, 1918

V. pennsylvanica (Saussure, 1857)

V. vulgaris (Linnaeus, 1758)

VII. Lepidoptera

A. Arctiidae

1. Arachuis

A. picta Packard, 1864

A. caja utahensis (Henry Edwards, 1886)

3. Apantesis

A. nevadensis (Grote and Robinson, 1866)

A. ornata (Packard, 1864)

A. parthenice (Kirby, 1837)

A. proxima (Guerin-Meneville, 1844)

A. williamsi tooele Barnes and McDunnough, 1910

A. williamsi form determinata (Neumoegen, 1881)

4. Diacrisia

D. vagans (Boisduval, 1852) D. virginica (Fabricius, 1798)

5. Ectypia

E. clio jessica (Barnes, 1900)

6. Estigmene

E. oregonsis (Stretch, 1873)

7. Halysidota

II. argentata subalpina French, 1890

II. maculata agassizi Packard, 1864

H. osluri Rothschild, 1909

H. tesellaris (J. E. Smith and Abbot, 1797)

8. Hemiliyalea

H. labecula (Grote, 1881)

9. Holomelina

H. fragilis (Strecker, 1878)

10. Isia

I. isabella (J. E. Smith and Abbott, 1797)

11. Leptarctia

L. californiae form decia (Boisduval, 1869)

12. Nemcophila

N. plantaginis (Linnaeus, 1758)

B. Lasiocampidae

1. Malacosoma

M. americanum (Fabricius, 1793)

M. californicum fragile (Stretch, 1881)

M. disstria Hubner, 1822

2. Phyllodesma

P. americana (Harris, 1841)

3. Tolype

T. glenwoodi Barnes, 1900

C. Lymantridae

Dasychira

D. vagans grisca (Barnes and McDunnough, 1913)

D. Nymphalidae

1. Aglsis

A. milberti Godart, 1819

2. Argunnis

A. leto Behr, 1862

A. nokomis Edwards, 1862

3. Basilarchia

B. lorguini Boisduval, 1852

4. Nymphalis

 \hat{N} . antiopa (Linnaeus, 1758)

5. Vanessa

V. atalanta Linnaeus, 1758 V. cardui Linnaens, 1758

V. carne Hubner, 1806

E. Saturniidae

I. Automeris

A. io (Fabricius, 1775)

2. Coloradia

C. pandora Blake, 1863

3. Hemileuca

II. eleganterina (Boisduval, 1852)

II. hera hera (Harris, 1841)

II. nevadensis Stretch, 1872

II. oliviae Cockerell, 1898

4. Platysamia

P. euryalus (Boisduval, 1855)

P. gloveri Strecker, 1872

VIII. Mallophaga

A. Gyropidae

Gliricola

G. porcelli (Schank, 1781)

G. ovalis Burmeister, 1838

B. Laemobothriidae

Laemobothrion

L. atrum (Nitzsch, 1818)

L. glutinans Nitzsch. 1861

L. maximum (Scopoli, 1763)

L. simile Kellogg, 1896

L. tinnunculi (Linnaeus, 1758)

L. vulturis (J. C. Fabricius, 1775)

C. Menoponidae

1. Actornithophilus

A. lacustris Clay, 1962

A. limarius Clay, 1962

A. luminosae (Kellogg, 1908)

A. mexicanus Emerson, 1953

A. ochraceus (Nitzsch, 1818)

A. paludosus Clay, 1962

A. patellatus (Piaget, 1890)

A. piceus lari (Packard, 1870)

A. piccus piccus (Denny, 1842)

A. stictus (Kellogg & Paine, 1911) A. totani (Schrank, 1803)

A. umbrinus (Burmeister, 1838)

A. uniseriatum (Piaget, 1880)

2. Amyrsidea

A. megalosoma (Overgaard, 1943)

A. perdicis (Denny, 1842)

3. Ardeiphilus

A. floridae Tuff, 1965

4. Austromenopon

A. aegialitidis (Durrant, 1906)

A. atrofulcum (Piaget, 1880)

A. durisetosum (Blagoveshtchensky, 1948)

A. himantopi Timmermann, 1954 A. limosae Timmermann, 1954 A. micrandum (Nitzsch, 1866)

A. sachtlebeni Timmermann, 1954

A. spenceri Timmermann, 1956

A. squatarolac Timmermann, 1954

A. transversum (Denny, 1842)

5. Bonomiella

B. columbae Emerson, 1957

6. Ciconiphilus

C. butoridiphagus Carriker, 1964

C. cygni Price & Beer, 1965

C. decimfasciatus (Boisduval & Lacordaire, 1835)

C. pectiniventris (Harrison, 1916)

7. Colpocephalum

C. brachysomum Kellogg & Chapman, 1902

C. flavescens (de 11aan, 1829) C. fregili Denny, 1842 C. impressum Rudow, 1866 C. kelloggi Osborn, 1902 C. leptopygos Nitzsch, 1874 C. nanum Piaget, 1890 C. napiforme Rudow, 1869 C. pectinatum Osborn, 1902 C. tausi (Ansari, 1951) C. turbinatum Denny, 1842 C. unciferum Kellogg, 1896

8. Comatomenopon C. thulae Tuff, 1967

9. Cuculiphilus C. alternatus (Osborn, 1902)

C. zerafae Ansari, 1955

10. Dennyus D. Bruneri (Carriker, 1903) D. spiniger Ewing, 1930

11. Eureum E. spenceri Emerson & Pratt, 1956

12. Hohorsticlla H. frontalis Carriker, 1949

13. Holomenopopn

H. clypeilargum Eichler, 1943 H. leucoxanthum (Burmeister, 1838) H. setigerum (Blagoveshtchensky, 1948) H. transvaalense (Bedford, 1920)

14. Kurodaia K. acadicac Price & Beer, 1963 K. flammei Price & Beer, 1963 K. fulvovasciata (Piaget, 1880) K. haliacti (Denny, 1842)

K. magna Emerson, 1960 K. painei (McGregor, 1912) K. subpachygaster (Piaget, 1880)

15. Machaerilaemus

M. amercanus (Ewing, 1930) M. clayac (Balat, 1966) M. malleus (Burmeister, 1838) M. mclospizac Emerson, 1954

16. Menacanthus

M. alaskensis (Kellogg & Chapman, 1902) M. annulatus (Giebel, 1874)

M. chrysophaeus (Kellogg, 1896)

M. distinctus (Kellogg & Chapman, 1899) M. eurysternum (Burmeister, 1838)

M. expansus (Osborn, 1896) M. gonophacus (Burmeister, 1838) M. mutabilis Blagoveshtchensky, 1940 M. perforatus (Piaget, 1880)

M. persignatus (Kellogg & Chapman, 1899) M. picicola (Packard, 1873)

M. robustus (Kellogg, 1896) M. stramineus (Nitzsch, 1818)

17. Menopon M. pallens Clay, 1949

18. Myrsidea

M. anaspila (Nitzsch, 1866)

M. conspica (Kellogg & Chapman, 1902)

M. culcullaris (Nitzsch, 1818) M. dissimilis (Kellogg, 1896) M. emersoni Clay, 1966 M. ineerta (Kellogg, 1896)

M. interrupta (Osborn, 1896) M. latifrons (Carriker, 1910)

M. melanorum (Kellogg, 1896) M. palloris (Carriker, 1903) M. picae (Linnaeus, 1758) M. quadrifasciata (Piaget, 1880) M. quadrimaculata (Carriker, 1902) M. ridulosa (Kellogg & Chapman, 1899) M. rustica (Giebel, 1874)

19. Nosopon N. lucidum (Rudow 1869)

20. Piagetiella P. peralis (Leidy, 1878)

21. Plegadiphilus P. plegadis (Dubinin, 1938)

22. Pseudomenopon P. insolens' (Kellogg, 1896) P. par (Kellogg, 1896) P. pilosum (Scopoli, 1763) P. quadrii Eichler, 1952 23. Triniton

T. anserinum (J. C. Fabricius, 1805) T. querquedulae (Linnaeus, 1758)

D. Philopteridae 1. Acidoproctus

A. maximus Piaget, 1878

2. Anaticola A. crassicornis cornicephalus (Zavaleta,

A. crassicornis crassicornis (Scopoli, 1763) A. crassicornis dafilensis Carriker, 1956 A. crassicornis depuratus (Nitzsch, 1866)

A. crassicornis hopkinsi Eichler, 1954 A. crassicornis mergiserrati (De Geer, 1778)

3. Anatoecus

A. cygni cmersoni Keler, 1960 A. dentatus affinis Keler, 1960 A. dentatus dentatus (Scopoli, 1763) A. dentatus ferrugineus (Giebel, 1874) A. icterodes bipunctatus (Giebel, 1874) A. icterodes boschadis Keler, 1960 A. icterodes icterodes (Nitzsch, 1818) A. icterodes marcui Keler, 1960 A. icterodes simmillimus Keler, 1960 A. icterodes tendeiroi Keler, 1960

4. Aquanirmus A. amercanus (Kellogg & Chapman, 1899)

5. Ardeicola A. botauri (Osborn, 1896)

A. cruscula Carriker, 1960 A. expallida Blagoveschtchensky, 1940

A. florida nigra Tuff, 1967 A. goisagi Uchida, 1953

A. rhaphidus (Nitzsch, 1866)

6. Bruelia

B. angustifrons (Carriker, 1902) B. argula (Burmeister, 1838) B. audax (Kellogg, 1899) B. biocellata (Piaget, 1880) B. brachythorax (Giebel, 1874) B. cedrorum (Piaget, 1880) B. deficiens (Piaget, 1885)

B. domestica (Kellogg & Chapman, 1899) B. ductilis (Kellogg & Chapman, 1899) B. iliaci brevicolor Ansari, 1956

B. interposita (Kellogg, 1899) B. limbata (Burmeister, 1838) B. longa (Kellogg, 1896)

B. longifrons Carriker, 1956 B. nebulosa (Burmeister, 1838) B. ornatissima (Giebel, 1874) B. peninsularis (Kellogg, 1899) B. rotundata (Osborn, 1896) B. straminea (Denny, 1842)

B. subtilis (Nitzseh, 1874)
B. tenuis (Burmeister, 1838)
B. xanthocephali (Osborn, 1896)
B. zeropunctata antiqua Ansari, 1956

B. zeropunctata zeropunctata Ansari, 1957

7. Carduiceps

C. cingulatus cingulatus (Denny, 1842) C. cingulatus clayae Timmermann, 1954 C. zonarius (Nitzsch, 1866)

8. Chelopistes
C. meleagridis (Linnaeus, 1758)

9. Cirrophthirus

C. tcstudinarius (Children, 1836)
10. Colinicola

C. docophoroides (Piaget, 1880)

11. Columbicola
C. baculoides

C. baculoides Paine, 1912C. macrourae (Wilson, 1941)

12. Craspedorrhynchus

C. americanus Emerson, 1960
C. aquilinus (Denny, 1842)
C. dilatatus (Rudow, 1869)
C. haematopus (Scopoli, 1763)
C. hirsutus Carriker, 1956
C. subhaematopus Emerson, 1960

Cuclogaster

 C. heterogrammicus (Nitzsch, 1866)

14. Cuculicola C. splendidus (Kellogg, 1899)

15. Cuculoecus C. coccygi (Osborn, 1895)

Cummingsiclla
 C. ambigua (Burmeister, 1838)
 C. longirostricola (Wilson, 1937)

C. longirostricola (Wil 17. Degecrriclla

D. discocephalus aquilarum Eichler, 1943 D. fulva (Giebel, 1874) D. fusca (Denny, 1842) D. nisus nisus (Giebel, 1866) D. nisus vagans (Giebel, 1874)

D. regalis (Giebel, 1866) D. rufa carruthi Emerson, 1953 D. rufa rufa (Burmeister, 1838)

Falcolipeurus

 F. marginalis Osborn, 1902
 F. suturalis (Rudow, 1869)

F. suturalis (Rudow, 1869)

19. Fulicoffula
F. americana Emerson, 1960

F. comstocki (Kellogg & Paine, 1911) F. distincta Emerson, 1960

F. longipila (Kellogg, 1896)

20. Goniocotes
G. chrysocephalus Giebel, 1874
G. microthorax (Stephans, 1829

G. chrysocephatus Glebel, 1874
G. microthorax (Stephans, 1829)
21. Goniodes

G. bonasus Emerson, 1948 G. centrocerci Simon, 1938 G. colchici Denny, 1842 G. dispar Burmeister, 1838
G. merriamanus Packard, 1873
G. nebraskensis Carriker, 1945
G. stefani Clay & Hopkins, 1955
G. submamillatus Emerson, 1950

22. Ibidoecus 1. bisignatus (Nitzsch, 1866)

23. Incidifrons
1. monachus (Kellogg & Paine, 1911)
1. transpositus (Kellogg, 1896)

Lagopoecus

 L. colchicus Emerson, 1949
 L. gambeli Emerson, 1949
 L. gibsoni Hopkins, 1947
 L. obscurus Emerson, 1948

L. perplexus (Kellogg & Chapman, 1899)

L. umbellus Emerson, 1950

Lipeurus
 L. maculosus Clay, 1938

Lunaceps

 L. holophaeus cabenisi Timmermann, 1954
 L. limosella clayae Timmermann, 1954
 L. numenii (Denny, 1842)

27. Mulcticola M. macrocephalus (Kellogg, 1896)

28. Ornithobius
O. gonipleurus Denny, 1842
O. waterstoni reconditus Timmermann, 1962

Oxylipeurus
O. corpulentus Clay, 1938
O. ellipticus (Keler, 1958)
O. mesopelios colchicus Clay, 1938
O. polytrapezius (Burmeister, 1838)

30. Pectinopygus
P. farallonii (Kellogg, 1896)
P. tardotti Flbel & Emerson, 1956

P. tordoffi, Elbel & Emerson, 1956
31. Perenirmus
P. arcticus Cariker, 1958
P. auritus (Scopoli, 1763)

P. gulosus (Nitzseh, 1866) P. jungens (Kellogg, 1896)

P. mirinotatus (Kellogg & Chapman, 1899)
P. quadripustulatus (Kellogg & Mann, 1912)

32. Philopterus
P. agelaii (Osborn, 1896)
P. americanus (Kellogg, 1899)
P. citrinellae curvirostrae (Sehrank, 1776)

P. corvi (Linnaeus, 1758) P. excisus domesticus (Kellogg, 1896) P. excisus major (Kellogg, 1896)

P. excisus microsomaticus Tandan, 1955 P. fringillae (Seopoli, 1772)

P. fringillae (Seopoli, 1772) P. garrulae (Piaget, 1880) P. hanzaki Balat, 1955

P. mirus (Kellogg & Chapman, 1899) P. ocellatus osborni Edwards, 1952

P. phillipi Emerson, 1953 P. picae (Denny, 1842) P. rufus (Kellogg, 1899) P. rutteri (Kellogg, 1899)

33. Physconelloides
P. spenceri Emerson & Ward, 1958
P. wisemani Emerson, 1960
P. zenaidurae (MeGregor, 1917)

34. Picicola

P. foedus (Kellogg & Chapman, 1899)

P. orpheus (Osborn, 1896)

P. snodgrassi (Kellogg, 1896)

35. Quadraceps

Q. alcyonae (Carriker, 1959)

Q. assimilis major (Kellogg, 1899)

Q. carrikeri Hopkins & Timmermann, 1954

Q. connexus (Kellogg & Mann, 1912)

Q. charadrii hospes (Nitzseh, 1866)

Q. falcigerus (Peters, 1931) Q. fimbriatus (Giebel, 1866)

Q. griseus (Rudow, 1869)

Q. hemichrous (Nitzsch, 1866)

Q. hiaticulae boephilus (Kellogg, 1896)

Q nigrolimbatus (Mjöberg, 1910) Q. phaeonotus (Nitzseh, 1866)

Q. punctatus sublingulatus Timmerman, 1952

Q. ravus (Kellogg, 1899)

Q. semifissus mexicanus Carriker, 1944

Q. similis (Giebel, 1866)

Q. zephyra (Timmermann, 1954)

36. Rallicola

R. advenus (Kellogg, 1896) R. kelloggi Emerson, 1957

R. mystax (Giehel, 1874)

R. ortygometrae subporzanae Emerson, 1957

37. Rhynonirmus

R. scolopacis (Denny, 1842)

38. Rotundiceps

R. cordatus (Osborn, 1896)

39. Saemundssonia

S. conica conica (Denny, 1842)

S. conica naumanni (Giebel, 1874)

S. kratochvili Balat, 1950

S. lari congener (Giebel, 1874)

S. lobaticeps (Giebel, 1874)

S. parvigenitalis Ward, 1955

S. platygaster nitzschi (Giebel, 1866)

S. platygaster platygaster (Denny, 1842)

S. scolopacisphaeopodis (Schrank, 1803)

S. tricolor Carriker, 1956

S. tringae (O. Fabricius, 1780)

40. Strigiphilus

S. acutifrons Emerson, 1961

S. aitkeni Clay, 1966

S. barbatus (Osborn, 1902)

S. cursor (Burmeister, 1838)

S. oculatus (Budow, 1870)

S. otus Emerson, 1955

S. speotyti (Osborn, 1896)

41. Sturnidoecus

S. simplex (Kellogg, 1896)

S. sturni (Schrank, 1776)

E. Ricinidae

1. Ricinus

R. angulatus (Kellogg, 1896)

R. arcuatus (Kellogg & Mann, 1912)

R. bombyeillae (Denny, 1842)

R. diffusus (Kellogg, 1896)

R. inexpectatus Balat, 1966

R. japonicus (Uchida, 1915)

R. medius Uchida, 1926

R. merulae (Durrant, 1906)

R. microcephalus (Kellogg, 1896)

R. picturatus (Carriker, 1902)

R. subhastatus (Durrant, 1906)

R. sucinaceus (Kellogg, 1896)

R. serratus (Durrant, 1906)

2. Trochiloecetes

T. lineatus (Osborn, 1896)

T. prominens (Kellogg & Chapman, 1899)

T. ochoterenai (Zavaleta, 1943)

F. Trichodectidae

1. Bovicola

B. bovis (Linnaeus, 1758)

B. caprae (Gurlt, 1843)

B. crassipes (Rudow, 1866)

B. equi (Denny, 1842)

B. limbatus (Gervais, 1844)

B. ouis (Schrank, 1781)

2. Eutrichophilus

E. setosus (Giebel, 1861)

3. Felicola

F. subrostrata (Burmeister, 1838)

4. Geomydoecus G. californicus (Chapman, 1897)

5. Neotrichodectes

N. osborni Keler, 1944

IX. Siphonaptera

A. Amphipsyllidae

Amphisylla

A. sibirica washingtoni Hubbard, 1954

B. Ceratophyllidae

1. Amphalius

A. necopinus (Jordan, 1925)

2. Ceratophyllus

C. affinis neglectus Smit, 1958

C. celsus celsus Jordan, 1926

C. garei Bothschild, 1902

C. niger C. Fox, 1908

C. petrochelidoni Wagner, 1936

3. Daetylopsylla

D. (Foxella) ignota apachina (C. Fox,

D. (Foxella) ignota arizonensis (Hubbard,

D. (Foxella) ignota comis Jordan, 1929

D. (Foxella) ignota ignota Baker, 1895 D. (Foxella) ignota recula (Jordan and

Rothschild, 1915)

D. (Foxella) ignota utahensis (Wagner, 1931)

D. (Foxelloides) minidoka Prince and Stark,

D. (Spicata) rara 1. Fox, 1940

4. Diamanus

D. montanus (Baker, 1895)

5. Malaraeus

M. bitterrootensis (Dunn, 1923)

M. euphorbi (Rothschild, 1905)

M. sinomus (Jordan, 1925)

M. telelinum (Rothschild, 1905)

M. vonfintelis Prince, 1959

6. Megabothris

M. abantis (Rothschild, 1905)

7. Monopsyllus

M. ciliatus kincaidi Hubbard, 1947

M. cyrturus (Jordan, 1929)

M. eumolpi americanus Hubbard, 1950

M. eumolpi cumolpi (Rothschild, 1905)

M. exilis (Jordan, 1937)

M. vison (Baker, 1904)

M. wagneri (Baker, 1904)

8. Nosopsyllus

N. fasciatus (Bosc d'Antie, 1801)

9. Opisocrostis

O. hirsutus (Baker, 1895)

O. labis (Jordan and Rothschild, 1922) O. tuberculatus cynomuris Jellison, 1939

O. tuberculatus tuberculatus (Baker, 1904)

Opisodasys

O. keeni keeni (Baker, 1896)

O. pseudarctomys (Baker, 1904)

11. Orchopeas

O. caedens caedens (Jordan, 1925)

O. howardii (Baker, 1895)

O. lcucopus (Baker, 1904)

O. ncotomae Auguston, 1943

O. nepos (Rothschild, 1905) O. sexdentatus agilis (Rothschild, 1905)

O. sexdentatus nevadensis (Jordan, 1929)

12. Oropsylla

O. idahocnsis (Baker, 1904)

T. acamantis medius Stark, 1970

T. acamantis utahensis (Wagner, 1936)

T. aridis campestris Prince, 1944

T. aridis hoffmani (Hubbard, 1949)

T. arizonensis (Baker, 1898)

T. bacchi bacchi (Rothschild, 1905)

T. baechi caducus (Jordan, 1930)

T. bacchi consimilis Stark, 1957

T. bacchi gladiolis (Jordan, 1925)

T. francisi barnesi (Stark, 1970)

T. francisi francisi (C. Fox, 1927)

T. pandorae pandorae Jellison, 1937

T. stanfordi (Wagner, 1936)

C. Hystrichopsyllidae

I. Anomiopsyllus

A. amphibolus Wagner, 1936

A. nudatus (Baker, 1898)

2. Atyphloceras

A. echis echis Jordan and Rothschild, 1915

A. multidentatus multidentatus (C. Fox, 1909)

3. Callistopsyllus

C. terinus (Rothschild, 1905)

4. Catallagia

C. decipiens Rothschild, 1915

C. newcyi Holland and Loshbaugh, 1958

5. Carteretta

C. carteri clavata Good, 1942

6. Conorhinopsylla

C. standfordi Stewart, 1930

7. Corrodopsylla

C. curvata curvata (Rothschild, 1915)

C. curvata obtusata (Wagner, 1929)

8. Ctenophthalmus

C. pseudagyrtes pseudagyrtes (Baker, 1904)

9. Delotelis

D. telogoni Rothschild, 1905

10. Epitedia

E. scapani (Wagner, 1936)

E. stanfordi Traub, 1944

E. testor (Rothschild, 1915)

E. wenmanni wenmanni (Rothschild, 1904)

11. Hystrichopsylla

II. dippici truncata Holland, 1957

11. linsdalei Holland, 1957

12. Jordanopsylla

J. allredi Traub and Tipton, 1951

13. Megarthroglossus

M. becki Tipton and Allred, 1951

M. divisus divisus (Baker, 1898)

M. procus Jordan and Rothschild, 1915

M. smiti Mendez, 1956

14. Meringis

M. dipodomys Kohls, 1938

M. hubbardi Kohls, 1938

M. jamesoni Hubbard, 1943

M. jewetti Hubbard, 1940

M. parkeri (Jordan, 1937)

15. Nearctopsylla

N. brooksi (Rothschild, 1904)

N. hyrtaci (Rothschild, 1904)

16. Neopsylla

N. inopina Rothschild, 1915

17. Phalacropsylla

P. allos Wagner, 1936

18. Rhadinopsylla

R. heiseri (McCoy. 1911)

R. sectilis goodi (Hubbard, 1941)

R. sectilis sectilis (Jordan and Rothschild, 1923)

R. fraterna (Baker, 1895)

19. Stenistomera

S. alpina (Baker, 1895)

S. hubbardi Egoscue, 1968

S. macrodactyla (Good, 1942)

D. Ischnopsyllidae

1. Myodopsylla

M. gentilis (Jordan and Rothschild, 1921)

2. Sternopsylla

S. distincta texana (C. Fox, 1914)

E. Leptosyllidae

1. Ctenophyllus

C. armatus terribilis (Rothschild, 1903)

2. Odontopsyllus

O. dentatus (Baker, 1904)

Ornithophaga

O. nearctica Holland and Loshbaugh, 1958

4. Peromyscopsylla

P. hamifer vigens (Jordan, 1937)

P. hesperomys adelpha (Rothschild, 1915)

P. hesperomys ravalliensis (Dunn, 1923)

P. sclenis (Rothschild, 1906)

F. Pulicidae

1. Cediopsylla

C. inacqualis inacqualis (Baker, 1895)

C. interrupta Jordan, 1925

- 2. Ctenocephalides C. felis felis (Bouche, 1835)
- 3. Echidnophaga E. gallinacea (Westwood, 1875)
- 4. Hoplopsyllus
 - H. (Euhoplopsyllus) glacialis affinis (Baker,
 - H. (Hoplopsyllus) anomalus (Baker, 1904)
- 5. Pulex P. irritans Linnaeus, 1758
- 6. Xenopsylla X. cheopis (Rothschild, 1903)
- G. Vermipsyllidae Chactopsulla C. stewarti Johnson, 1955

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